# DRAFT TOXICOLOGICAL PROFILE FOR COBALT

U.S. DEPARTMENT OF HEALTH AND HUMAN SERVICES
Public Health Service
Agency for Toxic Substances and Disease Registry

September 2001

## **DISCLAIMER**

The use of company or product name(s) is for identification only and does not imply endorsement by the Agency for Toxic Substances and Disease Registry.

## **UPDATE STATEMENT**

Toxicological profiles are revised and republished as necessary, but no less than once every three years. For information regarding the update status of previously released profiles, contact ATSDR at:

Agency for Toxic Substances and Disease Registry Division of Toxicology/Toxicology Information Branch 1600 Clifton Road NE, E-29 Atlanta, Georgia 30333

## **FOREWORD**

This toxicological profile is prepared in accordance with guidelines developed by the Agency for Toxic Substances and Disease Registry (ATSDR) and the Environmental Protection Agency (EPA). The original guidelines were published in the *Federal Register* on April 17, 1987. Each profile will be revised and republished as necessary.

The ATSDR toxicological profile succinctly characterizes the toxicologic and adverse health effects information for the hazardous substance described therein. Each peer-reviewed profile identifies and reviews the key literature that describes a hazardous substance's toxicologic properties. Other pertinent literature is also presented, but is described in less detail than the key studies. The profile is not intended to be an exhaustive document; however, more comprehensive sources of specialty information are referenced.

The focus of the profiles is on health and toxicologic information; therefore, each toxicological profile begins with a public health statement that describes, in nontechnical language, a substance's relevant toxicological properties. Following the public health statement is information concerning levels of significant human exposure and, where known, significant health effects. The adequacy of information to determine a substance's health effects is described in a health effects summary. Data needs that are of significance to protection of public health are identified by ATSDR and EPA.

Each profile includes the following:

- (A) The examination, summary, and interpretation of available toxicologic information and epidemiologic evaluations on a hazardous substance to ascertain the levels of significant human exposure for the substance and the associated acute, subacute, and chronic health effects;
- (B) A determination of whether adequate information on the health effects of each substance is available or in the process of development to determine levels of exposure that present a significant risk to human health of acute, subacute, and chronic health effects; and
- (C) Where appropriate, identification of toxicologic testing needed to identify the types or levels of exposure that may present significant risk of adverse health effects in humans.

The principal audiences for the toxicological profiles are health professionals at the Federal, State, and local levels; interested private sector organizations and groups; and members of the public. We plan to revise these documents in response to public comments and as additional data become available. Therefore, we encourage comments that will make the toxicological profile series of the greatest use.

Comments should be sent to:

Agency for Toxic Substances and Disease Registry Division of Toxicology 1600 Clifton Road, N.E. Mail Stop E-29 Atlanta, Georgia 30333

The toxicological profiles are developed in response to the Superfund Amendments and

### **Background Information**

The toxicological profiles are developed by ATSDR pursuant to Section 104(i) (3) and (5) of the Comprehensive Environmental Response, Compensation, and Liability Act of 1980 (CERCLA or Superfund) for hazardous substances found at Department of Energy (DOE) waste sites. CERCLA directs ATSDR to prepare toxicological profiles for hazardous substances most commonly found at facilities on the CERCLA National Priorities List (NPL) and that pose the most significant potential threat to human health, as determined by ATSDR and the EPA. ATSDR and DOE entered into a Memorandum of Understanding on November 4, 1992 which provided that ATSDR would prepare toxicological profiles for hazardous substances based upon ATSDR's or DOE's identification of need. The current ATSDR priority list of hazardous substances at DOE NPL sites was announced in the Federal Register on July 24, 1996 (61 FR 38451).

This profile reflects ATSDR's assessment of all relevant toxicologic testing and information that has been peer-reviewed. Staff of the Centers for Disease Control and Prevention and other Federal scientists have also reviewed the profile. In addition, this profile has been peer-reviewed by a nongovernmental panel and is being made available for public review. Final responsibility for the contents and views expressed in this toxicological profile resides with ATSDR.

Jeffrey P. Koplan, M.D., M.P.H.

Administrator

Agency for Toxic Substances and Disease Registry

COBALT vii

## QUICK REFERENCE FOR HEALTH CARE PROVIDERS

Toxicological Profiles are a unique compilation of toxicological information on a given hazardous substance. Each profile reflects a comprehensive and extensive evaluation, summary, and interpretation of available toxicologic and epidemiologic information on a substance. Health care providers treating patients potentially exposed to hazardous substances will find the following information helpful for fast answers to often-asked questions.

## Primary Chapters/Sections of Interest

- **Chapter 1: Public Health Statement**: The Public Health Statement can be a useful tool for educating patients about possible exposure to a hazardous substance. It explains a substance's relevant toxicologic properties in a nontechnical, question-and-answer format, and it includes a review of the general health effects observed following exposure.
- **Chapter 2: Relevance to Public Health**: The Relevance to Public Health Section evaluates, interprets, and assesses the significance of toxicity data to human health
- Chapter 3: Health Effects: Specific health effects of a given hazardous compound are reported by *route of exposure*, by *type of health effect* (death, systemic, immunologic, reproductive), and by *length of exposure* (acute, intermediate, and chronic). In addition, both human and animal studies are reported in this section.

**NOTE:** Not all health effects reported in this section are necessarily observed in the clinical setting. Please refer to the Public Health Statement to identify general health effects observed following exposure.

**Pediatrics:** Four new sections have been added to each Toxicological Profile to address child health issues:

Section 1.7 How Can (Chemical X) Affect Children?

Section 1.8 How Can Families Reduce the Risk of Exposure to (Chemical X)?

Section 3.6 Children's Susceptibility

Section 6.6 Exposures of Children

## Other Sections of Interest:

Section 3.7 Biomarkers of Exposure and Effect

Section 3.10 Methods for Reducing Toxic Effects

## ATSDR Information Center

*E-mail:* atsdric@cdc.gov *Internet:* http://www.atsdr.cdc.gov

COBALT viii

The following additional material can be ordered through the ATSDR Information Center:

Case Studies in Environmental Medicine: Taking an Exposure History—The importance of taking an exposure history and how to conduct one are described, and an example of a thorough exposure history is provided. Other case studies of interest include Reproductive and Developmental Hazards; Skin Lesions and Environmental Exposures; Cholinesterase-Inhibiting Pesticide Toxicity; and numerous chemical-specific case studies.

Managing Hazardous Materials Incidents is a three-volume set of recommendations for on-scene (prehospital) and hospital medical management of patients exposed during a hazardous materials incident. Volumes I and II are planning guides to assist first responders and hospital emergency department personnel in planning for incidents that involve hazardous materials. Volume III—Medical Management Guidelines for Acute Chemical Exposures—is a guide for health care professionals treating patients exposed to hazardous materials.

Fact Sheets (ToxFAQs) provide answers to frequently asked questions about toxic substances.

## Other Agencies and Organizations

The National Center for Environmental Health (NCEH) focuses on preventing or controlling disease, injury, and disability related to the interactions between people and their environment outside the workplace. Contact: NCEH, Mailstop F-29, 4770 Buford Highway, NE, Atlanta, GA 30341-3724 • Phone: 770-488-7000 • FAX: 770-488-7015.

The National Institute for Occupational Safety and Health (NIOSH) conducts research on occupational diseases and injuries, responds to requests for assistance by investigating problems of health and safety in the workplace, recommends standards to the Occupational Safety and Health Administration (OSHA) and the Mine Safety and Health Administration (MSHA), and trains professionals in occupational safety and health. Contact: NIOSH, 200 Independence Avenue, SW, Washington, DC 20201 • Phone: 800-356-4674 or NIOSH Technical Information Branch, Robert A. Taft Laboratory, Mailstop C-19, 4676 Columbia Parkway, Cincinnati, OH 45226-1998 • Phone: 800-35-NIOSH.

The National Institute of Environmental Health Sciences (NIEHS) is the principal federal agency for biomedical research on the effects of chemical, physical, and biologic environmental agents on human health and well-being. Contact: NIEHS, PO Box 12233, 104 T.W. Alexander Drive, Research Triangle Park, NC 27709 • Phone: 919-541-3212.

#### Referrals

The Association of Occupational and Environmental Clinics (AOEC) has developed a network of clinics in the United States to provide expertise in occupational and environmental issues. Contact:

AOEC, 1010 Vermont Avenue, NW, #513, Washington, DC 20005 • Phone: 202-347-4976 •
FAX: 202-347-4950 • e-mail: AOEC@AOEC.ORG • Web Page: http://www.aoec.org/.

COBALT ix

The American College of Occupational and Environmental Medicine (ACOEM) is an association of physicians and other health care providers specializing in the field of occupational and environmental medicine. Contact: ACOEM, 55 West Seegers Road, Arlington Heights, IL 60005 • Phone: 847-818-1800 • FAX: 847-818-9266.

COBALT xi

## **CONTRIBUTORS**

## CHEMICAL MANAGER(S)/AUTHORS(S):

Obaid M. Faroon, D.V.M., Ph.D. Henry Abadin, M.S.P.H. Sam Keith, M.S., C.H.P. ATSDR, Division of Toxicology, Atlanta, GA

Mark Osier, Ph.D.
Gary Diamond, Ph.D.
Gloria Sage, Ph.D.
Syracuse Research Corporation, North Syracuse, NY

#### THE PROFILE HAS UNDERGONE THE FOLLOWING ATSDR INTERNAL REVIEWS:

- 1. Health Effects Review. The Health Effects Review Committee examines the health effects chapter of each profile for consistency and accuracy in interpreting health effects and classifying end points.
- 2. Minimal Risk Level Review. The Minimal Risk Level Workgroup considers issues relevant to substance-specific minimal risk levels (MRLs), reviews the health effects database of each profile, and makes recommendations for derivation of MRLs.
- 3. Data Needs Review. The Research Implementation Branch reviews data needs sections to assure consistency across profiles and adherence to instructions in the Guidance.

COBALT xiii

#### PEER REVIEW

A peer review panel was assembled for cobalt. The panel consisted of the following members:

- 1. Dr. Herman Cember, C.H.P., Ph.D., PE., Adjunct Professor, School of Health Sciences, Purdue University, Lafayette, Indiana;
- 2. Dr. James Hansen, Ph.D., Senior Associate, Stratus Consulting Inc., Longmont, CO;
- 3. Dr. Dominique Lison, M.D., Ph.D., Vice-Chairman of the Doctoral School in Genetics and Immunology, Catholic University of Louvain, Brussels, Belgium;
- 4. Dr. Nancy Pedigo, Ph.D., Research Assistant Professor, Department of Pharmacology, University of Kentucky Medical Center, Lexington, KY.

These experts collectively have knowledge of cobalt's physical and chemical properties, toxicokinetics, key health end points, mechanisms of action, human and animal exposure, and quantification of risk to humans. All reviewers were selected in conformity with the conditions for peer review specified in Section 104(I)(13) of the Comprehensive Environmental Response, Compensation, and Liability Act, as amended.

Scientists from the Agency for Toxic Substances and Disease Registry (ATSDR) have reviewed the peer reviewers' comments and determined which comments will be included in the profile. A listing of the peer reviewers' comments not incorporated in the profile, with a brief explanation of the rationale for their exclusion, exists as part of the administrative record for this compound. A list of databases reviewed and a list of unpublished documents cited are also included in the administrative record.

The citation of the peer review panel should not be understood to imply its approval of the profile's final content. The responsibility for the content of this profile lies with the ATSDR.

COBALT xv

## **CONTENTS**

FOREWORD v
QUICK REFERENCE FOR HEALTH CARE PROVIDERS
CONTRIBUTORS
PEER REVIEW xiii
LIST OF FIGURES xix
LIST OF TABLES xxi
1. PUBLIC HEALTH STATEMENT
2. RELEVANCE TO PUBLIC HEALTH
2.1 BACKGROUND AND ENVIRONMENTAL EXPOSURES TO COBALT IN THE UNITED STATES
2.3 MINIMAL RISK LEVELS
3. HEALTH EFFECTS       27         3.1 INTRODUCTION       27         3.2 DISCUSSION OF HEALTH EFFECTS BY ROUTE OF EXPOSURE       27         3.2.1 Inhalation Exposure       29         3.2.1.1 Death       29         3.2.1.2 Systemic Effects       41         3.2.1.3 Immunological and Lymphoreticular Effects       49         3.2.1.4 Neurological Effects       49         3.2.1.5 Reproductive Effects       50         3.2.1.6 Developmental Effects       50         3.2.1.7 Cancer       50
3.2.2       Oral Exposure       52         3.2.2.1       Death       52         3.2.2.2       Systemic Effects       68         3.2.2.3       Immunological and Lymphoreticular Effects       75         3.2.2.4       Neurological Effects       76

		3.2.2.5	Reproductive Effects	. 76
		3.2.2.6	Developmental Effects	. 77
		3.2.2.7	Cancer	
	3.2.3	Dermal E	Exposure	
		3.2.3.1	Death	
		3.2.3.2	Systemic Effects	
		3.2.3.3	Immunological and Lymphoreticular Effects	
		3.2.3.4	Neurological Effects	
		3.2.3.5	Reproductive Effects	
		3.2.3.6	Developmental Effects	
		3.2.3.7	Cancer	
	3.2.4		Exposure	
	5.2.1	3.2.4.1	Death	
		3.2.4.2	Systemic Effects	
		3.2.4.3	Immunological and Lymphoreticular Effects	
		3.2.4.3	Neurological Effects	
		3.2.4.4		
			Reproductive Effects	
		3.2.4.6	Developmental Effects	
2.2	CENTOR	3.2.4.7	Cancer	
3.3		OXICITY		
3.4			CS	
	3.4.1		on	
		3.4.1.1	Inhalation Exposure	
		3.4.1.2	Oral Exposure	
		3.4.1.3	Dermal Exposure	
	3.4.2	Distribut		
		3.4.2.1	Inhalation Exposure	
		3.4.2.2	Oral Exposure	119
		3.4.2.3	Dermal Exposure	120
		3.4.2.4	Other Routes of Exposure	120
	3.4.3	Metaboli	sm	121
	3.4.4	Eliminati	ion	121
		3.4.4.1	Inhalation Exposure	121
		3.4.4.2	Oral Exposure	122
		3.4.4.3	Dermal Exposure	
		3.4.4.4	Other Routes of Exposure	
	3.4.5	Physiolo	gically Based Pharmacokinetic (PBPK)/Pharmacodynamic (PD)	
		Models		127
3.5	MECHA	ANISMS (	OF ACTION	142
	3.5.1	Pharmac	okinetic Mechanisms	142
	3.5.2		sms of Toxicity	
	3.5.3		o-Human Extrapolations	147
3.6			SRUPTION	148
3.7			JSCEPTIBILITY	150
3.8			OF EXPOSURE AND EFFECT	153
٥.٥	3.8.1		ers Used to Identify or Quantify Exposure to Cobalt	
	3.8.2		ers Used to Characterize Effects Caused by Cobalt	
3.9			WITH OTHER CHEMICALS	
			THAT ARE UNUSUALLY SUSCEPTIBLE	
3.11				
	3.11.1	Reducing	g Peak Absorption Following Exposure	100

		3.11.2 Reducing Body Burden	160
		3.11.3 Interfering with the Mechanism of Action for Toxic Effects	161
	3.12	ADEQUACY OF THE DATABASE	161
		3.12.1 Existing Information on Health Effects of Cobalt	161
		3.12.2 Identification of Data Needs	164
		3.12.3 Ongoing Studies	173
4	CHEN	AUCAL AND DIVICIOAL INFORMATION	175
4.		MICAL AND PHYSICAL INFORMATION	
	4.1 4.2	CHEMICAL IDENTITY	175
	4.2	PHYSICAL, CHEMICAL, AND RADIOLOGICAL PROPERTIES	1/3
5.	PROI	DUCTION, IMPORT/EXPORT, USE, AND DISPOSAL	185
	5.1	PRODUCTION	185
	5.2	IMPORT/EXPORT	191
	5.3	USE	191
	5.4	DISPOSAL	193
6.			197
	6.1	OVERVIEW	
	6.2	RELEASES TO THE ENVIRONMENT	202
	6.3	6.2.3 Soil	
	0.3	6.3.1 Transport and Partitioning	
		6.3.2 Transformation and Degradation	
		6.3.2.1 Air	
		6.3.2.2 Water	
		6.3.2.3 Sediment and Soil	
	6.4	LEVELS MONITORED OR ESTIMATED IN THE ENVIRONMENT	
	0.1	6.4.1 Air	
		6.4.2 Water	
		6.4.3 Sediment and Soil	
		6.4.4 Other Environmental Media	
	6.5	GENERAL POPULATION AND OCCUPATIONAL EXPOSURE	
	6.6	EXPOSURES OF CHILDREN	244
	6.7		245
	6.8	ADEQUACY OF THE DATABASE	246
		6.8.1 Identification of Data Needs	247
		6.8.2 Ongoing Studies	250
7	4 N T 4 '	LVTICAL ACTUODS	252
1.		LYTICAL METHODS	253
	7.1	BIOLOGICAL SAMPLES	253
		7.1.1 Internal Cobalt Measurements	253
	7.0	7.1.2 External Radiation Measurements	257
	7.2	ENVIRONMENTAL SAMPLES	259
			260
	7.2	7.2.2 Laboratory Analysis of Environmental Samples	260
	7.3	ADEQUACY OF THE DATABASE  7.3.1 Identification of Data Needs	<ul><li>266</li><li>266</li></ul>
		7.3.1 Identification of Data Needs	
		1.3.4 Ongoing studies	∠∪ /

COBALT xviii

8.	REG	ULATIONS AND ADVISORIES	269
9.	REFE	ERENCES	291
10	. GLC	DSSARY	377
ΑF	PENI	DICES	
	A.	ATSDR MINIMAL RISK LEVELS AND WORKSHEETS	A-1
	B .	USER'S GUIDE	B-1
	C.	ACRONYMS, ABBREVIATIONS, AND SYMBOLS	C-1
	D.	OVERVIEW OF BASIC RADIATION PHYSICS, CHEMISTRY AND BIOLOGY	D-1

COBALT xix

## **LIST OF FIGURES**

3-1.	Levels of Significant Exposure to Cobalt - Chemical Toxicity - Inhalation	38
3-2.	Levels of Significant Exposure to Cobalt - Chemical Toxicity - Oral	65
3-3.	Levels of Significant Exposure to Cobalt - Radiation Toxicity - External	93
3-4.	Transfer Parameters for Cobalt Following Inhalation of Cobalt Oxide (Co <sub>3</sub> O <sub>4</sub> ) Particles, Showing the Fractions of the Lung Content, L(t), and Time, t, Cleared Per Day by Each Route	114
3-5.	Conceptual Representation of a Physiologically Based Pharmacokinetic (PBPK) Model for a Hypothetical Chemical Substance	129
3-6.	Respiratory Tract Compartments in Which Particles May be Deposited	132
3-7.	Reaction of Gases or Vapors at Various Levels of the Gas-Blood Interface	133
3-8.	Compartment Model to Represent Time-Dependent Particle Transport in the Respiratory Tract	137
3-9.	The Human Respiratory Tract Model: Absorption into Blood	139
3-10.	ICRP Biokinetics Model for Cobalt	141
3-11.	Relation Between Mean Cobalt Exposure and Mean Blood Concentration of Cobalt in Exposed Workers	156
3-12.	Existing Information on Health Effects of Stable Cobalt	162
3-13.	Existing Information on Health Effects of Radioactive Cobalt	163
6-1.	Frequency of NPL Sites with Cobalt Contamination	198
6-2.	Frequency of NPL Sites with Cobalt 60 Contamination	199

COBALT xxi

## **LIST OF TABLES**

3-1.	Levels of Significant Exposure to Cobalt - Chemical Toxicity - Inhalation	30
3-2.	Levels of Significant Exposure to Cobalt - Chemical Toxicity - Oral	54
3-3.	Levels of Significant Exposure to Cobalt - Chemical Toxicity - Dermal	80
3-4.	Levels of Significant Exposure to Cobalt - Radiation Toxicity - External	82
3-5.	Genotoxicity of Cobalt In Vitro	109
3-6.	Initial (Day 3) Lung Deposits of Cobalt Oxide and Summary of Lung Retention at 90 and 180 Days	116
3-7.	Summary of Measurements of Retention and Excretion After Intragastric Administration of Cobalt Oxide ( $Co_3O_4$ ) Particles	117
3-8.	Peak Translocation and Average Mechanical Clearance Rates After Inhalation of Cobalt Oxide	123
3-9.	Summary of Measurements of Retention and Excretion of Cobalt Following Injection of Cobalt Nitrate Co(NO <sub>3</sub> ) <sub>2</sub> Solution	126
3-10.	Reference Respiratory Values for a General Caucasian Population at Different Levels of Activity	131
3-11.	Reference Values of Parameters for the Compartment Model to Represent Time-dependent Particle Transport from the Human Respiratory Tract	135
3-12.	Cobalt Exposure Concentrations and Amounts in the Blood and Urine of Subjects Examined	155
<b>4-</b> 1.	Chemical Identity of Cobalt and Selected Compounds	176
4-2.	Physical and Chemical Properties of Cobalt and Selected Compounds	180
4-3.	Principal Radioactive Cobalt Isotopes	184
5-1.	Current U.S. Manufacturers of Cobalt Metal and Selected Cobalt Compounds	187
5-2.	Facilities that Produce, Process, or Use Cobalt or Cobalt Compounds	189
6-1.	Releases to the Environment from Facilities that Produce, Process, or Use Cobalt or Cobalt Compounds	203
6-2.	Concentration of Cobalt in the Atmosphere	221
6-3.	Cobalt Levels in Water	225

COBALT xxii

5-4.	Cobalt Levels in Sediment	229
5-5.	Cobalt Levels in Food	232
6-6.	Cobalt Content of Miscellaneous Substances	235
6-7.	Mean Daily Dietary Intake of Cobalt for Selected Population Groups in Canada	237
5-8.	Cobalt Levels in Human Tissues and Fluids	239
<b>5-9</b> .	Ongoing Studies on Cobalt	251
7-1.	Analytical Methods for Determining Stable Cobalt in Biological Materials	254
7-2.	Analytical Methods for Determining Radioactive Cobalt in Biological Samples	256
7-3.	Analytical Methods for Determining Stable Cobalt in Environmental Samples	261
7-4.	Analytical Methods for Determining Radioactive Cobalt in Environmental Samples	263
7-5.	Ongoing Analytical Studies on Cobalt	268
8-1.	Regulations and Guidelines Applicable to Stable Cobalt	271
3-2.	Regulations and Guidelines Applicable to Radioactive Cobalt	281

## 1. PUBLIC HEALTH STATEMENT

This public health statement tells you about cobalt and the effects of exposure.

The Environmental Protection Agency (EPA) identifies the most serious hazardous waste sites in the nation. These sites make up the National Priorities List (NPL) and are the sites targeted for long-term federal cleanup activities. Cobalt has been found in at least 404 of the 1,585 current or former NPL sites. <sup>60</sup>Co has been found in at least 11 of the 1,585 current or former NPL sites. However, the total number of NPL sites evaluated for cobalt is not known. As more sites are evaluated, the sites at which cobalt is found may increase. This information is important because exposure to this substance may harm you and because these sites may be sources of exposure.

When a substance is released from a large area, such as an industrial plant, or from a container, such as a drum or bottle, it enters the environment. This release does not always lead to exposure. You are exposed to a substance only when you come in contact with it. You may be exposed by breathing, eating, or drinking the substance, or by skin contact.

If you are exposed to cobalt, many factors determine whether you'll be harmed. These factors include the dose (how much), the duration (how long), and how you come in contact with it. You must also consider the other chemicals you're exposed to and your age, sex, diet, family traits, lifestyle, and state of health.

## 1.1 WHAT IS COBALT?

Cobalt is a naturally-occurring element that has properties similar to those of iron and nickel. It has an atomic number of 27. There is only one stable isotope of cobalt, which has an atomic weight of 59. (An element may have several different forms, called isotopes, with different weights depending on the number of neutrons that it contains. The isotopes of an element, therefore, have different atomic weights [number of protons and neutrons], although the atomic number [number of protons] remains the same.) However, there are many unstable or

radioactive isotopes, two of which are commercially important, cobalt-60 and cobalt-57, also written as Co-60 or <sup>60</sup>Co and Co-57 or <sup>57</sup>Co, and read as cobalt sixty and cobalt fifty-seven. All isotopes of cobalt behave the same chemically and will therefore have the same chemical behavior in the environment and the same chemical effects on your body. However, isotopes have different weights and the radioactive isotopes have different radioactive properties, such as their half-life and the nature of the radiation they give off. The half-life of a cobalt isotope is the time that it takes for half of that isotope to give off its radiation and change into a different isotope. Radioactive isotopes are constantly changing into different isotopes by giving off radiation, a process referred to as radioactive decay. The new isotope may be a different element or the same element with a different mass.

Small amounts of cobalt are naturally found in rocks, soil, water, plants, and animals. Cobalt is also found in meteorites. Elemental cobalt is a hard, silvery grey metal. However, cobalt is usually found in the environment combined with other elements such as oxygen, sulfur, and arsenic. Small amounts of these "chemical compounds" can be found in rocks, soil, plants, and animals. Cobalt is even found in water in dissolved or ionic form, typically in small amounts. (Ions are atoms, collections of atoms, or molecules containing a positive or negative electric charge.) A biochemically important cobalt compound is vitamin  $B_{12}$  or cyanocobalamin. Vitamin  $B_{12}$  is essential for good health in animals and humans. Cobalt is not currently mined in the United States. Therefore, we obtain cobalt and its other chemical forms from imported materials and by recycling scrap metal that contains cobalt.

Cobalt metal is usually mixed with other metals to form alloys. The cobalt makes the alloy hard or resistant to wear and corrosion. These alloys are used in a number of military and industrial applications such as aircraft engines, magnets, and grinding and cutting tools. They are also used in artificial hip and knee joints. Cobalt compounds are used as colorants in glass, ceramics, and paints, as catalysts, and as paint driers. Cobalt colorants have a characteristic blue color. Cobalt compounds are also used as trace element additives in agriculture and medicine.

<sup>60</sup>Co is the most important radioisotope of cobalt. It is produced by bombarding natural cobalt, <sup>59</sup>Co, with neutrons in a nuclear reactor. <sup>60</sup>Co decays by giving off a beta ray (or electron) with a

half-life of 5.27 years, transforming into a stable isotope of nickel (atomic number 28). The decay is accompanied by the emission of high energy radiation called gamma rays. <sup>60</sup>Co is used as a source of gamma rays for sterilizing medical equipment and consumer products, radiation therapy for treating cancer patients, and for manufacturing plastics. <sup>60</sup>Co has also been used for food irradiation; depending on the radiation dose, this process may be used to sterilize food, destroy pathogens, extend the shelf-life of food, disinfest fruits and grain, delay ripening, and retard sprouting (e.g., potatoes and onions). <sup>57</sup>Co is used in medical and scientific research and has a half-life of 272 days. Another important cobalt isotope, <sup>58</sup>Co, is produced when nickel is exposed to a source of neutrons. Since nickel is used in nuclear reactors, <sup>58</sup>Co may be unintentionally produced and appear as a contaminant in cooling water released by nuclear reactors. <sup>60</sup>Co may be similarly produced from cobalt alloys in nuclear reactors and released as a contaminant in cooling water. <sup>58</sup>Co has a half-life of 71 days and gives off beta and gamma radiation in the process.

Quantities of radioactive cobalt can be measured in units of mass (grams), as is the stable isotope, but are normally measured in units of radioactivity (curies or becquerels). The becquerel (Bq) is a new international unit, and the curie (Ci) is a traditional unit; both are currently used. A becquerel is the amount of radioactive material in which 1 atom transforms every second, and a curie is the amount of radioactive material in which 37 billion atoms transform every second. For an overview of basic radiation physics, chemistry, and biology see Appendix D of this profile. For more information on radiation, see the *ATSDR Toxicological Profile for Ionizing Radiation*.

To learn more about the properties and uses of cobalt, see Chapters 5 and 6.

#### 1.2 WHAT HAPPENS TO COBALT WHEN IT ENTERS THE ENVIRONMENT?

Cobalt occurs naturally in soil, rock, air, water, plants, and animals. It may enter air and water, and settle on land from wind-blown dust, seawater spray, volcanic eruptions and forest fires and may additionally get into surface water from runoff and leaching when rainwater washes through soil and rock containing cobalt. Small amounts of cobalt may be released into the atmosphere

from coal-fired power plants and incinerators, vehicular exhaust, industrial activities relating to the mining and processing of cobalt-containing ores, and the production and use of cobalt alloys and chemicals. <sup>58</sup>Co and <sup>60</sup>Co may be released to the environment as a result of nuclear accidents, radioactive waste dumping in the sea or in radioactive waste landfills, and nuclear power plant operations.

Cobalt cannot be destroyed in the environment. It can only change its form or become attached or separated from particles. Cobalt released from power plants and other combustion processes are usually attached to very small particles. Cobalt contained in wind-borne soil is generally found in larger particles than those released from power plants. These large particles settle to the ground or are washed out of the air by rain. Cobalt that is attached to very small particles may stay in the air for many days. Cobalt released into water may stick to particles in the water column or to the sediment at the bottom of the body of water into which it was released, or remain in the water column in ionic form. The specific fate of cobalt will depend on many factors such as the chemistry of the water and sediment at a site as well as the cobalt concentration and water flow. Cobalt deposited on soil is often strongly attached to soil particles and therefore would not travel very far into the ground. However, the form of the cobalt and the nature of the soil at a particular site will affect how far cobalt will penetrate into the soil. Both in soil and sediment, the amount of cobalt that is mobile will increase under more acidic conditions. Ultimately, most cobalt ends up in the soil or sediment.

When plants grow in highly contaminated soil, they accumulate very small amounts of cobalt, especially in the parts of the plant that you eat most often like the fruit, grain, and seeds. While animals that eat these plants will accumulate cobalt, cobalt is not known to biomagnify (produce increasingly higher concentrations) up the food chain. Therefore, vegetables, fruits, fish, and meat that you consume will generally not contain high amounts of cobalt. Cobalt is an essential element, required for good health in animals and humans, and therefore, it is important that foodstuffs contain adequate quantities of cobalt.

<sup>60</sup>Co and <sup>58</sup>Co are moderately short-lived, man-made radioactive isotopes that are produced in nuclear reactors. Small amounts may be released to the environment as contaminants in cooling

water or in radioactive waste. However, these isotopes are not produced in nuclear weapons testing and are not associated with nuclear fallout. In the environment, radioactive isotopes of cobalt will behave chemically like stable cobalt. However, <sup>60</sup>Co and <sup>58</sup>Co will also undergo radioactive decay according to their respective half-lives, 5.27 years and 71 days.

For more information about what happens to cobalt in the environment, see Chapter 7.

#### 1.3 HOW MIGHT I BE EXPOSED TO COBALT?

Cobalt is widely dispersed in the environment. You may be exposed to small amounts of cobalt by breathing air, drinking water, and eating food containing it. Children may also be exposed to cobalt by eating dirt. You may also be exposed by skin contact with soil, water, cobalt alloys, or other substances that contain cobalt. Analytical methods used by scientists to determine the levels of cobalt in the environment generally do not determine the specific chemical form of cobalt present. Therefore, we do not always know the chemical form of cobalt a person may be exposed to. Similarly, we do not know what forms of cobalt are present at hazardous waste sites. Some forms of cobalt may be insoluble or so tightly attached to particles or embedded in minerals that they are not taken up by plants and animals. Other forms of cobalt that are weakly attached to particles may accumulate in plants and animals.

The concentration of cobalt in soil varies widely, generally ranging from about 1 to 40 ppm (1 ppm = 1 part of cobalt in a million parts of soil by weight), with an average level of 7 ppm. Soils containing less than about 3 ppm of cobalt are considered cobalt-deficient because plants growing on them do not have sufficient cobalt to meet the dietary requirements of cattle and sheep. Such cobalt-deficient soils are found in some areas in the southeast and northeast parts of the United States. On the other hand, soils near cobalt-containing mineral deposits, mining and smelting facilities, or industries manufacturing or using cobalt alloys or chemicals may contain much higher levels of cobalt.

Usually, the air contains very small amounts of cobalt, less than 1–2 nanograms per cubic meter (ng/m³). The amount of cobalt that you breathe in a day is much less than what you consume in

food and water. You may breathe in higher levels of cobalt in dust in areas near cobalt-related industries or near certain hazardous waste sites.

The concentration of cobalt in surface and groundwater in the United States is generally low—between 1 and 10 parts of cobalt in 1 billion parts of water (ppb) in populated areas; concentration may be hundreds or thousands times higher in areas that are rich in cobalt-containing minerals or in areas near mining or smelting operations. In most drinking water, cobalt levels are less than 1–2 ppb.

For most people, food is the largest source of cobalt intake. The average person consumes about 11 micrograms of cobalt a day in their diet. Included in this food is vitamin  $B_{12}$ , which is found in meat and diary products. The recommended daily intake of vitamin  $B_{12}$  is 6 micrograms (1 microgram = 1 millionth part of a gram).

You may also be exposed to higher levels of cobalt if you work in industries that make or use cutting or grinding tools, in metal mining, smelting, and refining, or in other industries that produce or use cobalt metal and cobalt compounds. Exposure will be reduced when exhaust systems are used in the workplace. Exposure will primarily result from breathing cobalt-containing dust.

When we speak of exposure to <sup>60</sup>Co, we are interested in exposure to the radiation given off by this isotope, primarily the gamma rays. The general population is rarely exposed to this radiation unless a person is undergoing radiation therapy. However, workers at nuclear facilities, irradiation facilities, or nuclear waste storage sites may be exposed to <sup>60</sup>Co or <sup>58</sup>Co. Exposures to radiation at these facilities are regulated and carefully monitored and controlled.

You can find more information on how you may be exposed to cobalt in Chapter 7.

### 1.4 HOW CAN COBALT ENTER AND LEAVE MY BODY?

Cobalt can enter your body when you breathe in air containing cobalt dust, when you drink water that contains cobalt, when you eat food that contains cobalt, or when your skin touches materials that contain cobalt. If you breathe in air that contains cobalt dust, the amount of inhaled cobalt that stays in your lungs depends on how big the dust particles are. The amount that is then absorbed into your blood depends on how well it will dissolve. If the particles dissolve easily, then it is easier for the cobalt to pass into your blood from the particles in your lungs. If the particles dissolve slowly, then they will remain in your lungs longer. Some of the particles will leave your lungs as they normally clean themselves out. Some of the particles will be swallowed into your stomach. The most likely way you will be exposed to excess cobalt is by eating contaminated food or drinking contaminated water. Levels of cobalt normally found in the environment, however, are not high enough to result in excess amounts of cobalt in food or water. The amount of cobalt that enters your body from food or water depends on many things including your state of health, the amount you eat or drink, and the number of days, weeks, or years you eat foods or drink fluids containing cobalt. If you do not have enough iron in your body, the body may absorb more cobalt from the foods you eat. Once cobalt enters your body, it goes into all tissues, mainly into the liver, kidney, and bones. After cobalt is breathed in or eaten, some of it leaves the body quickly in the feces. The rest is absorbed into the blood and then into the tissues throughout the body. The absorbed cobalt leaves the body slowly, mainly in the urine. Tests have shown that cobalt does not enter the body through normal skin, but it can if the skin has been cut.

Further information on how cobalt can enter or leave your body can be found in Chapter 3.

#### 1.5 HOW CAN COBALT AFFECT MY HEALTH?

Cobalt has both beneficial and harmful effects on human health. Cobalt is beneficial for humans because it is part of vitamin  $B_{12}$ , which is essential to maintain human health. Cobalt (0.16-1.0 mg cobalt/kg of body weight) has also been used as a treatment for anemia (less than normal number of red blood cells), particularly in pregnant women, because it causes red blood

cells to be produced. Cobalt also increases red blood cell production in healthy people, but only at very high exposure levels. Cobalt is also essential for the health of various animals, such as cattle and sheep. Exposure of humans and animals to levels of cobalt normally found in the environment is not harmful.

When too much cobalt is taken into your body, however, harmful health effects can occur. People who breathed air containing 0.038 mg cobalt/m³ (about 100,000 times the concentration normally found in air) for 6 hours had trouble breathing. Serious effects on the lungs, including asthma, pneumonia, and wheezing have been found in people exposed to 0.003 mg cobalt/m³ at work. People exposed to 0.007 mg cobalt/m³ at work have also developed allergies to cobalt that resulted in asthma and skin rashes. The general public, however, is not likely to be exposed to the same type or amount of cobalt dust that caused these effects in workers.

In the 1960s, some breweries added cobalt to beer to stabilize the foam (0.04–0.14 mg cobalt/kg). Some people who drank excessive amounts of beer (8–25 pints/day) experienced serious effects on the heart. In some cases, these effects resulted in death. Vomiting and nausea usually occurred before the effects on the heart were noticed. Cobalt is no longer added to beer so you will not be exposed from this source. The effects on the heart, however, may have also been due to the fact that the beer-drinkers had protein-poor diets and may have already had heart damage from alcohol abuse. Effects on the heart were not seen, however, in people with anemia treated with up to 1 mg cobalt/kg, or in pregnant women with anemia treated with 0.6 mg cobalt/kg. Effects on the thyroid were found in people exposed to 0.5 mg cobalt/kg for a few weeks. Vision problems were found in one man following treatment with 1.3 mg cobalt/kg for 6 weeks, but this effect has not been seen in other human or animal studies.

Being exposed to radioactive cobalt can be very dangerous to your health. If you come near radioactive cobalt, cells in your body can become damaged from gamma rays that can penetrate your entire body, even if you do not touch the radioactive cobalt. Radiation from radioactive cobalt can also damage cells in your body if you eat, drink, breathe, or touch anything that contains radioactive cobalt. The amount of damage depends on the amount of radiation to which you are exposed, which is related to the amount of activity in the radioactive material and the

length of time that you are exposed. Most of the information regarding health effects from exposure to radiation comes from exposures for only short time periods. The risk of damage from exposure to very low levels of radiation for long time periods is not known. If you are exposed to enough radiation, you might experience a reduction in white blood cell number, which could lower your resistance to infections. Your skin might blister or burn, and you may lose hair from the exposed areas. This happens to cancer patients treated with large amounts of radiation to kill cancer. Cells in your reproductive system could become damaged and cause temporary sterility. Exposure to lower levels of radiation might cause nausea, and higher levels can cause vomiting, diarrhea, bleeding, coma, and even death. Exposure to radiation can also cause changes in the genetic materials within cells and may result in the development of some types of cancer.

To protect the public from the harmful effects of toxic chemicals and to find ways to treat people who have been harmed, scientists use many tests.

One way to see if a chemical will hurt people is to learn how the chemical is absorbed, used, and released by the body. In the case of a radioactive chemical, it is also important to gather information concerning the radiation dose and dose rate to the body. For some chemicals, animal testing may be necessary. Animal testing may also be used to identify health effects such as cancer or birth defects. Without laboratory animals, scientists would lose a basic method to get information needed to make wise decisions to protect public health. Scientists have the responsibility to treat research animals with care and compassion. Laws today protect the welfare of research animals, and scientists must comply with strict animal care guidelines.

Studies in animals suggest that exposure to high amounts of nonradioactive cobalt during pregnancy might affect the health of the developing fetus. Birth defects, however, have not been found in children born to mothers who were treated with cobalt for anemia during pregnancy. The doses of cobalt used in the animal studies were much higher than the amounts of cobalt to which humans would normally be exposed.

Non-radioactive cobalt has not been found to cause cancer in humans or in animals following exposure in the food or water. Cancer has been shown, however, in animals who breathed cobalt or when cobalt was placed directly into the muscle or under the skin. Based on the animal data, the International Agency for Research on Cancer (IARC) has determined that cobalt is possibly carcinogenic to humans.

Much of our knowledge of cobalt toxicity is based on animal studies. Cobalt is essential for the growth and development of certain animals, such as cows and sheep. Short-term exposure of rats to high levels of cobalt in the air results in death and lung damage. Longer-term exposure of rats, guinea pigs, hamsters, and pigs to lower levels of cobalt in the air results in lung damage and an increase in red blood cells. Short-term exposure of rats to high levels of cobalt in the food or drinking water results in effects on the blood, liver, kidneys, and heart. Longer-term exposure of rats, mice, and guinea pigs to lower levels of cobalt in the food or drinking water results in effects on the same tissues (heart, liver, kidneys, and blood) as well as the testes, and also causes effects on behavior. Sores were seen on the skin of guinea pigs following skin contact with cobalt for 18 days. Generally, cobalt compounds that dissolve easily in water are more harmful than those that are hard to dissolve in water. More information on how cobalt can affect your health can be found in Chapter 3.

Much of what we know about the effects of radioactive cobalt comes from studies in animals. The greatest danger of radiation seen in animals is the risk to the developing animal, with even moderate amounts of radiation causing changes in the fetus. High radiation doses in animals have also been shown to cause temporary or permanent sterility and changes in the lungs, which affected the animals' breathing. The blood of exposed animals has lower numbers of white blood cells, the cells that aid in resistance to infections, and red blood cells, which carry oxygen in the blood. Radioactive cobalt exposures in animals have also caused genetic damage to cells, cancer, and even death.

More information on how cobalt can affect your health can be found in Chapter 3.

## 1.6 HOW CAN COBALT AFFECT CHILDREN?

This section discusses potential health effects from exposures during the period from conception to maturity at 18 years of age in humans.

Children can be exposed to cobalt in the same ways as adults. In addition, cobalt may be transferred from the pregnant mother to the fetus or from the mother to the infant in the breast milk. Children may be affected by cobalt the same ways as adults. Studies in animals have suggested that children may absorb more cobalt from foods and liquids containing cobalt. Babies exposed to radiation while in their mother's womb are believed to be much more sensitive to the effects of radiation than adults.

### 1.7 HOW CAN FAMILIES REDUCE THE RISK OF EXPOSURE TO COBALT?

If your doctor finds that you have been exposed to significant amounts of cobalt, ask whether your children might also be exposed. Your doctor might need to ask your state health department to investigate.

Children living near waste sites containing cobalt are likely to be exposed to higher environmental levels of cobalt through breathing, touching soil, and eating contaminated soil. Some children eat a lot of dirt. You should discourage your children from eating dirt. Make sure they wash their hands frequently and before eating. Discourage your children from putting their hands in their mouths or hand-to-mouth activity.

*Stable Cobalt.* Since cobalt is naturally found in the environment, we cannot avoid being exposed to it. However, the relatively low concentrations present do not warrant any immediate steps to reduce exposure. If you are accidentally exposed to large amounts of cobalt, consult a physician immediately.

*Radioactive Cobalt.* You are unlikely to be exposed to high levels of radioactive cobalt unless you are exposed as part of a radiotherapy treatment, there is an accident involving a cobalt

sterilization or radiotherapy unit, or there is an accidental release from a nuclear power plant. In such cases, follow the advice of public health officials who will publish guidelines for reducing exposure to radioactive material when necessary. Workers who work near or with radioactive cobalt should follow the workplace safety guidelines of their institution carefully to reduce the risk of accidental irradiation.

## 1.8 IS THERE A MEDICAL TEST TO DETERMINE WHETHER I HAVE BEEN EXPOSED TO COBALT?

We have reliable tests that can measure cobalt in the urine and the blood for periods up to a few days after exposure. The amount of cobalt in your blood or urine can be used to estimate how much cobalt you were exposed to. The tests are not able to accurately predict potential health effects following exposure to cobalt.

It is difficult to determine if a person has been exposed only to external radiation from radioactive cobalt unless the radiation dose was rather large. Health professionals examining people who have health problems similar to those resulting from radiation exposure would need to rely on additional information in order to establish if such people had been near a source of radioactivity. More information on medical tests can be found in Chapters 3 and 7.

## 1.9 WHAT RECOMMENDATIONS HAS THE FEDERAL GOVERNMENT MADE TO PROTECT HUMAN HEALTH?

The federal government develops regulations and recommendations to protect public health. Regulations <u>can</u> be enforced by law. Federal agencies that develop regulations for toxic substances include the Environmental Protection Agency (EPA), the Occupational Safety and Health Administration (OSHA), the Food and Drug Administration (FDA), and the Nuclear Regulatory Commission (NRC).

Recommendations provide valuable guidelines to protect public health but <u>cannot</u> be enforced by law. Federal organizations that develop recommendations for toxic substances include the

1. PUBLIC HEALTH STATEMENT

Agency for Toxic Substances and Disease Registry (ATSDR) and the National Institute for

Occupational Safety and Health (NIOSH), and the FDA.

Regulations and recommendations can be expressed in not-to-exceed levels in air, water, soil, or

food that are usually based on levels that affect animals; then they are adjusted to help protect

people. Sometimes these not-to-exceed levels differ among federal organizations because of

different exposure times (an 8-hour workday or a 24-hour day), the use of different animal

studies, or other factors.

Recommendations and regulations are also periodically updated as more information becomes

available. For the most current information, check with the federal agency or organization that

provides it. Some regulations and recommendations for cobalt include the following:

EPA requires that the federal government be notified if more than 1,000 pounds of cobalt (as the

bromide, formate, and sulfamate compounds) are released into the environment in a 24-hour

period. OSHA regulates levels of nonradioactive cobalt in workplace air. The limit for an

8-hour workday, 40-hour workweek is an average of 0.1 mg/m<sup>3</sup>.

1.10 WHERE CAN I GET MORE INFORMATION?

If you have any more questions or concerns, please contact your community or state health or

environmental quality department, your regional Nuclear Regulatory Commission office, or:

Agency for Toxic Substances and Disease Registry

Division of Toxicology

1600 Clifton Road NE, Mailstop E-29

Atlanta, GA 30333

\* Information line and technical assistance

Phone: 1-888-42-ATSDR (1-888-422-8737)

Fax: (404) 498-0057

#### 1. PUBLIC HEALTH STATEMENT

ATSDR can also tell you the location of occupational and environmental health clinics. These clinics specialize in recognizing, evaluating, and treating illnesses resulting from exposure to hazardous substances.

\* To order toxicological profiles, contact

National Technical Information Service 5285 Port Royal Road Springfield, VA 22161

Phone: 1-800 -553-6847 or (703) 605-6000

COBALT 15

# 2. RELEVANCE TO PUBLIC HEALTH

# 2.1 BACKGROUND AND ENVIRONMENTAL EXPOSURES TO COBALT IN THE UNITED STATES

Cobalt is a naturally-occurring element that has properties similar to those of iron and nickel. There is only one stable isotope of cobalt, <sup>59</sup>Co. However, there are many radioactive isotopes, two of which are commercially important, <sup>60</sup>Co and <sup>57</sup>Co. Cobalt is the 33<sup>rd</sup> most abundant element in the earth's crust. Its average concentrations in the earth's crust and in igneous rocks are 20–25 and 18 mg/kg, respectively. The United States is the world's largest consumer of cobalt. Cobalt is used in a number of essential military and industrial applications. The largest use of metallic cobalt is in superalloys that are used in gas turbine aircraft engines. Cobalt compounds are used as pigments in glass, ceramics, and paints; as catalysts in the petroleum industry; as paint driers; and as trace element additives in agriculture and medicine. <sup>60</sup>Co is produced by irradiating <sup>59</sup>Co with thermal neutrons in a nuclear reactor, and is used as a source of gamma rays for sterilizing medical equipment or consumer products, food irradiation, radiation therapy for treating cancer patients, and for manufacturing plastics. <sup>57</sup>Co decays to an excited state of <sup>57</sup>Fe, the most widely used x-ray source in Mössbauer spectroscopy.

The primary anthropogenic sources of cobalt in the environment are from the burning of fossil fuels and sewage sludge, phosphate fertilizers, mining and smelting of cobalt-containing ores, processing of cobalt-containing alloys, and industries that use or process cobalt compounds. Cobalt released to the atmosphere is deposited onto soil or water surfaces by wet and dry deposition. In soils, cobalt generally has low mobility and strong adsorption. However its mobility increases in moist, acidic soils. In water, cobalt largely partitions to sediment and suspended solids in the water column; however, the amount that is adsorbed to suspended solids is highly variable.

Exposure of the general population to cobalt occurs through inhalation of ambient air and ingestion of food and drinking water. In general, exposure from food sources is much greater than from drinking water and air. The cobalt intake in food has been estimated to be 5.0–40.0 μg/day. The general population is not significantly exposed to radioactive forms of cobalt. Cancer patients being treated with radiation therapy may be exposed to gamma rays from a <sup>60</sup>Co source; however, external exposure to gamma radiation is not unique to <sup>60</sup>Co, but is similar for all gamma-emitting radionuclides. Occupational exposure to cobalt occurs for workers in the hard metal industry (tool production, grinding, etc.) and in

industries such as coal mining, metal mining, smelting and refining, cobalt dye painters, and the cobalt chemical production industry. The concentrations of cobalt in the air of hard metal manufacturing, welding, and grinding factories may range from 1 to 300  $\mu$ g/m³, compared to normal atmospheric levels of 0.4–2.0  $\mu$ g/m³. Workers at nuclear facilities and nuclear waste storage sites may be exposed to potentially high levels of radioactive cobalt.

#### 2.2 SUMMARY OF HEALTH EFFECTS

Overview. Cobalt is essential in the body in that it is a component of cyanocobalamin (vitamin  $B_{12}$ ). Vitamin  $B_{12}$  acts as a coenzyme in many enzymatic reactions, most notably in a methyl transfer reaction that converts homocysteine to methionine and in a separate reaction that converts L-methylmalonylcoenzyme A (CoA) to succinyl-CoA. Vitamin  $B_{12}$  is also involved in some enzymes involved in hematopoiesis; deficiency can lead to pernicious anemia. It has been identified in liver, muscle, lung, lymph nodes, heart, skin, bone, hair, stomach, brain, pancreatic juice, kidneys, plasma, and urinary bladder of nonexposed subjects, with the highest cobalt concentration found in the liver. No other essential function of cobalt has been reported. The Recommended Dietary Allowance (RDA) for vitamin  $B_{12}$  for adults is 2.4 µg/day, which contains 0.1 µg of cobalt.

Cobalt has been found to produce adverse effects by the inhalation, oral, and dermal routes. Effects in humans following inhalation exposure to cobalt included lung effects (respiratory irritation, fibrosis, asthma, pneumonia, wheezing), cardiovascular effects (cardiomyopathy), liver and kidney congestion, ocular effects (congestion of the conjunctiva), and weight loss. Effects in humans observed following ingestion of cobalt, as cobalt sulfate in beer or as cobalt chloride as a treatment for anemia, included cardiomyopathy, gastrointestinal effects, visual disturbances, and thyroid effects. Cobalt dermatitis and sensitization as a result of dermal exposure to cobalt are well documented.

Exposure to sufficient levels of radiation from cobalt radionuclides may also produce adverse health effects, including respiratory pneumonitis and fibrosis, pericarditis, gastrointestinal effects, including atrophy and fibrosis of the stomach and intestines, reduced immunologic and hematologic indices, hypocellularity of bone marrow, dermal effects (including acneform reactions, hair loss, and skin degeneration), neurodegenerative and neurobehavioral effects, damage to reproductive tissues, teratogenesis, and cancer. These symptoms are the same as those experienced following exposure to other gamma-emitting isotopes.

Cobalt is essential for the growth and development of ruminants. Overexposure of other animals to cobalt resulted in effects similar to those in humans following inhalation and dermal exposure. After ingestion of cobalt, effects in animals were similar to effects in humans although some additional effects, including hypothermia, neurological effects (effects on reactivity), developmental effects (stunted fetuses), and reproductive effects (testicular degeneration and atrophy), were found in animals, but not humans. In the animal studies, the doses tested were higher than the documented levels to which humans have been exposed, or would be expected to be exposed.

Issues relevant to children are explicitly discussed in 3.6 Children's Susceptibility and 6.6 Exposures of Children.

**Death.** Lethal cardiomyopathy in humans was reported following repeated inhalation of airborne stable cobalt or ingestion of beer that contained cobalt. Inhalation exposure levels associated with cardiomyopathy have not been determined. In the 1960s, breweries in the United States, Canada, and Europe added cobalt salts to beer to improve foaming properties at the tap. Several deaths occurred among heavy beer drinkers who consumed beer containing 0.04–0.14 mg cobalt/kg/day (8–30 pints of beer daily). The addition of cobalt to beer has since been discontinued. Although the ingestion of cobalt was identified as a key causative factor in the beer drinkers cardiomyopathy, other etiologic factors were significant, including heavy alcohol consumption and related nutritional deficits. Repeated oral ingestion of 1 mg cobalt/kg/day to raise the hematocrit of anemic, but otherwise healthy, patients did not cause cardiac injury.

Acute exposure to high levels of radiation from a radioactive cobalt source resulted in the death of an exposed worker from acute radiation sickness. This result is not specific to cobalt radiation, and applies to all intense gamma ray emitting radionuclides.

In animals, deaths from inhalation exposure of stable cobalt were related to respiratory effects and secondary infections. Deaths in animals following oral exposure resulted from cardiomyopathy or from multiple lesions (kidney, liver, and heart lesions). Acute lethality in animals varies with the chemical form administered, with soluble compounds generally being more toxic than insoluble compounds. In rats, cobalt fluoride is more toxic than cobalt chloride by a factor of two. With the exception of tricobalt tetroxide, the  $LD_{50}$  values of the cobalt compounds for which acute oral lethality data are available (Table 3-2) all lie within the same order of magnitude when expressed in terms of the cobalt ion.

Tricobalt tetroxide ( $LD_{50} > 3,672$  mg cobalt/kg) is insoluble in water and, therefore, is relatively nontoxic. The lethal dose following exposure to cobalt radiation in animals varies with the species, portion of body exposed, dose rate, and duration of exposure.

**Systemic Effects.** The primary target organ systems for the effects of stable cobalt in humans are the respiratory system following inhalation exposure and the cardiac and hematopoietic systems following oral exposure. Following exposure to cobalt radiation, the most sensitive target is the developing animal, with pronounced effects also seen in the respiratory system, reproductive organs, gastrointestinal tract, blood, and nervous system.

Respiratory Effects. Effects on the respiratory system include irritation, fibrosis, asthma, pneumonia, and wheezing following inhalation exposure to stable cobalt. Individuals can develop a sensitivity to cobalt, and inhalation exposure to airborne cobalt can precipitate asthmatic attacks in sensitized individuals. Studies in animals report similar effects following inhalation exposure. Intermediate-duration inhalation studies in rats and mice reported that the larynx was the part of the respiratory tract most sensitive to the effects of cobalt, with the lungs, nose, and trachea being affected at higher exposure levels. Exposure to radiation may damage lung tissue, following a two-phase pattern. The first phase of damage usually consists of radiation pneumonitis, which occurs between 3 and 13 weeks after irradiation and is characterized by low-grade fever, mild exertional dyspnea, congestion, and unproductive cough. The second phase is characterized by radiation-induced lung fibrosis, emphysema, and pleural thickening.

Cardiovascular Effects. In humans, lethal cardiomyopathy resulted from oral and inhalation exposure to stable cobalt. Along with the severe cardiac effects, beer-cobalt cardiomyopathy was characterized by initial effects on the gastrointestinal system (vomiting, nausea, diarrhea), pulmonary rales and edema (resulting from the cardiac failure), liver injury (resulting from hepatic ischemia), and polycythemia. Beer-cobalt cardiomyopathy was similar to both alcoholic cardiomyopathy and beriberi, except that beer-cobalt cardiomyopathy had an abrupt onset, characterized by left ventricular failure, cardiogenic shock, polycythemia, and acidosis. Evidence that ingestion of ethanol was not required for development of cobalt cardiomyopathy came from studies in animals. A cardiomyopathy similar to that observed in humans occurred in guinea pigs after repeated exposure to cobalt (20 mg/kg/day) in foods, with or without ethanol consumption. Following exposure to cobalt radiation, cardiovascular effects consist mainly of pericarditis and alterations in endothelial cell permeability.

Gastrointestinal Effects. Gastrointestinal effects, including nausea, vomiting, and diarrhea, were reported in humans after ingestion of cobalt-contaminated beer and treatment with cobalt for anemia, though only a small percentage of anemic patients reported these symptoms. No effects on the gastrointestinal system were reported in animals after inhalation or oral exposure. Exposure to sufficient cobalt radiation may result in dramatic gastrointestinal effects, including vomiting, bloody stools, fibrosis, and atrophy of gastrointestinal cells.

Hematological Effects. Because stable cobalt induces polycythemia in humans following high-dose oral exposure, it has been used in the treatment of anemia. Polycythemia has not been observed in humans following inhalation exposure. Animal data show increased hematocrit and hemoglobin levels following both oral and inhalation exposure. Although increases in hematocrit in both humans and animals do not necessarily constitute an adverse effect, extreme elevation is known to increase the risk of clotting, vascular anomalies, and possible stroke. In contrast to the effects of stable cobalt, radiation from cobalt isotopes causes diminished levels of circulating erythrocytes and hemoglobin, as well as a diminished hematopoietic activity.

*Musculoskeletal Effects.* No studies were located demonstrating musculoskeletal effects in humans or animals following exposure to stable cobalt. The musculoskeletal system has been found to be relatively resistant to adverse effects following radiation exposure from radioactive cobalt and other radionuclides.

Hepatic Effects. No conclusive evidence that stable cobalt is a direct liver toxicant in humans has been reported following exposure to low levels; however, liver injury has been associated with cobalt-related cardiomyopathy following either inhalation or oral exposure. Although the mechanism for the liver effects is not known, it is likely that hepatic ischemia related to cardiovascular impairment is a significant causative factor. Liver injury was observed in animals orally exposed to near lethal levels of cobalt. Whether this represents a direct effect of cobalt on the liver or an indirect effect of cardiac impairment is not known. A direct effect of cobalt on the liver is plausible since the liver is the major site of accumulation of orally absorbed cobalt. Exposure to radiation from cobalt isotopes does not appear to cause significant hepatic changes unless the radiation dose is large.

**Renal Effects.** No conclusive evidence that stable cobalt is a kidney toxicant in humans has been reported. Congestion of the kidneys, however, has been associated with cobalt cardiomyopathy resulting from occupational exposure to cobalt. Effects on the proximal tubules of the kidneys were observed in

animals orally exposed to cobalt. Adverse effects on the kidneys of both humans and animals are a possibility because a substantial amount of cobalt absorbed into the blood is excreted in the urine. Data are not available on the effects of exposure to radiocobalt in humans, but animal studies have demonstrated a marked reduction in renal function following exposure to high levels of <sup>60</sup>Co radiation.

Endocrine Effects. Studies examining the endocrine effects of stable cobalt in humans have generally been equivocal. A decrease in iodine uptake by the thyroid resulted from acute oral exposure of humans to 1 mg cobalt/kg/day or longer-term exposure to 0.54 mg/kg/day. Exposure of animals to cobalt compounds has caused changes in histopathology of the thyroid and increased incidence of tumors of the adrenal medulla. In various species of animals, parenteral administration of cobalt resulted in cytotoxic effects on the alpha cells of the pancreas. Because this effect has never been reported in humans or animals following inhalation, oral, or dermal exposure to cobalt, the relevance of the effect to humans is not known. One study in humans exposed to cobalt radiotherapy showed that some patients developed hypothyroidism, with significant decreases in T<sub>4</sub> levels. Animal studies have shown no effect of cobalt radiation on levels of follicle stimulating hormone (FSH), luteinizing hormone (LH), prolactin, or testosterone.

**Dermal Effects.** No dermal effects have been reported following inhalation or oral exposure to stable cobalt compounds. Dermatitis is a common effect following dermal cobalt exposure in humans, and probably occurs due to an allergic reaction. Exposure to cobalt radiation may result in skin lesions, hair loss, and skin cancer.

*Ocular Effects.* Effects on the human eye have been observed following occupational exposure (congestion of the conjunctiva) and oral exposure (optic atrophy, impaired choroidal perfusion) to stable cobalt. Humans exposed during cobalt radiotherapy have developed vision disorders, including cataracts.

**Body Weight Effects.** Weight loss was found in workers occupationally exposed to stable cobalt. Similar weight loss was seen in animals. In addition, time- and dose-related hypothermia was observed in rats given cobalt orally. Available studies examining exposure to cobalt radiation have not examined alterations in body weight.

Immunological and Lymphoreticular Effects. Exposure to stable cobalt can lead to sensitization. In its most serious form, cobalt-sensitization can result in or exacerbate asthma. Dermal sensitization and related cobalt-dermatitis have also been described. The mechanism for cobalt sensitization is not completely understood. Antibodies to cobalt have been detected in individuals sensitized to cobalt, suggesting that a humoral immune response may be a component of the sensitization phenomenon. Exposure of humans to high radiation doses from cobalt isotopes results in a decrease in circulating white blood cells, particularly leucocytes and neutrophils.

**Neurological Effects.** No studies were located regarding neurological effects in humans following inhalation, oral, or dermal exposure to stable cobalt. Enhanced behavioral reactivity to stress, a slower rate of lever pushing, and effects on conditioned reflexes were observed in rats orally exposed to cobalt. The relevance of these findings to humans is not known. In rats, cobalt applied directly to the brain has been found to induce epilepsy and has been used extensively as a model toward a better understanding of epilepsy in humans. Isolated cases of neurological damage from cobalt radiation during radiotherapy in humans have been reported, but the results are not consistent. Animals exposed to cobalt radiation have shown changes in behavioral patterns.

Reproductive Effects. No studies were located regarding reproductive effects in humans following inhalation, oral, or dermal exposure to stable cobalt. Following both inhalation and oral exposure of animals to cobalt, adverse effects on the testes were observed (degeneration, atrophy, decreased weight). An increase in the length of the estrous cycle was also reported in female mice following inhalation exposure (NTP 1991). Because no effects on the reproductive system were found in patients who died as a result of beer-cobalt cardiomyopathy, the significance of the animal results to humans is not clear. While human data are limited, numerous animal studies have shown profound effects of cobalt radiation on the reproductive tissues of both sexes, including reduced sperm count, decreased testicular weight, decreased reproductive performance, diminished implantation, and decreased numbers of offspring per litter.

**Developmental Effects.** No obvious developmental effects were observed in human fetuses from mothers who were given stable cobalt orally to counteract decreases in hematocrit and hemoglobin levels that often occur during pregnancy. No studies were located regarding developmental effects in humans following inhalation or dermal exposure. Animal studies, however, reported that oral exposure to cobalt results in developmental effects including stunted fetuses, a decrease in the number of litters and average

litter weights, and an increase in the number of dead pups per litter. Toxic maternal effects were also observed in this study. The relevance of the effects found in animals to possible human effects is not known. However, exposure of male mice to cobalt (26 mg/kg/day) for 12 weeks had no effect on the offspring in the F1 generation. While human data on developmental effects following radiocobalt exposure are lacking, exposure of animals to cobalt radiation has shown dramatic effects on multiple organ systems, even at acute radiation doses as low as 10 rad (0.1 Gy) (see Section 3.2.4.6).

**Cancer.** Cobalt has not been shown to cause cancer in humans by the inhalation, oral, or dermal exposure routes. An occupational study reported an increased incidence of death from lung cancer (SMR=4.66) in workers exposed to cobalt, but the difference was not statistically significant. Occupational studies of hard metal exposure in humans, however, have demonstrated increased mortality from lung cancers. A lifetime inhalation exposure study of cobalt sulfate in rats and mice showed significant increases in tumor formation in both sexes of both species, with the respiratory tract being the primary site of tumor formation.

The induction of tumors (fibrosarcomas) following intramuscular injection of cobalt oxide into rats has been shown. No tumors were induced in mice after intramuscular injection of cobalt. Tumors were also induced following subcutaneous and intrathoracic injections in rats. The significance of these results to humans is not clear because these are not physiological routes of exposure and no tumors were found in humans with metal-alloy prostheses. IARC, however, has classified cobalt and cobalt compounds as group 2B, possible human carcinogens.

The carcinogenic effects of ionizing radiation are well-documented. Several human studies exist wherein cobalt radiation, given as external radiotherapy, later led to an increased incidence of cancer, generally of the skin of the treated areas.

#### 2.3 MINIMAL RISK LEVELS

#### Inhalation MRLs

• An MRL of  $1x10^{-4}$  mg cobalt/m³ has been derived for chronic-duration inhalation exposure (>365 days) to cobalt.

An MRL for inhalation exposure to cobalt was derived for chronic duration only. The chronic inhalation MRL of  $1x10^{-4}$  mg cobalt/m³ was based on a NOAEL of 0.0053 mg cobalt/m³ and LOAEL of  $15.1 \mu g$ 

cobalt/m³ for decreases in forced vital capacity (FVC), forced expiratory volume in one second (FEV<sub>1</sub>), forced expiratory flow between 25 and 75% of the FVC (MMEF), and mean peak expiratory flow rate (PEF) in diamond polishers. The National Toxicology Program (NTP) has conducted a chronic-duration carcinogenicity study in rats and mice. Exposure of rats and mice to aerosols of cobalt (as cobalt sulfate) at concentrations ranging from 0.11 to 1.14 mg cobalt/m³ for 2 years resulted in a spectrum of inflammatory, fibrotic, and proliferative lesions in the respiratory tract of male and female rats and mice. Squamous metaplasia of the larynx occurred in rats and mice at exposure concentrations of 0.11 mg cobalt/m³ and above, with severity of the lesion increasing with increased exposure concentration. Hyperplastic lesions of the nasal epithelium occurred in rats at concentrations of 0.11 mg cobalt/m³ and above, and in mice at concentrations of 0.38 mg cobalt/m³ and above. Both sexes of rats had greatly increased incidences (>90% incidence) of alveolar lesions at all exposure levels, including inflammatory changes, fibrosis, and metaplasia. Similar changes were seen in mice at all exposure levels, though the changes in mice were less severe. The study in diamond polishers, being a well-conducted study in humans, was selected as the critical study for the derivation of a MRL because it examined a human population and defined a NOAEL, neither of which occurred in the NTP study.

An acute inhalation MRL was not derived because the threshold was not defined for human effects and animal studies reported effects that were serious and occurred at levels above those reported in the few human studies. An acute-duration study of hard metal exposure in humans was not utilized for MRL derivation because the toxicity of hard metal is not directly due to cobalt metal, but rather to an interaction between cobalt metal and tungsten carbide. An intermediate-duration MRL was not derived because available studies did not examine the dose-response relationship at low doses; the chronic inhalation MRL should be protective for intermediate exposures (see Appendix A). The chronic inhalation MRL was derived by adjusting the NOAEL of 0.0053 mg cobalt/m³ for intermittent exposure (8 hours/24 hours x 5 days/7 days), and dividing by an uncertainty factor of 10 (for human variability). It should be noted that this MRL may not be protective for individuals already sensitive to cobalt.

## Oral MRLs

• An MRL of  $1x10^{-2}$  mg cobalt/kg-day has been derived for intermediate-duration oral exposure (<365 days) to cobalt.

An intermediate-duration MRL of 1x10<sup>-2</sup> mg cobalt/kg/day was derived based on a LOAEL of 1 mg cobalt/kg-day for polycythemia as reported in a study by Davis and Fields. The authors exposed six male volunteers to 120 or 150 mg/day of cobalt chloride (~1 mg Co/kg/day) for up to 22 days. Exposure to

cobalt resulted in the development of polycythemia in all six patients, with increases in red blood cell numbers ranging from 0.5 to 1.19 million (~16–20% increase above pre-treatment levels). Polycythemic erythrocyte counts returned to normal 9–15 days after cessation of cobalt administration. An 8-week study in rats also reported increases in erythrocyte number, with a NOEL of 0.6 mg/kg-day and a LOEL of mg/kg/day.

Oral MRL values were not derived for acute or chronic exposure to cobalt. An acute MRL was not derived because the reported effects in animals were serious and occurred at levels above those reported in the few human oral studies. No chronic oral studies were available for humans or animals; therefore, a chronic oral MRL was not derived for cobalt.

Acute-, intermediate-, and chronic-duration dermal MRLs were not derived for cobalt due to the lack of appropriate methodology for the development of dermal MRLs.

#### MRLs for External Exposure to Cobalt Isotopes

Two MRLs have been derived for ionizing radiation and are applicable to external exposure to radioisotopes of cobalt:

• An MRL of 400 mrem (4.0 mSv) has been derived for acute-duration external exposure to ionizing radiation (14 days or less).

The acute MRL is based on results of a study in which neurological effects of radiation, measured by intelligence test scores, were evaluated in children 10–11 years of age who had been exposed at critical stages of fetal development (gestation weeks 8–15) during the atomic bombing of Hiroshima and Nagasaki. When IQ scores were regressed on radiation dose estimates, IQ diminished linearly with increasing dose, resulting in an estimated decrease in IQ score of approximately 25 points per 100 rad (or 100 rem in dose equivalent) or 0.25 points/rem (25 points/Sv). To derive the MRL of 400 mrem (4.0 mSv), ATSDR divided the dose associated with a predicted change of 0.25 IQ points/rem by an uncertainty factor of 3 (for human variability and/or the potential existence of sensitive populations). ATSDR noted that a change in IQ points of 0.25 is less than the reported difference of 0.3 IQ points between separated and unseparated identical twins.

The Nuclear Regulatory Commission (NRC) set a radiation exposure limit of 500 mrem (5 mSv) for pregnant working women over the full gestational period. For the critical gestational period of

8–15 weeks, ATSDR believes that the acute MRL of 400 mrem (4 mSv) is consistent with the NRC limit and could be applied to either acute (0–14-day) or intermediate (15–365-day) exposure periods.

• An MRL of 100 mrem/year (1.0 mSv/year) above background has been derived for chronic-duration external ionizing radiation (365 days or more).

The MRL is based on the BEIR V report that the average annual effective dose of ionizing radiation to the U.S. population is 360 mrem/year (3.6 mSv/year), a dose not expected to produce adverse non-cancerous health effects. This dose is obtained mainly by naturally-occurring radiation from external sources, medical uses of radiation, and radiation from consumer products. An uncertainty factor of 3 (for human variability) was applied to the NOAEL of 360 mrem/year to derive the MRL of 100 mrem/year.

COBALT 27

### 3. HEALTH EFFECTS

#### 3.1 INTRODUCTION

The primary purpose of this chapter is to provide public health officials, physicians, toxicologists, and other interested individuals and groups with an overall perspective on the toxicology of cobalt. It contains descriptions and evaluations of toxicological studies and epidemiological investigations and provides conclusions, where possible, on the relevance of toxicity and toxicokinetic data to public health.

A glossary and list of acronyms, abbreviations, and symbols can be found at the end of this profile.

## 3.2 DISCUSSION OF HEALTH EFFECTS BY ROUTE OF EXPOSURE

To help public health professionals and others address the needs of persons living or working near hazardous waste sites, the information in this section is organized first by route of exposure (inhalation, oral, and dermal) and then by health effect (death, systemic, immunological, neurological, reproductive, developmental, genotoxic, and carcinogenic effects). These data are discussed in terms of three exposure periods: acute (14 days or less), intermediate (15–364 days), and chronic (365 days or more).

Levels of significant exposure for each route and duration are presented in tables and illustrated in figures. The points in the figures showing no-observed-adverse-effect levels (NOAELs) or lowest-observed-adverse-effect levels (LOAELs) reflect the actual doses (levels of exposure) used in the studies. LOAELS have been classified into "less serious" or "serious" effects. "Serious" effects are those that evoke failure in a biological system and can lead to morbidity or mortality (e.g., acute respiratory distress or death). "Less serious" effects are those that are not expected to cause significant dysfunction or death, or those whose significance to the organism is not entirely clear. ATSDR acknowledges that a considerable amount of judgment may be required in establishing whether an end point should be classified as a NOAEL, "less serious" LOAEL, or "serious" LOAEL, and that in some cases, there will be insufficient data to decide whether the effect is indicative of significant dysfunction. However, the Agency has established guidelines and policies that are used to classify these end points. ATSDR believes that there is sufficient merit in this approach to warrant an attempt at distinguishing between "less serious" and "serious" effects. The distinction between "less serious" effects and "serious" effects is considered to be important because it helps the users of the profiles to identify levels of exposure at which

major health effects start to appear. LOAELs or NOAELs should also help in determining whether or not the effects vary with dose and/or duration, and place into perspective the possible significance of these effects to human health.

The significance of the exposure levels shown in the Levels of Significant Exposure (LSE) tables and figures may differ depending on the user's perspective. Public health officials and others concerned with appropriate actions to take at hazardous waste sites may want information on levels of exposure associated with more subtle effects in humans or animals (LOAELs) or exposure levels below which no adverse effects (NOAELs) have been observed. Estimates of levels posing minimal risk to humans (Minimal Risk Levels or MRLs) may be of interest to health professionals and citizens alike.

Estimates of exposure levels posing minimal risk to humans (Minimal Risk Levels or MRLs) have been made for cobalt. An MRL is defined as an estimate of daily human exposure to a substance that is likely to be without an appreciable risk of adverse effects (noncarcinogenic) over a specified duration of exposure. MRLs are derived when reliable and sufficient data exist to identify the target organ(s) of effect or the most sensitive health effect(s) for a specific duration within a given route of exposure. MRLs are based on noncancerous health effects only and do not consider carcinogenic effects. MRLs can be derived for acute, intermediate, and chronic duration exposures for inhalation and oral routes. Appropriate methodology does not exist to develop MRLs for dermal exposure.

Although methods have been established to derive these levels (Barnes and Dourson 1988; EPA 1990), uncertainties are associated with these techniques. Furthermore, ATSDR acknowledges additional uncertainties inherent in the application of the procedures to derive less than lifetime MRLs. As an example, acute inhalation MRLs may not be protective for health effects that are delayed in development or are acquired following repeated acute insults, such as hypersensitivity reactions, asthma, or chronic bronchitis. As these kinds of health effects data become available and methods to assess levels of significant human exposure improve, these MRLs will be revised.

A User's Guide has been provided at the end of this profile (see Appendix B). This guide should aid in the interpretation of the tables and figures for Levels of Significant Exposure and the MRLs.

Studies have shown that soluble cobalt compounds are generally more acutely toxic than insoluble cobalt compounds. When expressed in terms of the cobalt ion for the sake of comparison, however, the

differences are within an order of magnitude and therefore do not warrant presentation in separate LSE tables and figures. Therefore, data regarding both soluble and insoluble cobalt compounds are presented in Tables 3-1, 3-2, 3-3, and 3-4.

# 3.2.1 Inhalation Exposure

#### 3.2.1.1 Death

*Stable Cobalt.* Conclusive evidence for human deaths related to inhalation exposure to cobalt has not been reported; however, results of several studies and case reports suggest a possible relationship between exposure and deaths from lung cancer and cardiomyopathy, respectively.

In general, available cohort studies in humans have not reported a significant increase in total mortality as a result of cobalt exposure. Several studies have noted increased mortality rates resulting from lung cancer following occupational exposure to cobalt, either as a mixture of cobalt compounds (Mur et al. 1987) or as hard metal, a metal alloy with a tungsten carbide and cobalt matrix (Lasfargues et al. 1994; Moulin et al. 1998). Fatal cases of hard metal disease (Figueroa et al. 1992; Ruokonen et al. 1996) and cardiomyopathy (Barborik and Dusek 1972) believed to have resulted from occupational cobalt exposure have also been reported. However, in the majority of these and other reported occupational studies, co-exposure to other substances was common, and unable to be corrected for in the analysis.

Cobalt can be lethal in animals if inhalation exposure is sufficiently high or prolonged. The acute LC<sub>50</sub> for a 30-minute inhalation exposure in rats was 165 mg cobalt/m³ as cobalt hydrocarbonyl (Palmes et al. 1959). Exposure to 9 mg cobalt/m³ as cobalt hydrocarbonyl for 6 hours/day, 5 days/week for 3 months resulted in 16 deaths out of 75 rats (Palmes et al. 1959). Death was reported in rats and mice exposed to 19 mg cobalt/m³ (but not 1.9 mg cobalt/m³) as cobalt sulfate over 16 days, but exposure to 11.4 mg cobalt/m³ over 13 weeks was lethal only to mice and not to rats (Bucher et al. 1990; NTP 1991). Exposure to 1.14 mg cobalt/m³ as cobalt sulfate for 104 weeks resulted in no increase in mortality in rats and mice of either sex (Bucher et al. 1999; NTP 1998). Lethal levels for each species and duration category are recorded in Table 3-1 and plotted in Figure 3-1.

Table 3-1. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Inhalation

		-			LOAEL		· ————
Key to <sup>a</sup> figure	Species (strain)	Exposure/ duration/ frequency	System	NOAEL (mg/m3)	Less serious (mg/m3)	Serious (mg/m3)	Reference Chemical Form
Α	CUTE EX	POSURE					
s	ystemic						Maradan ak al
1	Human	6 hr	Resp		0.038 (bronchial irritation, reduced FVC)		Kusaka et al. 1986b Hard Metal
2	Rat SD-Jcl	5 hr	Resp	2.72			Kyono et al. 1992 Metal
3	Rat SD-Jcl	4 d	Resp		2.12 M (Slight damage to respiratory tissues, assessed by electron microscopy)		Kyono et al. 1992 Metal
4	Rat	30 min	Resp	7	26 (edema)	83 (severe edema)	Palmes et al. 1959 Hydrocarbony

Table 3-1. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Inhalation (continued)

						LOAEL	·		
Key to	Species (strain)	Exposure/ duration/ frequency	System	NOAEL (mg/m3)	Less ser (mg/m		Serie (mg		Reference Chemical Form
11	NTERMED	IATE EXPOS	URE						
C	Death								NTP 1991
5	Rat	16 d 5 d/wk 6 hr/d		1.9			19	(2/5 males died)	Sulfate
6	Mouse	13 wk 5 d/wk 6 hr/d		3.8			11.4	(2 males died)	NTP 1991 Sulfate
•	Systemic								NTD 4004
7	Rat	13 wk 5 d/wk	Resp		0.11	(laryngial squamous metaplasia and polyps)	0.38	(chronic inflammation of larynx)	NTP 1991 Sulfate
		6 hr/d	Cardio		11.4	(increase in severity of cardiomyopathy)			
			Hemato		1.14	И (polycythemia)⁵			
			Renal Bd Wt	11.4	11.4	(15% lower body weight in males)			
8	Rat	3 mo 5 d/wk	Resp		9	(lung inflamm)			Palmes et al. 1959 Hydrocarbonyl
		7 h/d	Hemato		9	(10% increase in hemoglobin) <sup>b</sup>			
			Bd Wt	9					

Table 3-1. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Inhalation (continued)

	Exposure/					LOAEL			
Key to <sup>‡</sup> figure		exposure/ duration/ frequency	System	NOAEL (mg/m3)	Less ser (mg/m		Serio (mg/r	us	Reference Chemical Form
		404	Doop	0.2	1.9	(respiratory tract	19	(necrosis)	NTP 1991
9	Mouse	16d 5 d/wk	Resp	0.2	1.0	inflammation)			Sulfate
		6 hr/d	Cardio	76					
			Gastro	76					
			Musc/skel	76					
			Hepatic				19	(necrosis)	
			Renal	76					
			Dermal	76				•	
					0.11	(larynx metaplasia)	1.14° F	(acute inflam of nose)	NTP 1991
10	Mouse	13 wk 5 d/wk	Resp		0.11	(larylix metaplasia)		,	Sulfate
		6 hr/d					3.8 N	1 (acute inflam of nose)	
			Gastro	11.4					
			Hemato	11.4					
			Musc/skel	11.4					
			Hepatic	11.4					
			Renal	11.4					
			Dermal	11.4					
			Bd Wt		11.4	(13-20% decrease in body weight)			
11	Gn Pig (Hartley)	66 d	Resp				2.4 F	<ul> <li>(Increased lung weight, increased retention of lavage fluid)</li> </ul>	Camner et al. 1993 Chloride
12	Gn Pig	3 mo 5 d/wk 7 h/d	Hemato		9	(5% increase in hemoglobin)⁵			Palmes et al. 1959 Hydrocarbony

Table 3-1. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Inhalation (continued)

						LOAEL			
Key to	Species (strain)	Exposure/ duration/ frequency	System	NOAEL (mg/m3)	Less ser (mg/m		Serio (mg/		Reference Chemical Form
13	Dog	3 mo	Hemato	9					Palmes et al. 1959
		3d/wk 7h/d							Hydrocarbonyl
			Bd Wt		9	(wt loss)			
14	Rabbit	4 mo 5 d/wk 6 h/d	Resp		0.4	(moderate lung inflammation)	2.0	(severe lung inflammation)	Johansson et al. 1987
45	Rabbit	4 mo	Resp	0.5 M					Johansson et al. 1991
15	Kappit	4 1110	rtoop						Chloride
16	Rabbit	4 mo	Resp		0.61	M (Histologic alterations in pulmonary tissue; altered BAL parameters)			Johansson et al. 1992 Chloride
17	Pig	3 mo 5d/wk	Resp		0.1	(decr compliance)			Kerfoot 1975 Metal
		6hr/d	Cardio		0.1	(EKG changes)			
			Hepatic	1.0					
			Renal Bd Wt	1.0	0.1	(decr wt gain)			
	Immunolog	ical/Lympho	eticular						NTP 1991
18	Rat	16 d 5 d/wk 6 hr/d			19	(necrosis of thymus)			Sulfate
19	Mouse	13 wk 5 d/wk 6 hr/d			11.4	(lymph node hyperplasia)			NTP 1991 Sulfate

Table 3-1. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Inhalation (continued)

					LOAEL		
Key to		Exposure/ duration/ frequency	System	NOAEL (mg/m3)	Less serious (mg/m3)	Serious (mg/m3)	Reference Chemical Form
ſ	Veurologica	ıl					NTD 4004
20	Rat	16 d 5 d/wk 6 hr/d			19 (congestion of vessels in brain)		NTP 1991 Sulfate
21	Mouse	16 d 5 d/wk 6 hr/d			19 (congestion of vessels in brain)		NTP 1991 Sulfate
	Reproductiv	ve					AUTT 4004
22	Rat	16 d 5 d/wk 6 hr/d				19 M (testes atrophy)	NTP 1991 Sulfate
23	Mouse	13 wk 5 d/wk 6 hr/d			1.14 M (decreased sperm motility)	11.4 (testes atrophy- increased length estrous cycle)	NTP 1991 Sulfate

Table 3-1. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Inhalation (continued)

					LOAEL		
(ey to figure	Species (strain)	Exposure/ duration/ frequency	System	NOAEL (mg/m3)	Less serious (mg/m3)	Serious (mg/m3)	Reference Chemical Form
С	HRONIC I	EXPOSURE					
S	ystemic						Deng et al.
24	Human	occup	Resp	0.0175			1991
							Metal
25	Human	occup	Resp		0.1355 (Decreased FEV1 and FVC ~10%; increased		Gennart and Lauwerys 1990
					cough, sputum, dyspnea)		Hard-Metal
26	Human	occup	Resp	0.0053 <sup>d</sup>	0.0151 (Decreased FEV1, FVC increased cough and	;	Nemery et al. 1992
					upper airway irritation)		Metal
07	Human	occup	Endocr		0.05 F (Decreased thyroid		Prescott et al. 1992
27	riuillali	оссар	2.1.400.		volume; increases in T₄ and FT₄I levels)		Zinc-Silicate Dye
28	Human	occup	Resp			.007 (asthma)	Shirakawa et al. 1988
							Hard Metal
29	Human	occup	Resp			.051 (interst lung dis)	Sprince et al. 1988
29	Human	0000p	1-				Hard Metal

Table 3-1. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Inhalation (continued)

		F							
ey to	Species	Exposure/ duration/ frequency	System	NOAEL (mg/m3)	Less ser (mg/m		Serio (mg/i		Reference Chemical Form
	Human	ıman 8 yr Resp		0.125	(Dyspnoea and wheezing)			Swennen et al. 1993	
						Wildozing			Metal
			Hemato		0.125	(Decreased red cell counts ~5%; decreased total hemoglobin ~4%)			
			Endocr		0.125	(Slight (~7%) decrease in T3 levels)			
			Dermal		0.125	(Eczema and erythema)			
	5.4	104 wk	Boon				0.11	(Hyper- and metaplasia of	NTP 1998
31	Rat (Fischer- 344)		Resp					respiratory tract tissues; pulmonary fibrosis)	Sulfate
			D		0.11	(Laryngial metaplasia)			NTP 1998
32	Mouse (B6C3F1)	104 wk	Resp		0.11	(Laryngiai motapiasis)			Sulfate
33	Hamster	life	Resp				7.9	(emphysema)	Wehner et al. 1977
		5d/wk 7h/d							Oxide
			Bd Wt	7.9					
ſ	mmunologic	cal/Lymphor	eticular						
34	Human	occup			0.007	(sensitization)			Shirakawa et al. 1986a
									Hard Metal
35	Human	8 yr			0.125	(Increased white cell count by 19%)			Swennen et al. 1993

Table 3-1. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Inhalation (continued)

					LOAEL	
Key to <sup>a</sup> figure	Species (strain)			Serious (mg/m3)	Reference Chemical Form	
	ancer	-			0.38 F (alveoloar/bronchiolar	NTP 1998
	Rat	104 wk			neoplasms)	Sulfate
(	(Fischer- 344	)			1.14 M (alveoloar/bronchiolar neoplasms)	
					1.14 F (pheochromocytoma)	
					0.38 F (Combined	NTP 1998
	Mouse (B6C3F1)	104 wk			alveolar/bronchiolar adenoma/carcinoma)	Sulfate
					1.14 M (Combined alveolar/bronchiolar adenoma/carcinoma)	

The number corresponds to entries in Figure 3-1.

Bd Wt = body weight; Cardio = cardiovascular; d = day(s); Derm = dermal; Endocr = endocrine; F = female; Gastro = gastrointestinal; Gn Pig = guinea pig; Hemato = hematological; hr = hour(s); LOAEL = lowest-observed-adverse-effect level; M = male; mo = month(s); Musc/skel = muscular/skeletal; NOAEL = no-observed- adverse-effect level; (occup) = occupational; Resp = respiratory; wk = week(s); yr = year(s);

An increase in hemoglobin or red blood cells (polycythemia) is not necessarily considered an adverse effect.

Differences in levels of health effects and cancer effects between males and females are not indicated in Figure 3-1. Where such differences exist, only the levels of effect for the most sensitive gender are

<sup>\*</sup>Used to derive a chronic inhalation Minimal Risk Level (MRL) of 1x10<sup>-4</sup> mg Co/m<sup>3</sup>; dose adjusted for intermittent exposure, and divided by an uncertainty factor of 10 (for human variability).

mg/m3 100 **⊕**4r 10 O4r O2r **1**3r

Systemic

Figure 3-1. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Inhalation Acute (≤14 days)

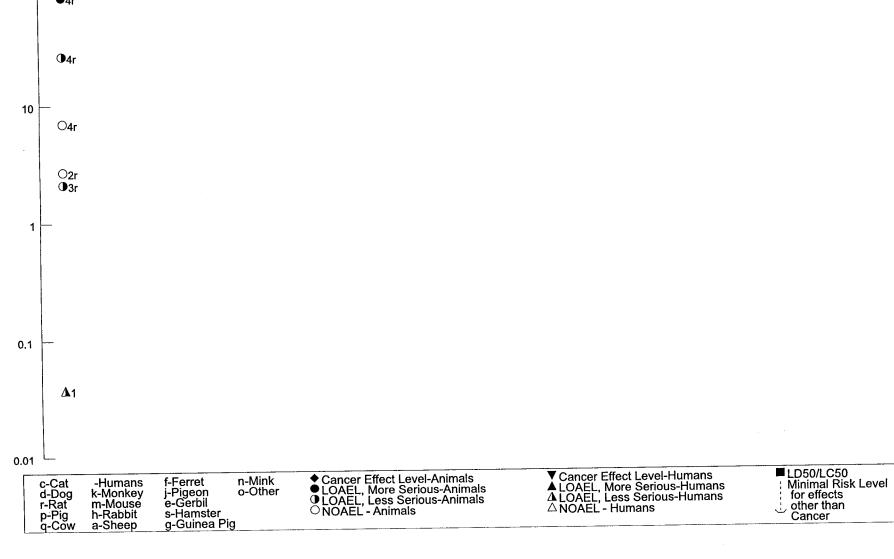


Figure 3-1. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Inhalation (continued)
Intermediate (15-364 days)

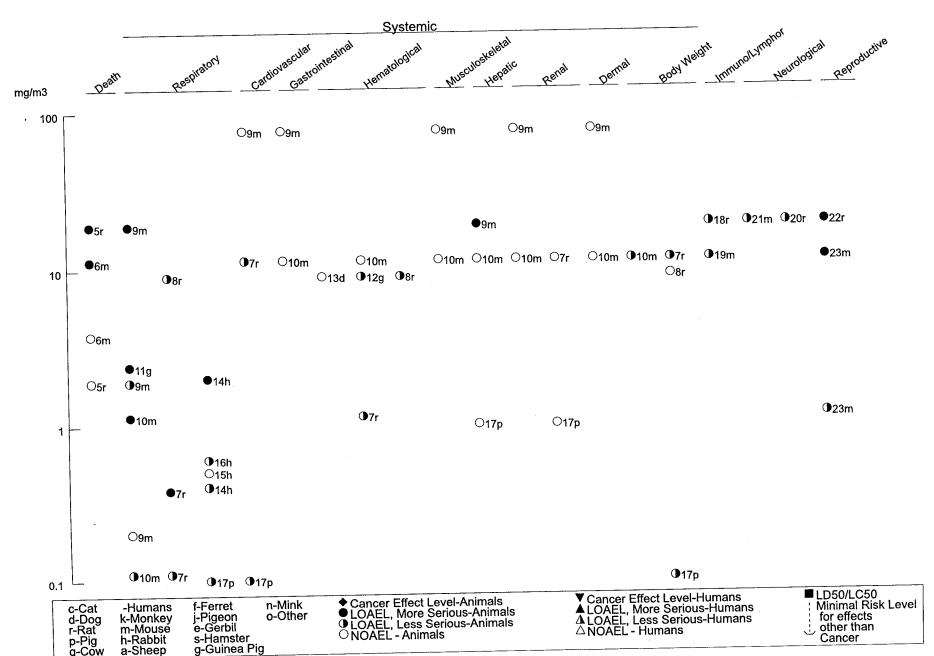
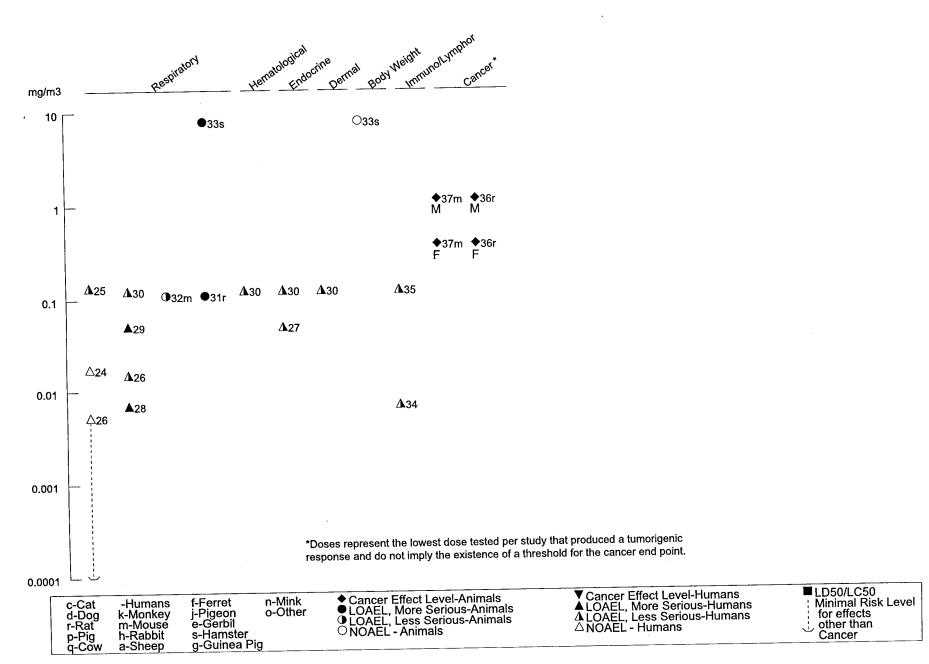


Figure 3-1. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Inhalation (*continued*)

Chronic (≥365 days)



**Radioactive Cobalt.** No studies were located regarding lethal effects in humans or animals following inhalation exposure to radioactive cobalt compounds.

# 3.2.1.2 Systemic Effects

Inhalation of cobalt by humans and/or animals resulted in respiratory, cardiovascular, hematological, hepatic, renal, ocular, and body weight effects. For each effect, the highest NOAEL values and all reliable LOAEL values for each species and duration category are reported in Table 3-1 and plotted in Figure 3-1.

# Respiratory Effects.

*Stable Cobalt.* The respiratory system is one of the primary target organs of inhaled cobalt. A significant decrease in ventilatory function due to chronic bronchial obstruction was observed in humans exposed to 0.038 mg cobalt/m³ as hard metal dust for 6 hours (Kusaka et al. 1986a).

Hard metal is a metal alloy with a tungsten carbide and cobalt matrix. It is used to make cutting tools because of its hardness and resistance to high temperature. Studies (Davison et al. 1983; Harding 1950) suggest that cobalt and not tungsten carbide is the probable causative agent for the respiratory effects observed in hard metal workers (see Section 3.5).

The effects of chronic occupational exposure to cobalt compounds on the respiratory system in humans are well-documented. These effects include respiratory irritation, diminished pulmonary function, wheezing, asthma, pneumonia, and fibrosis and occurred at exposure levels ranging from 0.007 to 0.893 mg cobalt/m³ (exposure from 2 to 17 years) (Anttila et al. 1986; Davison et al. 1983; Demedts et al. 1984a, 1984b; Deng et al. 1991; Gennart and Lauwerys 1990; Gheysens et al. 1985; Hartung et al. 1982; Kusaka et al. 1986b, 1996a, 1996b; Nemery et al. 1992; Raffn et al. 1988; Rastogi et al. 1991; Ruokonen et al. 1996; Shirakawa et al. 1988, 1989; Sprince et al. 1988; Swennen et al. 1993; Tabatowski et al. 1988; Van Cutsem et al. 1987; Zanelli et al. 1994). These effects have been observed in workers employed as hard metal workers, diamond polishers, and plate painters (painting with cobalt blue dye).

Kusaka et al. (1986b) described an acute exposure of 15 healthy young men to hard metal dust containing 38 μg cobalt/m³ for 6 hours. FVC was reduced, but no dose-response relation could be discerned. By contrast, 42 workers occupationally exposed to hard metal showed no decrease in ventilatory function at 85 μg cobalt/m³, but significant changes in FEV<sub>1</sub> (forced expiratory volume in 1 second) at 126 μg cobalt/m³ (Kusaka et al. 1986b). Several other studies of hard metal workers have shown respiratory effects, including decreased ventilatory function, wheezing, asthma, and fibrosis (Kusaka et al. 1996a, 1996b; Ruokonen et al. 1996; Zanelli et al. 1994), but have had less complete reports of exposure.

Swennen et al. (1993) performed a cross-sectional study on 82 workers in a cobalt refinery. Workers were examined for cobalt in blood and urine, a number of erythropoietic variables, thyroid metabolism, pulmonary function, skin lesions, and several serum enzymes. The concentrations of cobalt in blood and in urine after the shift were significantly correlated with those in air. The exposed workers complained more often (p<0.05) of dyspnoea and wheezing and had significantly more skin lesions (eczema, erythema) than control workers. A dose-effect relation was found between the reduction of the FEV and the intensity of the current exposure to cobalt, as assessed by measurement of cobalt in blood or urine.

Gennart and Lauwerys (1990) examined the ventilatory functions of 48 diamond polishing workers, relative to 23 control workers. Exposure occurred mainly in one of two rooms, with mean airborne concentrations of 15.2 and 135.5 μg cobalt/m³. Significant decreases in ventilatory function were found in exposed workers relative to controls. Duration of exposure played a significant factor, with no significant differences in workers who had been exposed for 5 years or less. Inhalation exposure to cobalt salts (exposure levels not reported) among glass bangle workers resulted in decreases in decreased ventilatory function, generally restrictive in nature, relative to controls (Rastogi et al. 1991). Swennen et al. (1993) reported that workers exposed to airborne cobalt metal, salts, or oxides (mean concentration 125 μg/m³, range 1–7,700) showed an increased prevalence of dyspnea and wheezing.

Nemery et al. (1992) conducted a cross-sectional study of cobalt exposure and respiratory effects in diamond polishers. Exposure occurred mainly from the generation of airborne cobalt resulting from the use of cobalt-containing polishing discs. The study groups were composed of 194 polishers working in 10 different workshops, and were divided into control, low-, and high-exposure groups. The low-exposure group (n=102) was exposed to an average of 5.3 µg cobalt/m³, based on personal sampling measurements, while the high dose group (n=92) received 15.1 µg cobalt/m³; there was considerable overlap in the total range of concentrations for the low- and high-exposure groups. Workers in the high-

exposure group were more likely than those in the other groups to complain about respiratory symptoms; the prevalence of eye, nose, and throat irritation and cough, as well as the fraction of these symptoms related to work, were significantly increased in the high-exposure group. Workers in the high-exposure group also had significantly reduced lung function compared to controls and low-exposure group workers, as assessed by FVC (forced vital capacity), FEV<sub>1</sub>, MMEF (forced expiratory flow between 25 and 75% of the FVC) and mean PEF (peak expiratory flow rate). Results in the low-exposure group did not differ from controls. Based on the NOAEL of 5.3 µg cobalt/m³ for decreased ventilatory function in exposed workers, a chronic inhalation MRL of 1x10<sup>-4</sup> mg cobalt/m³ was calculated as described in the footnote in Table 3-1. It should be noted that this MRL value may not be protective for some hypersensitive individuals.

Animals exposed to aerosols of cobalt oxides and cobalt sulfate developed respiratory effects that varied in severity with exposure level and duration. A single 30-minute exposure of rats to relatively high levels (26–236 mg cobalt/m³ as cobalt hydrocarbonyl) resulted in congestion, edema, and hemorrhage of the lung (Palmes et al. 1959). Prolonged exposure (3–4 months) of rats and rabbits to mixed cobalt oxides (0.4–9 mg cobalt/m³) resulted in lesions in the alveolar region of the respiratory tract characterized histologically by nodular accumulation of Type II epithelial cells, accumulations of enlarged highly vacuolated macrophages, interstitial inflammation, and fibrosis (Kyono et al. 1992; Johansson et al. 1984, 1987, 1991, 1992; Palmes et al. 1959). The lesions appeared to regress when exposure was terminated (Palmes et al. 1959). Guinea pigs sensitized to cobalt by repeated dermal application and then exposed to 2.4 mg cobalt/m³ as cobalt chloride showed pulmonary inflammatory changes (altered BAL fluid recovery, increased neutrophils and eosinophils following BAL) that were different than those in exposed animals not sensitized to cobalt (Camner et al. 1993). Decreased lung compliance was found in pigs exposed to 0.1 mg cobalt/m³ as cobalt dust for 3 months (Kerfoot 1975). Lifetime exposure of hamsters to 7.9 mg cobalt/m³ as cobalt oxide resulted in emphysema (Wehner et al. 1977).

Necrosis and inflammation of the respiratory tract epithelium (larynx, trachea, bronchioles, nasal turbinates) were reported in rats exposed to 19 mg cobalt/m³ and mice exposed to 1.9 mg cobalt/m³ (and above) as cobalt sulfate over 16 days (Bucher et al. 1990; NTP 1991). Exposure of rats and mice to cobalt as cobalt sulfate for 13 weeks resulted in adverse effects on all parts of the respiratory tract, with the larynx being the most sensitive part (Bucher et al. 1990; NTP 1991). At concentrations of 0.11 mg cobalt/m³ and above, rats and mice had squamous metaplasia of the larynx. Histiocytic infiltrates in the lung were also reported at similar levels in both the rats and mice. In rats, chronic inflammation of the

larynx was found at 0.38 mg cobalt/m³ and above, and more severe effects on the larynx, nose, and lung were reported at higher exposures. In mice, acute inflammation of the nose was found at 1.14 mg cobalt/m³ and above, and more severe effects on the larynx, nose, and lung were reported at higher exposures.

Exposure of rats and mice to aerosols of cobalt (as cobalt sulfate) at concentrations from 0.11 to 1.14 mg cobalt/m³ for 2 years resulted in a spectrum of inflammatory, fibrotic, and proliferative lesions in the respiratory tract of male and female rats and mice (Bucher et al. 1999; NTP 1998). Squamous metaplasia of the larynx occurred in rats and mice at exposure concentrations of 0.11 mg cobalt/m³ and above, with severity of the lesion increasing with increased exposure concentration. Hyperplastic lesions of the nasal epithelium occurred in rats at concentrations of 0.11 mg cobalt/m³ and above, and in mice at concentrations of 0.38 mg cobalt/m³ and above. Both sexes of rats had greatly increased incidences (>90% incidence) of alveolar lesions at all exposure levels, including inflammatory changes, fibrosis, and metaplasia. Similar changes were seen in mice at all exposure levels, though the changes in mice were less severe.

**Radioactive Cobalt.** No studies were located regarding respiratory effects in humans or animals after inhalation exposure to radioactive cobalt compounds.

## Cardiovascular Effects.

Stable Cobalt. Occupational exposure of humans to cobalt dust has been shown to result in cardiomyopathy characterized by functional effects on the ventricles and enlargement of the heart (Barborik and Dusek 1972; Horowitz et al. 1988; Jarvis et al. 1992), but the exposure levels associated with cardiac effects of inhaled cobalt in humans have not been determined. Jarvis et al. (1992) reported on two patients (exposure histories not specified) who were admitted to the emergency room for cardiac failures, which were believed to be associated with cobalt exposure. Barborik and Dusek (1972) reported a case of a 41-year-old man who was admitted to the hospital with cardiac failure following occupational exposure to cobalt; cobalt concentrations in heart, liver, lung, spleen, and kidney were elevated over two control patients. Horowitz et al. (1988) reported that in a cohort of 30 hard metal workers (exposure histories not specified), significant decreases in exercise right ventricular ejection fraction (EF) were seen in workers with abnormal chest x-rays relative to those with normal chest x-rays. It is possible that these effects were secondary to the respiratory effects of inhaled cobalt. It was concluded that cobalt is a weak

cardiomyopathic agent following occupational exposure (Horowitz et al. 1988). Cardiomyopathy is a characteristic toxic effect of cobalt following oral exposure in both humans and animals (Section 3.2.2.2).

In rats, exposure to 11.4 mg cobalt/m³ as cobalt sulfate over 13 weeks resulted in a marginal increase in the severity of cardiomyopathy as compared to controls (minimal-mild versus minimal) (Bucher et al. 1990; NTP 1991). Cardiomyopathy was not observed in mice exposed to 76 mg cobalt/m³ or less as cobalt sulfate over 16 days (Bucher et al. 1990; NTP 1991), nor in mice or rats exposed to up to 1.14 mg cobalt/m³ for 2 years (Bucher et al. 1999; NTP 1998). Electrocardiogram abnormalities that may reflect ventricular impairment have been observed in miniature swine (n=5) exposed to 0.1 mg cobalt dust/m³ for 6 hours/day, 5 days/week for 3 months (Kerfoot 1975).

**Radioactive Cobalt.** No studies were located regarding cardiovascular effects in humans or animals after inhalation exposure to radioactive cobalt compounds.

#### **Gastrointestinal Effects.**

*Stable Cobalt.* No studies were located regarding gastrointestinal effects in humans after inhalation exposure to stable cobalt.

No histological lesions were reported in the esophagus, stomach, duodenum, ileum, jejunum, cecum, colon, or rectum of rats or mice of either sex exposed to 76 mg cobalt/m³ or less as cobalt sulfate for 16 days, up to 11.4 mg cobalt/m³ for 13 weeks, or up to 1.14 mg cobalt/m³ for 104 weeks (Bucher et al. 1990, 1999; NTP 1991, 1998).

*Radioactive Cobalt.* No studies were located regarding gastrointestinal effects in humans or animals after inhalation exposure to radioactive cobalt compounds.

# Hematological Effects.

**Stable Cobalt.** Swennen et al. (1993) reported slightly, but significantly, decreased levels of red cells and total hemoglobin (~4–5% decreases) in a group of 82 workers occupationally exposed to 0.125 mg

cobalt/m<sup>3</sup>. No other studies were located regarding hematological effects in humans after inhalation exposure to cobalt.

Increased levels of hemoglobin and increased numbers of basophils and monocytes have been observed in rats and guinea pigs, but not in dogs, exposed to 9 mg cobalt/m³ as cobalt hydrocarbonyl for 3 months (Palmes et al. 1959). Polycythemia was reported in rats, but not mice, exposed to 1.14 mg cobalt/m³ as cobalt sulfate for 13 weeks (Bucher et al. 1990; NTP 1991).

*Radioactive Cobalt.* No studies were located regarding hematological effects in humans or animals after inhalation exposure to radioactive cobalt compounds.

#### Musculoskeletal Effects.

*Stable Cobalt.* No studies were located regarding musculoskeletal effects in humans after inhalation exposure to cobalt.

No histological lesions were reported in the sternebrae, including the bone marrow, of rats or mice exposed to 76 mg cobalt/m³ or less as cobalt sulfate for 16 days, up to 11.4 mg cobalt/m³ for 13 weeks, or up to 1.14 mg cobalt/m³ for 104 weeks (Bucher et al. 1990, 1999; NTP 1991, 1998) (see the above section on respiratory effects for detailed descriptions of exposure conditions).

*Radioactive Cobalt.* No studies were located regarding musculoskeletal effects in humans or animals after inhalation exposure to radioactive cobalt compounds.

## **Hepatic Effects.**

*Stable Cobalt.* Congestion of the liver was observed upon autopsy of a metal worker (exposure history not reported) who had been occupationally exposed to an unknown level of cobalt for 4 years (Barborik and Dusek 1972). The cause of death was determined to be cardiomyopathy.

Necrosis and congestion of the liver were observed in both rats and mice that died following exposure to 19 mg cobalt/m<sup>3</sup> as cobalt sulfate over 16 days (Bucher et al. 1990; NTP 1991). No histological effects

on the liver were found in pigs exposed to up to 1.0 mg cobalt/m³ as cobalt metal dust for 3 months (Kerfoot 1975).

**Radioactive Cobalt.** No studies were located regarding hepatic effects in humans or animals after inhalation exposure to radioactive cobalt compounds.

#### Renal Effects.

*Stable Cobalt.* Congestion of the kidneys was observed upon autopsy of a metal worker who had been occupationally exposed to an unknown level of cobalt for 4 years (Barborik and Dusek 1972). The cause of death was determined to be cardiomyopathy.

A significant increase in the relative weight of the kidneys was reported in male rats exposed to 0.11 mg cobalt/m³ and above as cobalt sulfate for 13 weeks (Bucher et al. 1990; NTP 1991). No effects were observed upon histological examination of the kidneys in rats or mice following exposure to 76 mg cobalt/m³ or less as cobalt sulfate for 16 days, up to 11.4 mg cobalt/m³ for 13 weeks, or up to 1.14 mg cobalt/m³ for 104 weeks (Bucher et al. 1990, 1999; NTP 1991, 1998). No histological effects on the kidneys were found in pigs exposed to up to 1.0 mg cobalt/m³ as cobalt metal for 3 months (Kerfoot 1975).

*Radioactive Cobalt.* No studies were located regarding renal effects in humans or animals after inhalation exposure to radioactive cobalt compounds.

#### **Dermal Effects.**

*Stable Cobalt.* No studies were located regarding dermal effects in humans or animals after inhalation exposure to stable cobalt.

**Radioactive Cobalt.** No studies were located regarding dermal effects in humans or animals after inhalation exposure to radioactive cobalt compounds.

#### Ocular Effects.

*Stable Cobalt.* Congestion of the conjunctiva was observed in a metal worker after occupational exposure to an unknown level of cobalt for 4 years (Barborik and Dusek 1972). Upon autopsy, the cause of death was determined to be cardiomyopathy.

No histological lesions were reported in the eyes or on the skin of rats or mice exposed to 76 mg cobalt/m³ or less as cobalt sulfate for 16 days, up to 11.4 mg cobalt/m³ for 13 weeks, or up to 1.14 mg cobalt/m³ for 104 weeks (Bucher et al. 1990, 1999; NTP 1991, 1998).

*Radioactive Cobalt.* No studies were located regarding ocular effects in humans or animals after inhalation exposure to radioactive cobalt compounds.

# **Body Weight Effects.**

*Stable Cobalt.* Weight loss, measured individually from time of initial examination throughout followup, was observed in a group of five diamond polishers suffering from cobalt-induced interstitial lung disease (Demedts et al. 1984b), but the exposure level of cobalt was not reported.

Decreased body weight, relative to controls at study termination, was reported in both rats and mice exposed to 19 mg cobalt/m³ as cobalt sulfate over 16 days or to 11.4 mg cobalt/m³ for 13 weeks (Bucher et al. 1990; NTP 1991). A 13-week exposure to 11.4 mg cobalt /m³ resulted in ruffled fur in male rats, with no clinical signs reported in female rats or either sex of mice (Bucher et al. 1990; NTP 1991). Chronic exposure of rats and mice to up to 1.14 mg cobalt/m³ did not result in decreased body weight (Bucher et al. 1999; NTP 1998).

Weight loss was found in dogs, but not rats or guinea pigs, exposed for 3 months to cobalt at a level of 9 mg cobalt/m³ as cobalt hydrocarbonyl (Palmes et al. 1959). Lifetime exposure of hamsters to a similar concentration (7.9 mg cobalt/m³ as cobalt oxide) did not result in decreased body weight gain (Wehner et al. 1977).

*Radioactive Cobalt.* No studies were located regarding body weight effects in humans or animals after inhalation exposure to radioactive cobalt compounds.

# 3.2.1.3 Immunological and Lymphoreticular Effects

Stable Cobalt. Cobalt is known to function as a hapten, resulting in the generation of antibodies against cobalt-protein complexes. Although the minimum exposure level associated with cobalt sensitization has not been determined, sensitization has been demonstrated in hard metal workers with work-related asthma who have experienced prolonged occupational exposure (>3 years) to levels ranging from 0.007 to 0.893 mg cobalt/m³ (Shirakawa et al. 1988, 1989). The lower end of this range, 0.007 mg/m³, is reported in Table 3-1 and plotted in Figure 3-1 as a LOAEL. The sensitization phenomenon includes the production of IgE and IgA antibodies to cobalt (Bencko et al. 1983; Shirakawa et al. 1988, 1989). Exposure to inhaled cobalt chloride aerosols can precipitate an asthmatic attack in sensitized individuals (Shirakawa et al. 1989), believed to be the result of an allergic reaction within the lungs.

Necrosis of the thymus was reported in rats exposed to 19 mg cobalt/m³ as cobalt sulfate over 16 days, and hyperplasia of the mediastinal lymph nodes was found in mice exposed to 11.4 mg cobalt/m³ for 13 weeks (Bucher et al. 1990; NTP 1991). Tests of immunological function, however, were not performed on the rats or mice.

*Radioactive Cobalt.* No studies were located regarding immunologic effects in humans or animals after inhalation exposure to radioactive cobalt compounds.

# 3.2.1.4 Neurological Effects

*Stable Cobalt.* Occupational exposure to cobalt in humans has been reported to cause several effects on the nervous system, including memory loss (Wechsler Memory Scale-Revised), nerve deafness, and a decreased visual acuity (Jordan et al. 1990; Meecham and Humphrey 1991). It should be noted, though, that both of these studies had small numbers of subjects (n=38 for Jordan et al. 1990, n=1 for Meecham and Humphrey 1991), and exposure characterization was not reported.

Congestion in the vessels of the brain/meninges was reported in rats and mice exposed to 19 mg cobalt/m³ or greater as cobalt sulfate over 16 days (Bucher et al. 1990; NTP 1991).

*Radioactive Cobalt.* No studies were located regarding neurological effects in humans or animals after inhalation exposure to radioactive cobalt compounds.

# 3.2.1.5 Reproductive Effects

*Stable Cobalt.* No studies were located regarding reproductive effects in humans after inhalation exposure to cobalt.

Testicular atrophy was reported in rats, but not in mice, exposed to 19 mg cobalt/m³ as cobalt sulfate over 16 days (Bucher et al. 1990; NTP 1991). Following exposure of mice to cobalt (as cobalt sulfate) for 13 weeks, a decrease in sperm motility was found at 1.14 mg cobalt/m³, and testicular atrophy was found at 11.4 mg cobalt/m³. A significant increase in the length of the estrous cycle was reported in female mice exposed to 11.4 mg cobalt/m³ for 13 weeks (Bucher et al. 1990; NTP 1991). No effects on the male or female reproductive systems were observed in rats similarly treated for 13 weeks (Bucher et al. 1990; NTP 1991), or in mice or rats exposed to up to 1.14 mg cobalt/m³ for 104 weeks (Bucher et al. 1999; NTP 1998).

*Radioactive Cobalt.* No studies were located regarding reproductive effects in humans or animals after inhalation exposure to radioactive cobalt compounds.

#### 3.2.1.6 Developmental Effects

*Stable Cobalt.* No studies were located regarding developmental effects in humans or animals after inhalation exposure to cobalt.

**Radioactive Cobalt.** No studies were located regarding developmental effects in humans or animals after inhalation exposure to radioactive cobalt compounds.

#### 3.2.1.7 Cancer

*Stable Cobalt.* The mortality of a cohort of 1,143 workers in a plant that refined and processed cobalt and sodium was analyzed (Mur et al. 1987); the French national population mortality data were used as a control. An increase in deaths due to lung cancer was found in workers exposed only to cobalt

(standardized mortality ratio (SMR) of 4.66; four cases in the exposed group versus one case in the controls). In a study within the cohort that controlled for date of birth, age at death, and smoking habits, 44% (four workers) in the group exposed to cobalt and 17% (three workers) in the group not exposed to cobalt died of lung cancer. The authors, however, indicated that the difference was not statistically significant and that the workers were exposed to both arsenic and nickel as well as cobalt. The nonneoplastic lung diseases commonly found in cobalt-exposed workers (see Section 3.2.1.2) were not reported in this group. These lung diseases may have been present in these workers, but if they were not listed as the cause of death on the death certificate, they would not have been mentioned. Inhalation was probably a prominent route of exposure to cobalt; however, oral and dermal exposure probably occurred as well. No adjustments were made for smoking habits, and the exposure levels of cobalt were not reported. A followup study of this cohort (Moulin et al. 1993) did not report significant increases in mortality due to respiratory or circulatory diseases. Similarly, no increase in the SMR for lung cancer was noted in exposed workers, relative to controls. While an elevated SMR for lung cancer was seen in maintenance workers (SMR=1.80, 95% confidence interval [CI]=0.78–3.55), it was not statistically significant.

Lasfargues et al. (1994) reported on the mortality of a cohort of 709 male workers in a French hard metal plant, using the national rates for French males for comparison. The overall mortality did not differ from expected, but there was a significant increase in mortality due to cancer of the trachea, bronchus, and lung (SMR=2.13, 95% CI=1.02–3.93). Smoking alone did not account for the lung cancer excesses, though smoking could not be entirely ruled out.

A cohort of 5,777 males and 1,682 females who were exposed occupationally to cobalt (concentrations ranging from 1 to 515 μg/m³, means of exposure levels ranging from 39.37 to 169.0 μg/m³) and tungsten carbide was examined by Moulin et al. (1998). A significantly increased mortality rate (SMR=1.30, 95% CI=1.00–1.66) was seen in exposed workers, when compared to the national average. Within this study, 61 cases and 180 controls were selected for a case-control study of cancer risk. When exposures during the last 10 years were ignored, a significant increase in lung cancer risk (SMR=1.93, 95% CI=1.03–3.62) relative to controls was seen among workers simultaneously exposed to cobalt and tungsten carbide. Adjustments for smoking and for coexposures to other carcinogens did not change the results, though occupational risk was greatest among smokers.

Wild et al. (2000) reported on a cohort of 2,216 male hard metal workers who had been employed for at least 3 months. The total mortality was not increased in workers, relative to local mortality rates. However, lung cancer mortality was significantly increased (SMR=1.70, 95% CI=1.24–2.26). The risks increased with exposure scores, even after adjustment for smoking and coexposure to other known or suspected carcinogens.

Treatment with 7.9 mg cobalt/m³ as cobalt oxide intermittently for a lifetime did not increase the incidence of malignant or benign tumors in hamsters (Wehner et al. 1977).

NTP (1998, Bucher et al. 1999) exposed groups of rats and mice of both sexes to 0, 0.11, 0.38, or 1.14 mg cobalt/m³ as cobalt sulfate for 2 years. Increased incidence of alveolar/bronchiolar neoplasms was noted following lifetime exposure of male rats to 1.14 mg cobalt/m³ and in female rats exposed to 0.38 mg cobalt/m³ (Bucher et al. 1999; NTP 1998). Statistical analysis revealed that tumors occurred in both sexes with significantly positive trends of rats. Similarly, mice of both sexes exposed to 1.14 mg cobalt/m³ showed an increase in alveolar/bronchiolar neoplasms, again with lung tumors occurring with significantly positive trends.

**Radioactive Cobalt.** No studies were located regarding carcinogenic effects in humans or animals after inhalation exposure to radioactive cobalt compounds.

### 3.2.2 Oral Exposure

#### 3.2.2.1 Death

Stable Cobalt. Lethal cardiomyopathy was reported in people who consumed large quantities of beer containing cobalt sulfate (Alexander 1969, 1972; Morin et al. 1971). The deaths occurred during the early to mid 1960s, at which time, breweries in Canada, the United States, and Europe were adding cobalt to beer as a foam stabilizer (Alexander 1969, 1972; Morin et al. 1971); this practice has been discontinued. Deaths occurred following ingestion of beer containing 0.04–0.14 mg cobalt/kg/day for a period of years (approximately 8–30 pints of beer each day). Approximately 43% of the patients admitted to the hospital with cardiomyopathy died within several years. "Acute mortality" (death within several days of admission) accounted for 18% of the deaths (Alexander 1972). It should be noted, however, that the cardiomyopathy may have also been due to the fact that the beer-drinkers had protein-poor diets and

may have had prior cardiac damage from alcohol abuse. Treatment of both pregnant and nonpregnant anemic patients with doses of cobalt (0.6–1 mg/kg/day) that were much higher than the doses in the beer did not result in effects on the heart (Davis and Fields 1958; Holly 1955). A 19-month-old male child who swallowed an unknown amount of a cobalt chloride solution died approximately 6.5 hours after ingestion, despite repeated induced vomiting, gastric lavage, and supportive therapy (Jacobziner and Raybin 1961).

Oral LD<sub>50</sub> values for several cobalt compounds have been determined in Wistar rats (FDRL 1984a, 1984b, 1984c; Singh and Junnarkar 1991; Speijers et al. 1982). The LD<sub>50</sub> values ranged from 42.4 mg cobalt/kg as cobalt chloride to 317 mg cobalt/kg as cobalt carbonate. An LD<sub>50</sub> of 3,672 mg cobalt/kg was also found for tricobalt tetraoxide, a highly insoluble cobalt compound (FDRL 1984c). The exact cause of death in rats is unknown, but effects on the heart, liver, gastrointestinal tract, and kidneys have been observed. In Sprague-Dawley rats, death has been reported to occur at 161 mg cobalt/kg given by gavage as cobalt chloride (Domingo and Llobet 1984). In male Swiss mice, the LD<sub>50</sub> values for cobalt chloride and cobalt sulfate have been reported to be 89.3 and 123 mg cobalt/kg, respectively (Singh and Junnarkar 1991).

Following 5 weeks of exposure to 20 mg cobalt/kg/day as cobalt sulfate by gavage, 20–25% of the guinea pigs died (Mohiuddin et al. 1970). The animals were given cobalt sulfate alone or in combination with ethanol (as part of a liquid diet) to compare the effects seen in animals to those seen in humans suffering from beer-cobalt cardiomyopathy. Although effects on the heart were found in the treated animals, alcohol did not appear to intensify the toxic effect.

The  $LD_{50}$  and all reliable LOAEL values for each species and duration category are reported in Table 3-2 and plotted in Figure 3-2.

*Radioactive Cobalt.* No studies were located regarding lethal effects in humans or animals after oral exposure to radioactive cobalt compounds.

Table 3-2. Levels of Significant Exposure to Cobalt - Chemical Toxicity Oral

		Exposure/				LOAEL		
Key to <sup>a</sup> figure		duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	Less serious (mg/kg/day)	Seric (mg/kg		Reference Chemical Form
	ACUTE I	XPOSURE						
	Death							
	Rat	1x				161.1		Domingo and Llobet 1984
	(Sprague- Dawley)	(GW)						Chloride
	Rat (Wistar)	1x (GW)				42.4	(LD50)	Singh and Junnarkar 1991
		(311)						Chloride
3	Rat (Wistar)	1x (GW)				194	(LD50)	Singh and Junnarkar 1991
		(311)						Sulfate
4	Rat	1 x				159	(LD50)	Speijers et al. 1982
	(Wistar)	(GO)						Oxide
5	Rat	1 x				168	(LD50)	Speijers et al. 1982
	(Wistar)	(GW)						Acetate
6	Rat	1 x				190	(LD50)	Speijers et al. 1982
	(Wistar)	(GW)						Chloride
7	Rat (Wistar)	1 x				140	(LD50)	Speijers et al. 1982
	(VVISIAI)	(GW)						Bromide
8	Rat (Wistar)	1 x			·.	161	(LD50)	Speijers et al. 1982
	(+*15141)	(GW)						Sulfate

Table 3-2. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Oral (continued)

		Exposure/ duration/				LOAE	<u>L</u>		
Key to <sup>a</sup> figure	Species (Strain)	frequency (Specific route)	System	NOAEL (mg/kg/day)		serious g/day)	Serio (mg/kg		Reference Chemical Form
9	Rat	1 x					91	(LD50)	Speijers et al. 1982
	(Wistar)	(GO)							Fluoride
	Rat	1 x					187	(LD50)	Speijers et al. 1982
	(Wistar)	(GO)							Phosphate
	Rat	1 x					109	(LD50)	Speijers et al. 1982
	(Wistar)	(GW)							Bromide
	Mouse	1x					123	(LD50)	Singh and Junnarkar
	(Swiss- Webster)	(GW)							1991
						•			Sulfate
13	Mouse (Swiss-	1x					89.3	(LD50)	Singh and Junnarkar
	Webster)	(GW)							1991 Chloride
	0 (11-								
	Systemic				4.0	( )			Roche and
14	Human	2 wk	Endocr		1.0	(decreased lodine uptake in thyroid)			Layrisse 1958
		(C)				apiane in injustry			Chloride
15	Rat	1x	Hemato		161.1	(increased hematocrit 8%) <sup>b</sup>			Domingo and Llobet 1984
		(GW)				070)			Chloride
16	Rat	1 x	Other	110	209	(Clinical signs, including			FDRL 1984a
		(GW)				decreased activity, ataxia, diarrhea, salivation)			Sulfate

Table 3-2. Levels of Significant Exposure to Cobalt - Chemical Toxicity Oral (continued)

		Exposure/ duration/		_					
Key to figure		frequency	System	NOAEL (mg/kg/day)		serious kg/day)	Serio (mg/kg		Reference Chemical Form
17	Rat	1 x	Other	•	149	(Decreased activity,			FDRL 1984b
	(Sprague- Dawley)	(GO)				diarrhea)			Carbonate
18	Rat	1x	Renal		19.4	(Increased urinary			Singh and Junnarkar
	(Wistar)	(GW)				output)			1991
									Sulfate
19	Rat	1 x	Cardio	109.6			176.6	(proliferative interstitial tissues	S, Speijers et al. 1982
	(Wistar)	(GO)						swollen muscle fibers, focal myocardial degeneration)	Fluoride
			Hepatic	42.6			68.2	(hyperemia)	
			Renal		42.6	(swollen proximal tubules)	176.6	(degeneration of proximal tubules)	
			Other				109.6	(hypothermia)	
20	Rat	1 x	Cardio				794.5	(hemorrhage)	Speijers et al. 1982
	(Wistar)	(GO)							Oxide
			Hepatic				157.3	(hyperemia)	
			Renal				157.3	(hyperemia)	
			Other				157.3	(hypothermia)	
21	Mouse	48 hr	Hemato		76.4 N	M (Alteration in electrophoretic profile of			Bryan and Bright, 1973
	(Swiss- Webster)	(VV)				serum proteins)			Chloride
22	Mouse	3 mo	Hemato	76.4 M					Bryan and Bright, 1973
	(Swiss- Webster)	(VV)							Chloride

Table 3-2. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Oral (continued)

		Exposure/ duration/		_		LOA	<b>NEL</b>	
Key to <sup>a</sup>	Species	frequency (Specific route)	System	NOAEL (mg/kg/day)		serious kg/day)	Serious (mg/kg/day)	Reference Chemical Form
	Neurolog	ical						
	Rat (Wistar)	1x (GW)			19.4	(Mild depression of spontaneous activity, muscle tone, and respiration)		Singh and Junnarkar 1991 Sulfate
	Rat (Wistar)	1x (GW)			4.25	(Mild depression of spontaneous activity, muscle tone, and respiration)		Singh and Junnarkar 1991 Chloride
	Developn	nental						
25	Rat	Gd 6-15 (GW)		24.8				Paternian et al. 1988 Chloride
26	Mouse	Gd 8-12 (GW)		81.7				Seidenberg 1986 Chloride

Table 3-2. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Oral (continued)

		Exposure/ duration/					Reference Chemical Form		
Key to		frequency (Specific route)	NOAEL System (mg/kg/day)		Less serious (mg/kg/day)			Serio (mg/kg	
	INTERM	EDIATE EXPO	SURE						
	Death								
27	Human	NR					0.04	(death)	Morin et al. 1971
		(W)							Sulfate
28	Gn Pig	5 wk					20	(death)	Mohiuddin et al. 1970
		(F)							Sulfate
	Systemic	<b>:</b>							
29	Human	NR	Cardio				0.07	(beer-cobalt cardiomyopathy)	Alexander 1972
		(W)							
									Sulfate
30	Human	1x/d 25 d	Hemato		1	(polycythemia) <sup>b.c</sup>			Davis and Fields 1958
		(C)							
31	Human	12-32 wk	Gastro		0.18	(nausea)			Duckham and Lee 1976b
		(C)							Chloride
			Hemato		0.18	(increased hemoglobin, 23-102% increase) <sup>b</sup>			
32	Human	90 d	Gastro		0.5	(gastric intolerance)			Holly 1955
		(C)	Llamata	0.6					Chloride
			Hemato Hepatic	0.6 0.6					

Table 3-2. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Oral (continued)

		Exposure/ duration/		_		LOAE	L		
Key to <sup>a</sup> figure		frequency (Specific route)	NOAEL System (mg/kg/day	NOAEL (mg/kg/day)		erious g/day)	Serio (mg/kg		Reference Chemical Form
33	Human	NR	Resp		0.04	(edema)			Morin et al. 1971
		(W)							Sulfate
			Cardio				0.04	(beer-cobalt cardiomyopathy)	1
			Gastro		0.04	(vomiting, nausea)			
			Hepatic		0.04	(necrosis)			
34	Human	10-25 d 1x/d	Other		0.54	(decreased lodine uptake)			Paley et al. 1958
		(C)							
35	Human	12-32 wk 7d/wk	Hemato		0.16	(increased hemoglobin) <sup>b</sup>			Taylor et al. 1977
		(C)							Chloride
36	Rat	4 wk	Bd Wt		3.79 N	(45-65% reduction in			Chetty et al.
	(Sprague- Dawley)	(F)				body weight gain)			1979 Chloride
37	Rat	8 wk 1x/d	Bd Wt		4.2	(33% decrease in body weight gain)			Clyne et al. 1988
		(F)							
38	Rat	3 mo	Resp		30.2	(increased lung weight 33%)			Domingo et al. 1984
		(VV)	Cardio		30.2	(increased heart weight 9.4%)			
			Gastro	30.2					
			Hemato		30.2	(increased hematocrit 29%) <sup>b</sup>			
			Musc/skel	30.2					
			Hepatic	30.2				•	
			Renal	30.2					

Table 3-2. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Oral (continued)

		Exposure/ duration/			LOAEL					
Cey to		frequency (Specific route)	System	NOAEL (mg/kg/day)	Less so (mg/kg			Seriou (mg/kg/	_	Reference Chemical Form
39	Rat	8 wk	Cardio					26	(degeneration)	Grice et al. 1969
		(F)								
40	Rat	24 wk	Cardio					8.4 M	(Left ventricular hypertrophy and impaired ventricular	Haga et al. 1996
	(Sprague- Dawley)	(F)							function)	Sulfate
41	Rat	4 mo	Resp	18						Holly 1955
		(G)		40						Chloride
			Cardio	18						
			Gastro	18						
			Hepatic	18				18	(tubular necrosis)	
		i	Renal					10	(tubulai necrosis)	
		·	Hemato		18	(erythrocytosis) <sup>b</sup>				
42	Rat	7 mo 6 d/wk	Hemato	0.05	0.5	(increased RBC, hemoglobin) <sup>b</sup>				Krasovskii and Fridlyand 1971
		(GW)	Hepatic	2.5						
43	Rat	3 wk	Cardio					12.4 M	(Incipient, multifocal	Morvai et al. 1993
	CFY	(G)							myocytolysis, with degeneration of myofibrilles)	Chloride
			Bd Wt		12.4 M	(Decreased body weigh 8%)	ht			
44	Rat	150 d 5 d/wk	Hemato		10	(increased hemoglobin	) <sub>p</sub>			Murdock 1959 Chloride
		(GW)	Hepatic		10	(increased weight 17%	)			
		(GVV)	Renal					10	(necrosis of tubular lining cells)	
			Bd Wt	10						

Table 3-2. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Oral (continued)

		Exposure/ duration/		_	L.	OAEL	_
Key to <sup>a</sup> figure	Openies	frequency (Specific route)	System	NOAEL (mg/kg/day)	Less serious (mg/kg/day)	Serious (mg/kg/day)	Reference Chemical Form
45		8 wk	Hemato	8.4 M			Pehrsson et al. 1991
	(Sprague- Dawley)						Sulfate
	24,		Bd Wt			8.4 M (>20% decrease from appropriate control)	
	Rat	12-16 d	Bd Wt	10.6 M			Saker et al. 1998
	(Sprague- Dawley)	(W)					Chloride
	<b></b> ,		Metab		10.6 M (Decreased serum glucose levels in diabetic rats, but not control rats)		
47	Rat	6 wk 7 d/wk	Hemato	0.6	2.5 (polycythemia) <sup>b</sup>		Stanley et al. 1947
		(C)					Chloride
48	Rat	3 d	Bd Wt	20 M	100 M (<20% reduction of bod weights)	у	Wellman et al. 1984
	(Long- Eva	<sup>ns)</sup> (F)			, , , , , , , , , , , , , , , , , , ,		Chloride
49	Mouse	45 d	Endocr			26 F (Necrosis and inflammation of thyroid)	Shrivastava et al. 1996
	Parkes	(W)				o, any, one,	Chloride
50	Gn Pig	5 wk	Cardio			20 (cardiomyopathy)	Mohiuddin et al. 1970
		(F)	Bd Wt	20			
51	Dog	4 wk 7 d/wk	Hemato		5 (polycythemia) <sup>b</sup>		Brewer 1940
		(F)					

Table 3-2. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Oral (continued)

		Exposure/ duration/		_		LOAEL			<del></del>
Key to <sup>a</sup>	Opcoico	frequency (Specific route)	System	NOAEL (mg/kg/day)	Less so (mg/ko		Seriou (mg/kg/		Reference Chemical Form
	Immunok	ogical/Lymphore	eticular						
52	Rat	4 wk					3.79 M	(Atrophy of the thymus)	Chetty et al. 1979
	(Sprague- Dawley)	(F)							Chloride
53	Rat	7 mo 6 d/wk		0.05	0.5	(decreased phagocytic ability)			Krasovskii and Fridlyand 1971 Chloride
		(GW)							Chloride
	Neurolog	jical							
54	Rat	57 d			20 M	(Increased latency during			Bourg et al. 1985
	(Sprague- Dawley)	(W)				retention testing)			Chloride
55	Rat	57 d			20	(increased reactivity)			Bourg et al. 1985
		(VV)							Chloride
56	Rat	7 mo 6 d/wk		0.05	0.5	(mildly increased latent reflex)	2.5	(pronounced increase in latent reflex)	Krasovskii and Fridlyand 1971
		(GW)							Chloride
57	Rat	30 d			4.96 M	(Alterations in			Mutafova- Yambolieva et
	(Wistar)	(VV)				sympathetically-induced contractility of vas			al. 1994
						deferens)			Chloride
58	Rat	69 d		5	20	(changes in schedule			Nation et al. 1983
		(F)				training, conditioned suppression, and mixed schedule training tests)			1905
59	Rat	30 d			6.44 M	(Alterations in cholinergic			Vassilev et al. 1993
	(Wistar)	(W)				sensitivity)			Nitrate
59		30 d			6.44 M	suppression, and mixed schedule training tests)			

Table 3-2. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Oral (continued)

		Exposure/ duration/		_	LOA	EL	
Key to <sup>®</sup> figure	Opcoico	frequency (Specific route)	System	NOAEL (mg/kg/day)	Less serious (mg/kg/day)	Serious (mg/kg/day)	Reference Chemical Form
	Rat	3 d		20 M	100 M (Saccharin and food aversion)		Wellman et al. 1984
	(Long- Evan	s) (F)			<b>,</b>		Chloride
	Reproduc	tive					
	Rat	98 days				20 M Pronounced histologic alteration of seminiferous	Corrier et al. 1985
	(Sprague- Dawley)	(F)				tubules	Chloride
62	Rat	90 d			30.2 M 26% decrease in		Domingo et al. 1984
	(Sprague- Dawley)	(W)			testicular weight		Chloride
63	Rat	98 d 7 d/wk				13.25 (testicular degeneration)	Mollenhauer et al. 1985
		(F)					
64	Rat	69 d		5		20 M (testicular atrophy)	Nation et al. 1983
		(F)					Chloride
65	Mouse	13 wk				43.4 M (Irreversible testicular degeneration)	Anderson et al. 1992
	(CD-1)	(W)				degeneration)	Chloride
66	Mouse	13 wk				43.4 M (Testicular degeneration)	Anderson et al. 1993
	(CD-1)	(W)					Chloride
67	Mouse	13 wk			23 (reversible testicular		Pedigo et al. 1988
		(W)			degeneration)		Chloride
68	Mouse	10 wk				58.9 M (Reduced pregnant females	Pedigo et al. 1993
	(B6C3F1)	(W)				and pups per litter; reduced fertility)	Chloride

Table 3-2. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Oral (continued)

	Exposure/ duration/		_				
ey to <sup>a</sup> Species figure (Strain)	cies frequency	System	NOAEL (mg/kg/day)	Less serious (mg/kg/day)	Serio (mg/kg		Reference Chemical Form
Deve	elopmental						
69 Hum	an 90 d		0.6				Holly 1955
	(C)						Chloride
70 Rat	Gd 14- Ld 21				5.4	(stunted pup growth)	Domingo et al. 1985
	(G)						Chloride

<sup>\*</sup>The number corresponds to entries in Figure 3-2.

Bd Wt = body weight; (C) = capsule; Cardio = cardiovascular; d = day(s); Derm = dermal; Endocr = endocrine; (F) = feed; F = female; (G) = gavage; Gd = gestation day; (GO) = gavage oil; (GW) = gavage-water; Gastro = gastrointestinal; Hemato = hematological; hr = hour(s); Ld = lactation day; LD<sub>30</sub> = dose producing 50% death; LOAEL = lowest-observed-adverse-effect level; M = male; Metab = metabolism; mo = month(s); NOAEL = no-observed- adverse-effect level; NS = not specified; (W) = drinking water; wk = week(s); x = times.

<sup>&</sup>lt;sup>b</sup>An increase in hemoglobin, hematocrit, or red blood cells is not necessarily considered an adverse effect.

<sup>&#</sup>x27;Used to derive an intermediate oral MRL; concentration was divided by an uncertainty factor of 100 (10 for use of a LOAEL and 10 for human variability), resulting n an MRL of 0.01 mg/kg/day

\*\*\*DRAFT FOR PUBLIC COMMENT\*\*\*

Figure 3-2. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Oral Acute (≤14 days)

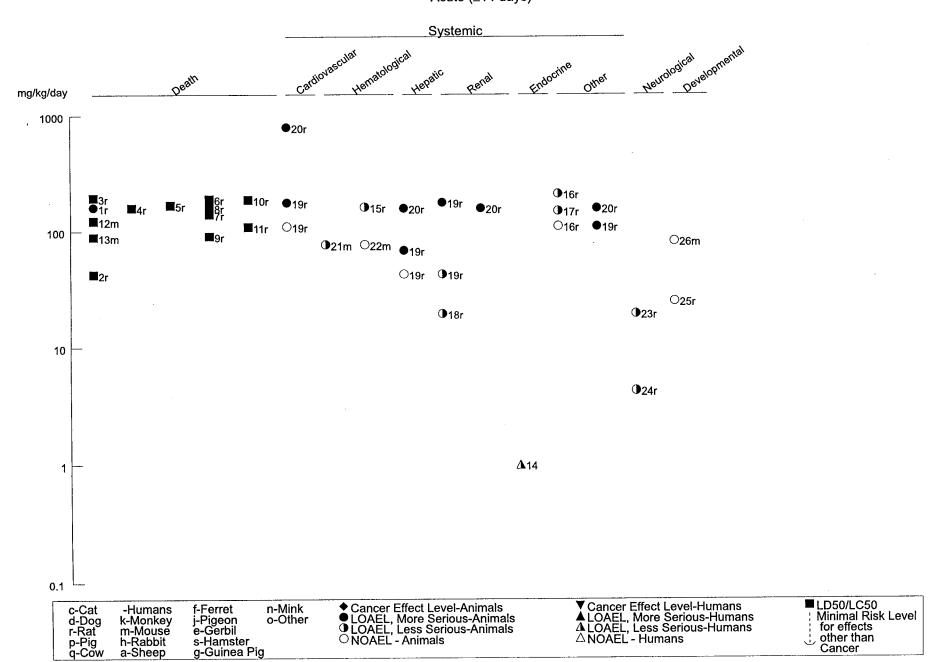


Figure 3-2. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Oral (continued)

Intermediate (15-364 days)

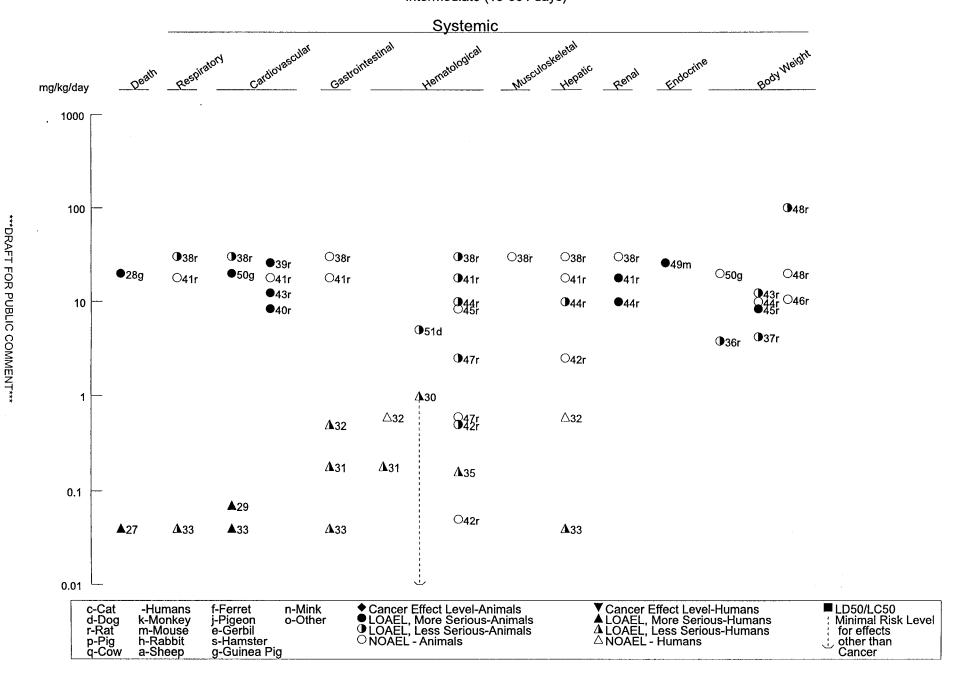
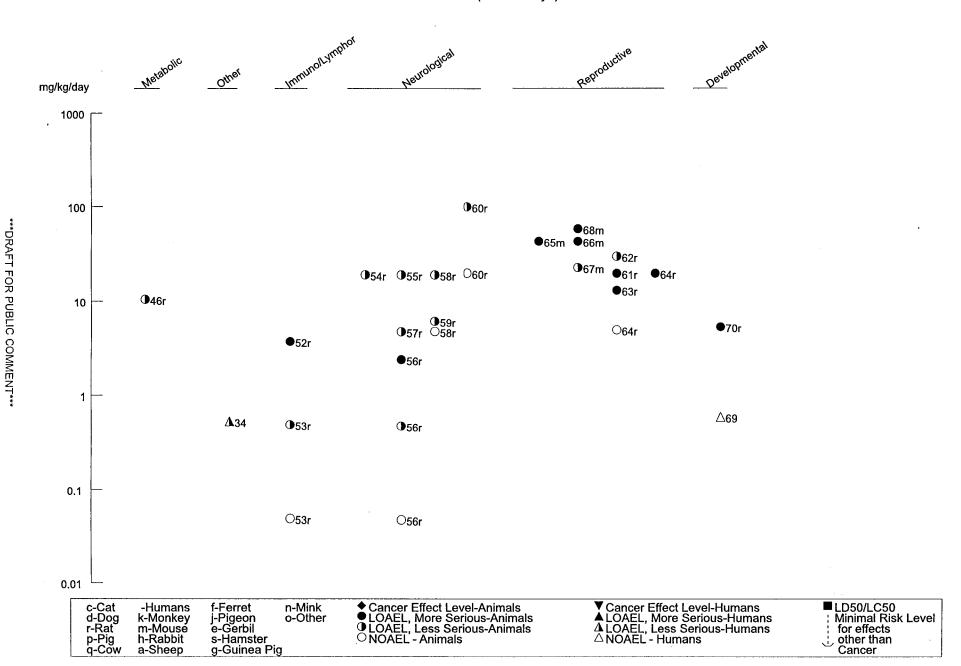


Figure 3-2. Levels of Significant Exposure to Cobalt - Chemical Toxicity - Oral (*continued*)

Intermediate (15-364 days)



## 3.2.2.2 Systemic Effects

Oral cobalt exposure in humans and/or animals resulted in respiratory, cardiovascular, gastrointestinal, hematological, musculoskeletal, hepatic, renal, ocular, thyroid, hypothermic, and body weight effects. For each effect, the highest NOAEL values and all reliable LOAEL values for each species and duration category are reported in Table 3-2 and plotted in Figure 3-2.

### Respiratory Effects.

Stable Cobalt. In 50 patients with beer-cobalt cardiomyopathy, pulmonary rales and pulmonary edema were observed and were attributed to cobalt-induced cardiac failure (Morin et al. 1971). These patients had ingested an average of 0.04 mg cobalt/kg/day in beer containing cobalt sulfate that was added to stabilize the foam. It should be noted that these patients consumed significant quantities of alcohol, and the effect that this may have had on the symptoms seen is not known. These symptoms may also be secondary to the cardiac effects, which are discussed below.

A significant increase in the weight of the lungs, without morphological or histological changes, was found in rats that received 30.2 mg cobalt/kg/day as cobalt chloride in drinking water for 3 months, as compared with controls (Domingo et al. 1984). No morphological changes were seen in the lungs of rats treated with 18 mg cobalt/kg/day for 4 months (Holly 1955).

*Radioactive Cobalt.* No studies were located regarding respiratory effects in humans or animals after oral exposure to radioactive cobalt compounds.

## Cardiovascular Effects.

Stable Cobalt. Beer-cobalt cardiomyopathy was observed in people who heavily consumed beer containing cobalt sulfate as a foam stabilizer (Alexander 1969, 1972; Morin et al. 1971). The beer drinkers ingested an average of 0.04 mg cobalt/kg/day (Morin et al. 1971, n=50) to 0.14 mg cobalt/kg/day for a period of years (Alexander 1969, 1972, n=28). The cardiomyopathy was characterized by sinus tachycardia, left ventricular failure, cardiogenic shock, diminished myocardial compliance, absence of a myocardial response to exercise or catecholamine, enlarged heart, pericardial effusion, and extensive intracellular changes (changes in the myofibers, mitochondria, glycogen, and lipids). The beer-cobalt

cardiomyopathy appeared to be similar to alcoholic cardiomyopathy and beriberi, but the onset of beer-cobalt cardiomyopathy was very abrupt. It should be noted, however, that the cardiomyopathy may have also been due to the fact that the beer-drinkers had protein-poor diets and may have had prior cardiac damage from alcohol abuse. Treatment of both pregnant and nonpregnant anemic patients for 90 days with doses of cobalt (0.6–1 mg/kg/day as cobalt chloride) that were much higher than the doses in the beer did not result in effects on the heart (Davis and Fields 1958; Holly 1955).

Approximately 40–50% of the patients admitted to the hospital with cardiomyopathy died within several years. In a followup study, 0–43% of the survivors showed a residual cardiac disability and 23–41% had abnormal electrocardiograms (Alexander 1972).

In an experiment designed to simulate conditions leading to beer-cobalt cardiomyopathy in humans, guinea pigs were given 20 mg cobalt/kg/day as cobalt sulfate by gavage either alone or in combination with ethanol (as part of a liquid diet) for 5 weeks (Mohiuddin et al. 1970). The experiment resulted in cardiomyopathy, which was characterized by abnormal EKGs; increased heart weights; lesions involving the pericardium, myocardium, and endocardium; and disfigured mitochondria. Alcohol did not intensify the cardiac effects. Myocardia changes (proliferative interstitial tissue, swollen muscle fibers, and focal degeneration) were also found in rats following a single dose of 176.6 mg cobalt/kg administered by gavage as cobalt fluoride or a single dose of 795 mg cobalt/kg administered as cobalt oxide (Speijers et al. 1982).

Three weeks of exposure to 12.4 mg cobalt/kg/day as cobalt chloride in male rats resulted in cardiac damage, presenting as incipient, multifocal myocytolysis, with degeneration of myofibrilles (Morvai et al. 1993). After longer-term exposure (2–3 months) of rats to 26–30.2 mg cobalt/kg/day as cobalt sulfate in the diet or as cobalt chloride in the drinking water, degenerative heart lesions (Grice et al. 1969) and an increase in heart weight were found (Domingo et al. 1984). Exposure of rats to 8.4 mg cobalt/kg/day as cobalt sulfate resulted in left ventricular hypertrophy and impaired left ventricular systolic and diastolic functions in an isolated working rat heart model (Haga et al. 1996).

**Radioactive Cobalt.** No studies were located regarding cardiovascular effects in humans or animals after oral exposure to radioactive cobalt compounds.

#### Gastrointestinal Effects.

Stable Cobalt. The first signs of the beer-cobalt cardiomyopathy syndrome were gastrointestinal effects and included nausea, vomiting, and diarrhea (Morin et al. 1971). Signs of heart failure subsequently appeared. These individuals had ingested an average of 0.04 mg cobalt/kg/day for a period of years during which cobalt sulfate was added to beer as a foam stabilizer; however, it is likely that alcohol consumption was also a factor.

In pregnant women given cobalt supplements (alone or combined with iron) to prevent the decrease in hematocrit and hemoglobin levels commonly found during pregnancy (n=78), a small percentage of those treated complained of gastric intolerance (Holly 1955). The women were treated with 0.5–0.6 mg cobalt/kg/day as cobalt chloride for 90 days. Similarly, nausea was reported in one anemic patient following treatment with 0.18 mg cobalt/kg/day as cobalt chloride (Duckham and Lee 1976b).

No morphological changes in the gastrointestinal system were observed following exposure of 20 male rats for 3 months to 30.2 mg cobalt/kg/day as cobalt chloride in the drinking water (Domingo et al. 1984) or exposure for 4 months to 18 mg cobalt/kg/day as cobalt chloride by gavage (Holly 1955).

**Radioactive Cobalt.** No studies were located regarding gastrointestinal effects in humans or animals after oral exposure to radioactive cobalt compounds.

### Hematological Effects.

Stable Cobalt. Cobalt has been shown to stimulate the production of red blood cells in humans. Davis and Fields (1958) exposed six apparently normal men, ages 20–47, to a daily dose of cobalt chloride, administered as a 2% solution diluted in either water or milk, for up to 22 days. Five of the six received 150 mg cobalt chloride per day for the entire exposure period, while the sixth was started on 120 mg/day and later increased to 150 mg/day. Blood samples were obtained daily from free-flowing punctures of fingertips at least 2 hours after eating, and at least 15 hours after the last dosage of cobalt. Blood was analyzed for red blood cell counts, hemoglobin percentage, leukocyte counts, reticulocyte percentages, and thrombocyte counts. Exposure to cobalt resulted in the development of polycythemia in all six subjects, with increases in red blood cell numbers ranging from 0.5 to 1.19 million (~16–20% increase above pre-treatment levels). Polycythemic erythrocyte counts returned to normal 9–15 days after

cessation of cobalt administration. Hemoglobin levels were also increased by cobalt treatment, though to a lesser extent than the erythrocyte values, with increases of 6–11% over pretreatment values. In five of the six subjects, reticulocyte levels were elevated, reaching at least twice the pre-experiment values. Thrombocyte and total leukocyte counts did not deviate significantly from pretreatment values. From the LOAEL of 1 mg/kg-day identified by this study, an intermediate-duration oral MRL of 1x10<sup>-2</sup> mg/kg-day was derived.

Increased levels of erythrocytes were also found following oral treatment of anephric patients (with resulting anemia) with 0.16–1.0 mg cobalt/kg/day daily as cobalt chloride for 3–32 weeks (Duckham and Lee 1976b; Taylor et al. 1977). The increase in hemoglobin resulted in a decreased need for blood transfusions. Treatment of pregnant women for 90 days with 0.5–0.6 mg cobalt/kg/day as cobalt chloride, however, did not prevent the reduction in hematocrit and hemoglobin levels often found during pregnancy (Holly 1955).

Significantly increased erythrocyte (polycythemia), hematocrit, and hemoglobin levels were found in animals treated orally with cobalt chloride as a single dose of 161 mg cobalt/kg (Domingo and Llobet 1984) or with longer-term exposure (3 weeks to 2 months) to 0.5 mg/kg/day and above (Brewer 1940; Davis 1937; Domingo et al. 1984; Holly 1955; Krasovskii and Fridlyand 1971; Murdock 1959; Stanley et al. 1947). Of particular note is an 8-week study in rats (Stanley et al. 1947), which reported dose- and time-related increases in erythrocyte number following oral administration of cobalt chloride, with an apparent NOAEL of 0.6 mg cobalt/kg/day and a LOAEL of 2.5 mg cobalt/kg/day. Changes in the levels of other blood proteins (transferrin, several haptoglobulins, and ceruloplasmin) were noted in male Swiss mice following 4, 24, and 48 hours of treatment with 76.4 mg cobalt/kg as cobalt chloride in the drinking water (Bryan and Bright 1973). Exposure for 3 weeks or 3 months to 76.4 mg cobalt/kg as cobalt chloride in the drinking water resulted in no alterations in serum proteins examined.

*Radioactive Cobalt.* No studies were located regarding hematological effects in humans or animals after oral exposure to radioactive cobalt compounds.

#### Musculoskeletal Effects.

*Stable Cobalt.* No studies were located regarding musculoskeletal effects in humans after oral exposure to cobalt.

No morphological changes were found in the skeletal muscle of rats exposed to 30.2 mg cobalt/kg/day as cobalt chloride in the drinking water for 3 months (Domingo et al. 1984). This NOAEL in rats for intermediate-duration exposure is reported in Table 3-2 and plotted in Figure 3-2.

**Radioactive Cobalt.** No studies were located regarding musculoskeletal effects in humans or animals after oral exposure to radioactive cobalt compounds.

# **Hepatic Effects.**

Stable Cobalt. Liver injury was evident in patients with beer-cobalt cardiomyopathy, characterized by central hepatic necrosis accompanied by increased levels of serum bilirubin and serum enzymes (serum glutamic oxaloacetic transaminase [SGOT], serum glutamic pyruvic transaminase [SGPT], lactate dehydrogenase [LDH]), creatine phosphokinase, ornithine carbamyl transferase, isocitric dehydrogenase, aldolase) (Alexander 1972; Morin et al. 1971). The hepatic injury may have resulted from ischemia, secondary to the cardiac effects of cobalt, and/or from excessive alcohol consumption. The cardiomyopathy resulted from the ingestion of beer containing 0.04 mg cobalt/kg/day as cobalt sulfate that had been added as a foam stabilizer (Morin et al. 1971). Liver function tests were found to be normal in pregnant women receiving up to 0.6 mg cobalt/kg/day as cobalt chloride for 90 days for treatment of the decreases in hematocrit and hemoglobin levels commonly found during pregnancy (Holly 1955).

Data from animals have also indicated that cobalt has hepatic effects. Hyperemia of the liver and cytoplasmic changes in hepatocytes (clumpy cytoplasm located along the cell membrane) were found in rats administered a single dose of 68.2 mg cobalt/kg as cobalt fluoride or a single dose of 157.3 mg cobalt/kg as cobalt oxide (Speijers et al. 1982).

Increased liver weight (17%) was found in rats exposed to 10 mg cobalt/kg/day (also as the chloride) for 5 months (Murdock 1959). No morphological or enzymatic changes were found in the livers of rats

exposed to 2.5–30.2 mg cobalt/kg as cobalt chloride by gavage or as cobalt chloride in the drinking water for 3–7 months (Domingo et al. 1984; Holly 1955; Krasovskii and Fridlyand 1971).

**Radioactive Cobalt.** No studies were located regarding hepatic effects in humans or animals after oral exposure to radioactive cobalt compounds.

#### Renal Effects.

Stable Cobalt. No studies were located regarding renal effects in humans after oral exposure to cobalt.

Acute and prolonged exposure to cobalt results in renal tubular degeneration in rats. Renal injury, evidenced by histologic alteration of the proximal tubules, was observed in rats after a single oral exposure to 42 mg cobalt/kg as cobalt fluoride (Speijers et al. 1982) and after exposure to 10–18 mg cobalt/kg/day as cobalt chloride for 4–5 months (Holly 1955; Murdock 1959). A slightly decreased urinary output was observed in rats exposed to 19.4 mg cobalt/kg as cobalt sulfate, but not in rats exposed to 4.25 mg cobalt/kg as cobalt chloride (Singh and Junnarker 1991).

**Radioactive Cobalt.** No studies were located regarding renal effects in humans or animals after oral exposure to radioactive cobalt compounds.

### **Dermal Effects.**

*Stable Cobalt.* No studies were located regarding dermal effects in humans or animals after oral exposure to cobalt.

**Radioactive Cobalt.** No studies were located regarding dermal effects in humans or animals after oral exposure to radioactive cobalt compounds.

# Ocular Effects.

*Stable Cobalt.* Severe visual disturbances (optic atrophy, impaired choroidal perfusion) developed in a man who was treated with cobalt chloride for pancytopenia and hypercellular bone marrow (Licht et al. 1972). He received 1.3 mg cobalt/kg daily for four series of treatments with a total duration of 6 weeks.

However, no other cases of visual disturbances due to therapeutic administration of cobalt have been reported, and no such effects have been observed in humans or animals.

**Radioactive Cobalt.** No studies were located regarding ocular effects in humans or animals after oral exposure to radioactive cobalt compounds.

## **Body Weight Effects.**

Stable Cobalt. No effects on body weight in animals were found following longer-term (1–5 months) exposure of rats to 10–30.2 mg cobalt/kg/day as cobalt chloride (Bourg et al. 1985; Domingo et al. 1984; Murdock 1959) or of guinea pigs to 20 mg cobalt/kg/day as cobalt sulfate (Mohiuddin et al. 1970). A significant decrease (33%) in body weight gain was observed following 8 weeks of exposure of rats to 4.2 mg cobalt/kg/day as cobalt sulfate (Clyne et al. 1988).

*Radioactive Cobalt.* No studies were located regarding body weight effects in humans or animals after oral exposure to radioactive cobalt compounds.

#### Metabolic Effects.

Stable Cobalt. Treatment of rats with 10.6 mg Co/kg/day as CoCl<sub>2</sub> in the drinking water for 12–16 days resulted in a significant decrease in serum glucose levels in diabetic rats, but not in control rats (Saker et al. 1998)

*Radioactive Cobalt.* No studies were located regarding metabolic effects in humans or animals after oral exposure to radioactive cobalt compounds.

# Other Systemic Effects.

Stable Cobalt. Hypothermia occurred in rats following a single oral dose of 157 mg cobalt/kg given as cobalt oxide or a single dose of 110 mg cobalt/kg given as cobalt fluoride (Speijers et al. 1982). The hypothermia was time- and dose-related. Hypothermia was reported as an effect during  $LD_{50}$  studies with other cobalt compounds, but the exact dose for the onset of hypothermia with these compounds was not

reported (Speijers et al. 1982). Other chemical signs noted in LD<sub>50</sub> studies include decreased activity, ataxia, diarrhea, and salivation (FDRL 1984a, 1984b).

**Radioactive Cobalt.** No studies were located regarding other systemic effects in humans or animals after oral exposure to radioactive cobalt compounds.

## 3.2.2.3 Immunological and Lymphoreticular Effects

Stable Cobalt. Cobalt is known to function as a hapten, resulting in the generation of antibodies against cobalt-protein complexes. Allergic dermatitis has been reported in some cobalt-sensitized people following oral challenge with cobalt. Several patients with eczema of the hands were challenged orally with 1 mg cobalt as cobalt sulfate given in tablet form once per week for 3 weeks (0.014 mg/kg/day). A flaring of the eczema was considered to be a positive allergic response to cobalt (Veien et al. 1987). Using both the oral challenge test and dermal patch tests, it was determined that the cobalt allergy was systemically induced. The exposure level associated with sensitization to cobalt was not established. After sensitization, allergic reactivity may be independent of dose. Cobalt has been found to be a sensitizer following inhalation exposure (Section 3.2.1.3). This LOAEL value was not reported in Table 3-2 because sensitized individuals only represent a small percent of the population.

A case report of a 6-year-old boy who had ingested approximately 1.7 mg of cobalt chloride reported neutropenia by 7 hours post-exposure (Mucklow et al. 1990). Thymic atrophy was reported in male Sprague-Dawley rats exposed to 3.79 mg cobalt/kg/day as cobalt chloride in the feed for 4 weeks (Chetty et al. 1979). A deterioration in immunological reactivity, manifested by a decline in phagocytic activity, was reported in rats following 6–7 months of treatment with 0.5 mg cobalt/kg and above as cobalt chloride (Krasovskii and Fridlyand 1971). This value is presented in Table 3-2 and Figure 3-2.

*Radioactive Cobalt.* No studies were located regarding immunological or lymphoreticular effects in humans or animals after oral exposure to radioactive cobalt compounds.

## 3.2.2.4 Neurological Effects

*Stable Cobalt.* No studies were located regarding neurological effects in humans after oral exposure to stable cobalt.

In Wistar rats, a single gavage of 4.25 mg cobalt/kg as cobalt chloride resulted in a moderate reduction in spontaneous activity, muscle tone, touch response, and respiration, while 19.4 mg cobalt/kg as cobalt sulfate caused a mild reduction the same parameters (Singh and Junnarkar 1991). In rats exposed to 4.96 mg cobalt/kg/day as cobalt chloride for 30 days in the drinking water, cobalt led to changes in sympathetically mediated contractile activity of isolated rat vas deferens (Mutafova-Yambolieva et al. 1994). Similarly, rats exposed to 6.44 mg cobalt/kg/day as cobalt nitrate in the drinking water showed an increased sensitivity and decreased maximal response to a cholinergic agonist (Vassilev et al. 1993). In rats exposed to 20 mg cobalt/kg/day as cobalt chloride for 57 days in the drinking water, cobalt enhanced behavioral reactivity to stress (the animals were less likely to descend from a safe platform to an electrified grid) (Bourg et al. 1985). Rats exposed to the same dose in the diet for 69 days showed a slower rate of lever pressing than controls but no change in behavioral reactivity to stress (Nation et al. 1983). Longer-term exposure of rats to cobalt chloride (7 months) resulted in a significant increase in the latent reflex period at 0.5 mg cobalt/kg and above as cobalt chloride, and a pronounced neurotropic effect (disturbed conditioned reflexes) at 2.5 mg cobalt/kg (Krasovskii and Fridlyand 1971).

The NOAEL value and the LOAEL value for rats for intermediate duration are reported in Table 3-2 and plotted in Figure 3-2.

**Radioactive Cobalt.** No studies were located regarding neurologic effects in humans or animals after oral exposure to radioactive cobalt compounds.

### 3.2.2.5 Reproductive Effects

*Stable Cobalt.* No studies were located regarding reproductive effects in humans after oral exposure to stable cobalt.

Testicular degeneration and atrophy have been reported in rats exposed to 13.25–58.9 mg cobalt/kg/day as cobalt chloride for 2–3 months in the diet or drinking water (Corrier et al. 1985; Domingo et al. 1984;

Mollenhauer et al. 1985; Nation et al. 1983; Pedigo et al. 1988, Pedigo and Vernon 1993), or in mice exposed to 43.4 mg cobalt/kg/day as cobalt chloride for 13 weeks in the drinking water (Anderson et al. 1992, 1993).

The highest NOAEL and all reliable LOAEL values for rats in the intermediate-duration category are reported in Table 3-2 and plotted in Figure 3-2.

*Radioactive Cobalt.* No studies were located regarding reproductive effects in humans or animals after oral exposure to radioactive cobalt compounds.

# 3.2.2.6 Developmental Effects

Stable Cobalt. No developmental effects on human fetuses were observed following treatment of pregnant women with cobalt chloride to raise hematocrit and hemoglobin levels that are often depressed during pregnancy. Dosages up to 0.6 mg cobalt/kg/day for 90 days were given (Holly 1955). Examination of the fetuses, however, was limited to the reporting of obvious birth defects, and exposure only occurred in the final trimester.

Oral exposure of female rats to cobalt chloride at 5.4 mg cobalt/kg/day or 21.8 mg cobalt/kg/day from gestation day 14 through lactation day 21 has been shown to result in stunted growth and decreased survival, respectively, of newborn pups (Domingo et al. 1985b). The effects on the offspring occurred at levels that also caused maternal toxicity (reduced body weight and food consumption and altered hematological measurements) and might be an indirect effect of maternal toxicity rather than a direct effect of cobalt on the fetus (Domingo et al. 1985b). Teratogenic effects were not observed. In contrast, no effects on fetal growth or survival were found following exposure of rats to 24.8 mg cobalt/kg/day as cobalt chloride during gestation days 6–15 (Paternian et al. 1988). In mice, exposure to 81.7 mg cobalt/kg/day as cobalt chloride during gestation days 8–12 was reported to have no effect on fetal growth or mortality in mice (Seidenberg et al. 1986).

The highest NOAEL and all reliable LOAEL values for each species and duration category are reported in Table 3-2 and plotted in Figure 3-2.

*Radioactive Cobalt.* No studies were located regarding developmental effects in humans or animals after oral exposure to radioactive cobalt compounds.

#### 3.2.2.7 Cancer

Stable Cobalt. In a survey assessing the correlation between cancer mortality and trace metals in water supplies (10 basins) throughout the United States, no correlation was found between cancer mortality and the level of cobalt in the water (Berg and Burbank 1972). Cobalt levels of 1–19  $\mu$ g/L, with resulting human exposures ranging from 0.03 to 0.54  $\mu$ g/kg/day, were reported.

No studies were located regarding carcinogenic effects in animals after oral exposure to stable cobalt.

*Radioactive Cobalt.* No studies were located regarding carcinogenic effects in humans or animals after oral exposure to radioactive cobalt compounds.

## 3.2.3 Dermal Exposure

### 3.2.3.1 Death

*Stable Cobalt.* No studies were located regarding lethal effects in humans after dermal exposure to cobalt.

No mortality was observed in guinea pigs dermally exposed to 51.75 mg cobalt/kg 5 days/week as dicobalt octacarbonyl for a total of 18 applications (Kincaid et al. 1954).

**Radioactive Cobalt.** No studies were located regarding lethal effects in humans or animals after dermal exposure to radioactive cobalt compounds.

# 3.2.3.2 Systemic Effects

No studies were located regarding respiratory, cardiovascular, gastrointestinal, hematological, musculoskeletal, hepatic, renal, or ocular effects in humans or animals after dermal exposure to stable or radioactive cobalt.

#### **Dermal Effects.**

Stable Cobalt. Dermatitis is a common result of dermal exposure to cobalt in humans which has been verified in a large number of studies (Alomar et al. 1985; Bedello et al. 1984; Dooms-Goossens et al. 1980; Fischer and Rystedt 1983; Kanerva et al. 1988, 1998; Marcussen 1963; Pryce and King 1990; Swennen et al. 1993; Romaguera et al. 1982; Valer et al. 1967). Using patch tests and intradermal injections, it has been demonstrated that the dermatitis is probably caused by an allergic reaction to cobalt. Contact allergy was reported in 22 of 223 (9.9%) nurses who were tested with a patch test of 1.0% cobalt chloride (Kie f-Đwierczy ka and Kr"cisz 2000). Exposure levels associated with the development of dermatitis have not been identified.

In animals, scabs and denuded areas were found after six doses of 51.75 mg cobalt/kg (5 days/week) as dicobalt octacarbonyl were applied to the shaved abdomens (uncovered area of approximately 50 cm<sup>2</sup>) of guinea pigs (Kincaid et al. 1954). By the 11<sup>th</sup> dose, the lesions disappeared. No adverse effects were observed in vehicle controls (methylethylketone). It is not known whether or not a similar reaction would result from metallic or inorganic forms of cobalt. This LOAEL value is reported in Table 3-3.

**Radioactive Cobalt.** No studies were located regarding dermal effects in humans or animals after dermal exposure to radioactive cobalt compounds.

### 3.2.3.3 Immunological and Lymphoreticular Effects

Stable Cobalt. Cobalt-induced dermatitis is well documented in the literature, and the studies indicate that cobalt is a sensitizer (Alomar et al. 1985; Dooms-Goossens et al. 1980; Fischer and Rystedt 1983; Goh et al. 1986; Kanerva et al. 1988; Marcussen 1963; Valer et al. 1967). Patch testing and intradermal injections were performed, but exposure levels of cobalt were not reported. Interrelationships exist between nickel and cobalt sensitization (Bencko et al. 1983; Rystedt and Fisher 1983). In guinea pigs, nickel and cobalt sensitization appear to be interrelated and mutually enhancing (Lammintausta et al. 1985), though cross-reactivity was not reported to occur.

Single or multiple dermal exposures of BALB/c mice to CoCl<sub>2</sub> in dimethylsulfoxide or in ethanol resulted in an increased cellular proliferation in the local lymph node assay in a concentration-dependant manner (Ikarashi et al. 1992a). The effect of three consecutive exposures to varying concentrations of

Table 3-3. Levels of Significant Exposure to Cobalt - Chemical Toxicity Dermal

	Exposure/				LOAEL		
Species	Duration/ Frequency	System	NOAEL (mg/kg/day)	Less sei (mg/kg/		Serious (mg/kg/day)	Reference Chemical Forn
ACUTE EXI	POSURE						
lmmunologi	cal/Lymphore	eticular					
Rat (Fischer- 344)	1x/d 3d		3.84 F	9.6 F	(Increased proliferation of lymphatic cells)		lkarashi et al. 1992b Chloride
Mouse (BALB/c)	1x or 1x/d for 3 d			10.8 F	(Increased proliferation of lymphatic cells)		Ikarashi et al. 1992a Chloride
Mouse CBA/N	1x/d 3 d		5.4 F	10.8 F	(Increased proliferation of lymphatic cells)		lkarashi et al. 1992b Chloride
Gn Pig (Hartley)	1x/d 3 d		7.39 F	14.7 F	(Increased proliferation of lymphatic cells)		lkarashi et al. 1992b Chloride
INTERMED	IATE EXPO	SURE					
Systemic					•		
Gn Pig (NS)	18 d 5 d/wk	Dermal		51.75	(skin lesions (scabs and denuded areas) at application site)		Kincaid et al. 1954

d = day(s); F = female; LOAEL = lowest-observed-adverse-effect level; NOAEL = no-observed- adverse-effect level; wk = week(s).

CoCl<sub>2</sub> in DMSO on lymph node proliferation was measured in rats, mice, and guinea pigs (Ikarashi et al. 1992b). Stimulation Indices of 3 or greater, indicated by the authors as a significant response, were reported for mice exposed to 1, 2.5, or 5% CoCl<sub>2</sub>, rats exposed to 2.5 or 5% CoCl<sub>2</sub>, and guinea pigs exposed to 5% CoCl<sub>2</sub>; these treatments resulted in dose levels of 10.8, 27, or 54.1 mg cobalt/kg/day for mice, 9.60 or 19.2 mg cobalt/kg/day for rats, and 14.7 mg cobalt/kg/day for guinea pigs.

No studies were located regarding the following health effects in humans or animals after dermal exposure to stable or radioactive cobalt:

- 3.2.3.4 Neurological Effects
- 3.2.3.5 Reproductive Effects
- 3.2.3.6 Developmental Effects

#### 3.2.3.7 Cancer

*Stable Cobalt.* No studies were located regarding carcinogenic effects in humans or animals after dermal exposure to cobalt.

*Radioactive Cobalt.* No studies were located regarding carcinogenic effects in humans or animals after dermal exposure to radioactive cobalt compounds.

# 3.2.4 External Exposure

This section contains information regarding health effects related to external exposure to radioactive cobalt sources. Radionuclides of cobalt may emit beta particles and/or gamma rays (see Table 3-4 and Figure 3-3), which are a health hazard in living organisms because they ionize the atoms that they hit while passing through the tissues of the body. Beta particles can travel appreciable distances in air, but travel only a few millimeters in solids. External exposure to beta particles may result in damage to skin and superficial body tissues at sufficiently high doses. Beta radiation is only a threat to internal organs if the radiation source is internalized. Gamma radiation, on the other hand, can easily pass completely through the human body and cause ionization of atoms in its path. The fraction of gamma rays that actually deposit energy and contributes to the radiation dose increases with tissue density (giving a higher dose to bone than soft tissue) and inversely with energy for most radionuclides of public interest. Several

Table 3-4. Levels of Significant Exposure to Cobalt - Radiation Toxicity - External

				LOAEL					
Key to <sup>a</sup> figure		Exposure/ duration/ frequency	NOAEL System (rad)		Less serious (rad)	Reference Chemical Form			
Δ	CUTE EXI	POSURE						·	
	)eath								
1	Human						2250 M	(Death)	Stavem et al. 1985
2	Mouse (BALB/c)						627	(30-day LD50 value, single exposure)	Darwezah et al. 1988
. 3	Mouse CBA/Ca.Lac	1x c.					1420 M	(Death)	Down et al. 1986
:	Systemic								
4	Human		Gastro	12.7					House et al. 1992
			Hemato	12.7					
5	Human		Dermal				159 N	1 (Severe alterations to skin o left hand)	f Klener et al. 1986
	·		Ocular				159 N	I (Progressive occlusion of vision of left eye)	
6	Human		Cardio				2250 N	// (left ventricular hypertrophy	) Stavem et al. 1985
			Gastro				2250 N	<ul> <li>(Pronounced atrophy in intestines; less severe in stomach)</li> </ul>	
			Hemato				2250 N	<ul> <li>(&gt;35% decrease in hemogolbin and &gt;90% decrease in thrombocytes)</li> </ul>	
			Renal		2250 M (Enlarged l	kidneys)			

Table 3-4. Levels of Significant Exposure to Cobalt - Radiation Toxicity - External (continued)

		F			LOAEL		
Key to		Exposure/ duration/ frequency	System	NOAEL (rad)	Less serious (rad)	Serious (rad)	Reference Chemical Form
7	Monkey (Rhesus)	30 min	Cardio		1000 M (Minor changes: increased heart rate; decreased blood pressure; variable cardiac output and total peripheral resistance)		Bruner 1977
8	Monkey (Rhesus)	1 hr	Cardio			10000 M (Pronounced decreases in mean arterial blood pressure and blood flow to the brain)	Cockerham et al. 1986
9	Rat (Wistar)		Cardio		2500 M (Increased brain uptake index)		Bezek et al. 1990
10	Rat (Sprague- Dawley)	1x	Resp			1500 M (Severe inflammation, pulmonary histopathology, fibrosis)	Lafuma et al. 1987
11	Mouse (Swiss- Webster)		Gastro		1000 M (Intestinal crypt cell damage, including necrosis and altered mitotic figures)		Devi et al. 1979
12	Mouse CBA/Ca.Lac.	1x	Resp		1330 M (Increased breathing rate)		Down et al. 1986
	Obi		Dermal		1800 M (Mild epilation)		
13	Mouse (Swiss- Webster)	24 hr	Hepatic		1000 M (Transient decrease in total liver protein)		Mazur et al. 1991

Table 3-4. Levels of Significant Exposure to Cobalt - Radiation Toxicity - External (continued)

		Fun course!							
Key to		Exposure/ duration/ frequency	NOAEL		Less serious (rad)		Seriou (rad	Reference Chemical Form	
14	Dog (Mongrel)	198 d	Cardio				4355	(Cardiac arrhythmia)	Dick et al. 1979
15.	Dog (Beagle)	10.44 min	Gastro		800	(Repeated emesis)			Gomez-de- Segura et al. 1998
16	Rabbit (New	1x	Dermal		1730 N	1 (Alopecia)			Cox et al. 1981
17	Zealand) Pig Large White	, 1x	Resp		900 F	Reversible decrease in ventilation capacity)	1090 F	(Irreversible decrease in ventilation capacity, histopathology, pulmonary atrophy)	Rezvani et al. 1989
18	Pig Large White	1x	Renal				874 F	(50% loss in effective renal plasma flow)	Robbins et al. 1989a
19	Pig Large White	1x	Renal		780 F	(Reversible changes in effective renal plasma flow and glomerular filtration rate)	980 F	(Persistent changes in effective renal plasma flow and glomerular filtration rate	Robbins et al. 1989b
			Hemato		780 i	(Slight decreases in erythrocytes, hemoglobin, and hematocrit)	1190 F	(Severe decreases in erythrocytes, hemoglobin, and hematocrit)	
20	Pig Large White	1x	Renal				557 F	(50% loss in effective renal plasma flow)	Robbins et al. 1989c
21	Pig Large White	1x	Renal				980 F	(Progressive inflammatory and degenerative changes the glomerulus)	Robbins et al. in 1991

Table 3-4. Levels of Significant Exposure to Cobalt - Radiation Toxicity - External (continued)

					LOAEL		
Key to		Exposure/ duration/ frequency	System	NOAEL (rad)	Less serious (rad)	Serious (rad)	Reference Chemical Form
	Ferret	2 hr	Gastro	49 M	77 M (Emesis with wretching)		King 1988
23	Baboon	3-4 wk, 1x/wk	Resp			3000 (Severe pulmonary fibrosis)	Collins et al. 1978
1	mmunologi	cal/Lymphore	eticular				
24	Human			12.7			House et al. 1992
25	Human				159 M (Minor reduction in white cell counts)		Klener et al. 1986
26	Human					2250 M (Pronounced decrease in lymphocytes and granulocytes)	Stavem et al. 1985
27	Mouse (Swiss- Webster)	24 hr			1000 M (>50% decrease in spleen weight and protein; increased spleen acid phosphatase)		Mazur et al. 1991
	Neurologica	al					
28	Rat (CD)	140 d		150 M	450 M (Reversible deficits in fixed-ratio behavior parameters)		Mele et al. 1988
29	Mouse (Swiss- Webster)	97 d		300 M	500 M (Reversible decreases in aggressive behavior)		Maier and Landauer 1989
30	Rabbit Burgundy fawn	12 hr			450 M (Altered firing rates and patterns of hippocampal neurons)		Bassant and Court 1978

Table 3-4. Levels of Significant Exposure to Cobalt - Radiation Toxicity - External (continued)

LOAEL									
Key to <sup>a</sup> figure	Species (strain)	Exposure/ duration/ frequency	System	NOAEL (rad)	Less serious (rad)		Seriou (rad		Reference Chemical Form
Reproductive									
	Rat (Sprague- Dawley)	1x			and a	reased testis weight altered matogenesis, with e evidence of very)			Cunningham and Huckins 1978
32	Rat (Wistar)	1x				rersible decrease in cular weight)			Laporte et al. 1985
D	)evelopme	ntal							
33	Monkey Squirrel	1x					100	(Developmental retardation, neurobehavioral deficits)	Brizzee et al. 1978
34	Rat (Sprague- Dawley)	1x					50 F	(Defective eye development and spinal curvature)	Bruni et al. 1994
35	Rat	1x					260	(Testicular trophy; adrenal atrophy)	Inano et al. 1989
36	Rat (Wistar)	1x			cyto	duced NADPH ochrome p450 uctase)			Inano et al. 1990
37	Rat (Wistar)	4d or 6d		11	(2.6	ghtly decreased 5%) brain weight in pring)	560	(decreased (13.1%) brain weight in offspring)	Reyners et al. 1992
38	Rat (Wistar)	1x					210 M	l (Testicular atrophy)	Suzuki et al. 1990

Table 3-4. Levels of Significant Exposure to Cobalt - Radiation Toxicity - External (continued)

		Exposure/				LOAEL			
Key to <sup>6</sup> figure		duration/ frequency	System	NOAEL (rad)	Less ser (rad)		Seriou (rad		Reference Chemical Form
39	Mouse (Swiss- Webster)	1x		5	10	(Decreased brain weight 3-4%, significantly increased microphthalmia)	50	(Increased fetal mortality and growth retardation)	Devi et al. 1994
40	Mouse (Swiss- Webster)	1x			25	(Decreased body weight 5%, liver weight 5%, and spleen weight 12%. Decreased spleen cellularity.)			Devi et al. 1998
41	Mouse (B6C3F1)	1x					100 M	(Increased number of tumor-bearing animals after in utero exposure)	Nitta et al. 1992
42	Mouse (Swiss- Webster)	1x					200	(Atrophy or lack of development of corpus callosum)	Schmidt and Lent 1987
43	Mouse	6d					20 F	(Altered neurobehavioral parameters, growth retardation)	Want et al. 1993
44	Mouse LACA	1x					50	(Delayed development, altered hindlimb splay)	Zhong et al. 1996
45	Hamster (Golden Syrian)	1x					200 F	(Severe developmental abnormalities of multiple organ systems, embryo death)	Harvey et al. 1962
46	Dog (Beagle)	1x					83	(Increased risk of thyroid neoplasia)	Benjamin et al 1997

Table 3-4. Levels of Significant Exposure to Cobalt - Radiation Toxicity - External (continued)

•					L	OAEL	
Key to	4 4 7	Exposure/ duration/ frequency	System	NOAEL (rad)	Less serious (rad)	Serious (rad)	Reference Chemical Form
47	Dog (Beagle)	1x		(, ad)		15.6 (Increased cancer-related mortality - multiple tumor types)	Benjamin et al. 1998b
48	Dog (Beagle)	1x		16	83 (Hypodontia)		Lee et al. 1989
49	Dog (Beagle)	1x				96 (Optic atrophy/degeneration	n) Schweitzer et al. 1987
(	Cancer						Ponjamin et al
50	Dog (Beagle)	1x				15.6 (Increased cancer-related mortality)	Benjamin et al. 1998b

Table 3-4. Levels of Significant Exposure to Cobalt - Radiation Toxicity - External (continued)

						LOAEL			
Key to	Species (strain)	Exposure/ duration/ frequency	System	NOAEL (rad)	Less serio	ous	Serio (rac		Reference Chemical Form
11	NTERMEDI	ATE EXPOS	JRE						
С	eath								Roscher and
51	Human						7500 F		Woodard 1969
5	Systemic								Fishman et al.
52	Human		Ocular				4800 F	(Progressive visual impairment and blindness)	1976
53	Human	22 - 35 d	Cardio				4623	(Persistent pericarditis)	Martin et al. 1975
54	Human	18 d	Gastro		3600	(Loose bowel movements, impaired absorption of vitamin B12)			McBrien 1973
55	Human	17 d	Dermal		4056 F	(Comedones, which were resolved with treatment)			Myskowski and Safai 1981
56	Human		Gastro				7500 F	(Severe gastrointestinal necrosis and fibrosis)	Roscher and Woodard 1969
57	Human	3 yr	Other (Hearing reduction)	2400					Thibadoux et al. 1980
58	Human	7 wk	Dermal		4700	(Reversible changes in skin pigmentation)			van Oort et al. 1984
59	Rat (albino)	10 wk	Other	2400 M	4800 N	(Transient alterations in incisor histopathology)	7200 I	(Lasting alterations in incison histopathology)	r Sweeney et al. 1977

		Table 3	-4. Levels of Significant Ex	posure to Cobalt - Radia	tion Toxicity-	External	(continued)
					LOAEL		
Key to <sup>a</sup>	Snecies	Exposure/	NOAEL	Less serious		Serie	ous

		<b></b>	-			LOAEL			
Key to		Exposure/ duration/ frequency	System	NOAEL (rad)	Less serious (rad)		Serious (rad)	•	Reference Chemical Form
60	Dog (Beagle)	150-300 d	Hemato				1125 M (	Aplastic anemia)	Seed et al. 1989
1	mmunologic	al/Lymphoret	icular						
61	Dog (Beagle)	150-300 d					1	Dose- and time-related eduction in granulocytes, monocytes, and ymphocytes)	Seed et al. 1989
!	Neurological						4000 =	Optic nerve damage,	Fishman et al.
62	Human						!	resulting in visual impairment and blindness)	
63	Human	9 mo				1	13150 F	(Neural necrosis and gliosis)	Liena et al. 1976
64	Human						5500 M	(Partial paralysis secondary to radiation myelopathy)	Sanyal et al. 1979
								(Partial paralysis secondary to radiation myelopathy)	
	Reproductiv	re							
65	Human	47 d					6600 M	(Calcification of the prostate	Keys and Reed 1980
66	Mouse	32 wk					1282 F	(Decreased offspring per litte and sterility)	er Searle et al. 1980

Table 3-4. Levels of Significant Exposure to Cobalt - Radiation Toxicity-	External	(continued)

				_			
	····	Exposure/				LOAEL	
Key to <sup>a</sup> figure	Species (strain)	duration/ frequency	System	NOAEL (rad)	Less serious (rad)	Serious (rad)	Reference Chemical Form
Са	ncer						
67 H	Human	NS				1800 F (Basal cell carcinon	na) Garcia-Silva et al. 1996
68 H	Human	8 mo				25150 M (Multiple basal cell carcinomas)	Wollenberg et al. 1995

ω
HEALT
HEFF

Eunocurol			L	OAEL	
Exposure/ duration/		NOAEL	Less serious	Serious	Reference
frequency	System				Chemical Form

#### **CHRONIC EXP**

Systemic

(strain)

69 Human

Key to Species

figure

3 yr

Cardio

13150 F (Endothelial hyperplasia, dysplasia, and fibrosis)

Liena et al.

1976

\*The number corresponds to entries in Figure 3-3.

Bd Wt = body weight; Cardio = cardiovascular; d = day(s); Endocr = endocrine; F = female; Gastro = gastrointestinal; Hemato = hematological; hr = hour(s); LOAEL = lowest-observed-adverse-effect level; M = male; Metab = metabolism; min = minute(s); mo = month(s); NOAEL = no-observed- adverse-effect level; NS = not specified; (occup) = occupational; Resp = respiratory; wk = week(s); yr = year(s).

Differences in levels of health effects and cancer effects between males and females are not indicated in Figure 3-3. Where such differences exist, only the levels of effect for the most sensitive gender are presented.

Figure 3-3. Levels of Significant Exposure to Cobalt - Radiation Toxicity - External Acute (≤14 days)

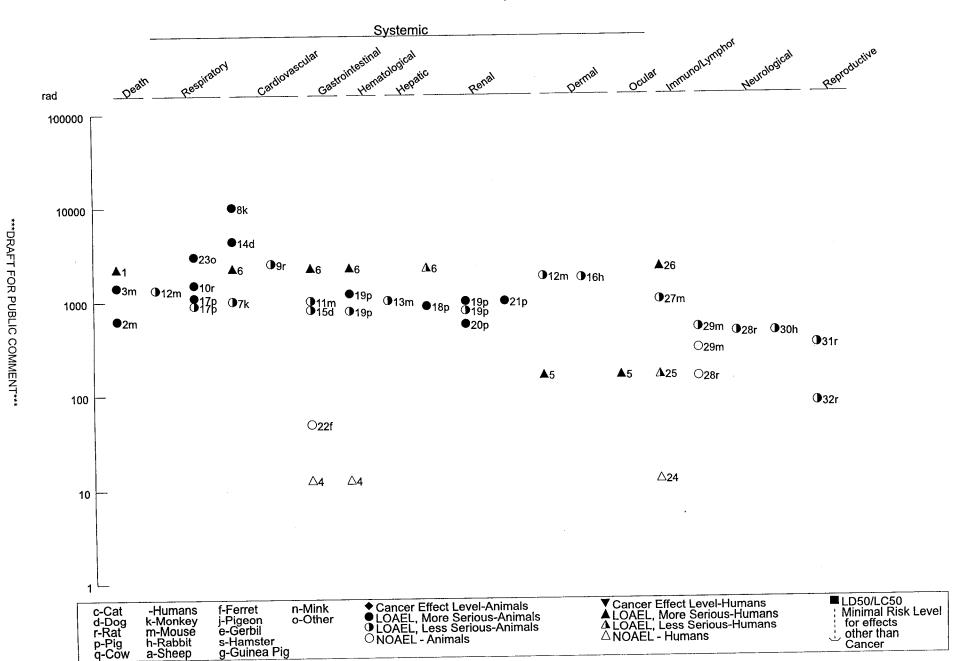


Figure 3-3. Levels of Significant Exposure to Cobalt - Radiation Toxicity - External (*continued*)

Acute (≤14 days)

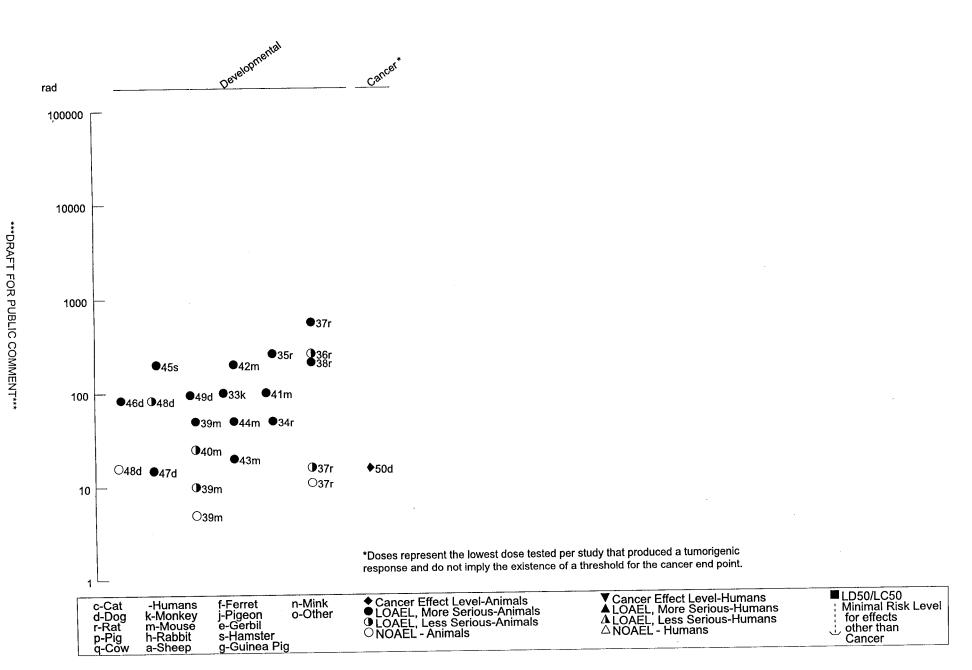
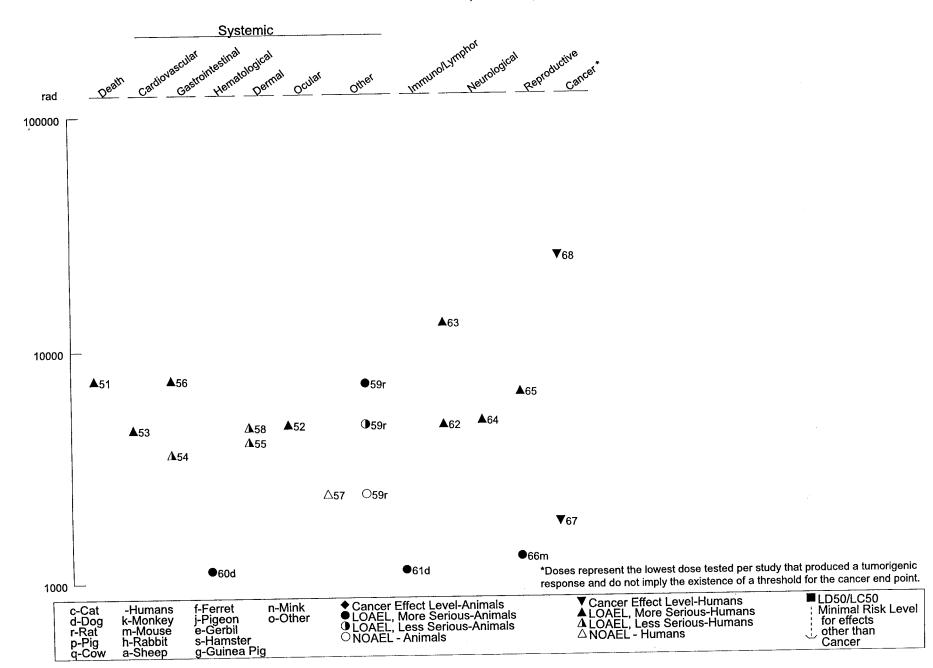


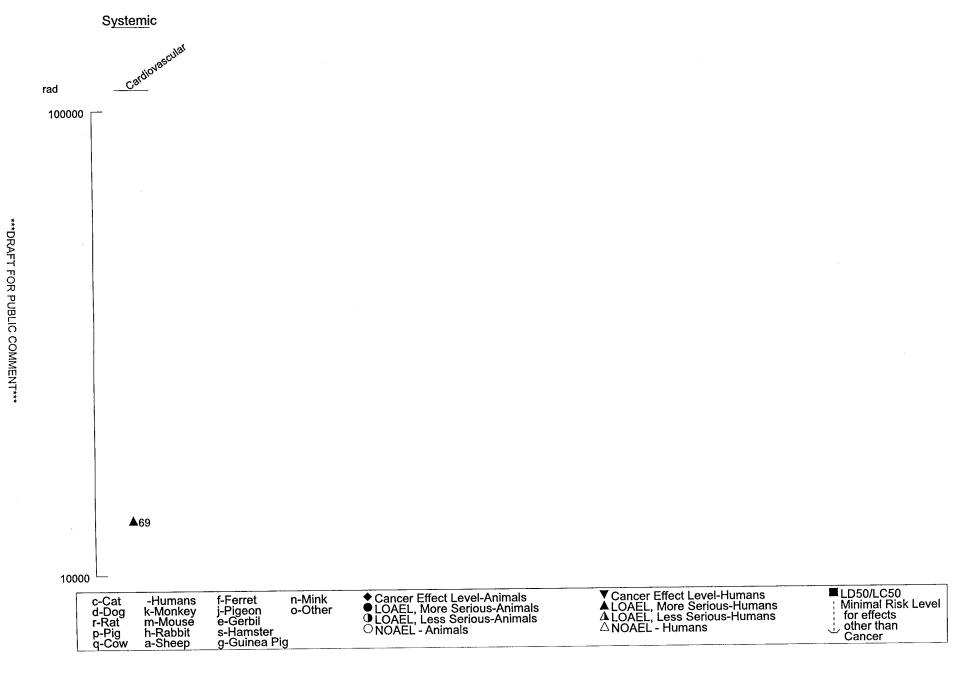
Figure 3-3. Levels of Significant Exposure to Cobalt - Radiation Toxicity - External (*continued*)

Intermediate (15-364 days)



\*\*\*DRAFT FOR PUBLIC COMMENT\*\*\*

Figure 3-3. Levels of Significant Exposure to Cobalt - Radiation Toxicity - External (continued) Chronic (≥365 days)



feet of concrete or a few inches of lead are typical shield thicknesses for protection from gamma rays. Because it is so highly penetrating, gamma radiation released by radionuclides such as cobalt is a radiation hazard to internal organs (ATSDR 1999; EPA 1997b). <sup>60</sup>Co gamma rays are commonly used for human radiotherapy.

The purpose of this section is to provide information regarding health effects associated with external exposure to a radioactive cobalt source. These health effects are not particular to cobalt, but apply to any radionuclide delivering the same beta and gamma radiation dose at a comparable dose rate. Refer to ATSDR (1999) for a detailed description of health effects from external exposure to ionizing radiation in general.

#### 3.2.4.1 Death

Exposure to high levels of external radiation, including radiation from cobalt radionuclides, may result in mortality. Stavem et al. (1985) reported a case in which a worker was exposed to 2,250 rad (22.5 Gy) within a few minutes time, resulting in death due to acute radiation sickness (depressed leukocyte counts, vomiting, diarrhea, etc.). Complications resulting from cobalt radiotherapy resulted in the death of a patient from severe gastrointestinal complications (Roschler and Woodard 1969).

Norris and Poole (1969) reported on the mortality of dogs exposed to  $^{60}$ Co gamma rays at a rate of 35 rad (0.35 Gy) per day for 40 days, resulting in a cumulative exposure of 1,400 rad (14 Gy). Twelve of 40 animals died prior to termination of the 40-day exposure period, 13 of 40 died within the 23-day post-exposure observation period, and 15 survived to the end of the study period, indicating an LD<sub>50</sub> of <1,400 rad at 35 rad/day. Darwezah et al. (1988) reported single whole-body exposure LD<sub>50</sub> values in mice of 913 rad (9.13 Gy) and 627 rad (6.27 Gy) at 6 and 30 days post-irradiation, respectfully. Down et al. (1986) reported a slightly higher LD<sub>50</sub> of 1,400–1,450 rad (14–14.5 Gy) for  $^{60}$ Co thoracic irradiation in mice at 26 days postirradiation. Several studies have demonstrated that decreasing the dose rate or the portion of the body exposed will increase the LD<sub>50</sub> for  $^{60}$ Co gamma rays (Darwezah et al. 1988; Down et al. 1986; Hanks et al. 1966; Page et al. 1968).

# 3.2.4.2 Systemic Effects

Respiratory Effects. Ionizing radiation is known to exert dramatic effects on the tissue of the lung (ATSDR 1999; Davis et al. 1992; Libshitz 1993; Roswit and White 1977), particularly at the high doses used in radiotherapy. The first phase of damage usually consists of radiation pneumonitis, which occurs between 3 and 13 weeks after irradiation and is characterized by low-grade fever, mild exertional dyspnea, congestion, and unproductive cough. The second phase is characterized by radiation-induced lung fibrosis, emphysema, and pleural thickening. Patients receiving radiotherapy treatment regimens of \$4,000 rad (40 Gy) to the chest region almost always develop radiographic changes in the lung (Davis et al. 1992), whereas lower therapeutic doses (2,500–3,000 rad, 25–30 Gy) generally result in a lower risk of adverse pulmonary symptoms (Davis et al. 1992; Roswit and White 1977). Prophylactic protective measures may be taken, and these injuries may be treated later if detected early enough in their progression (Roswit and White 1977).

At similar doses, studies in animals, including rats, mice, baboons, and pigs, using <sup>60</sup>Co radiation have also shown radiation pneumonitis and fibrosis, similar to effects seen in humans (Collins et al. 1978; Down et al. 1986; Lafuma et al. 1987; Rezvani et al. 1989). Other respiratory changes seen in animal experiments included an increased breathing rate, effects on the surfactant system, edema, increased pleural fluid content, pulmonary atrophy, and histologic alterations of the lung parenchyma (Bellet-Barthas et al. 1980; Collins et al. 1978; Down et al. 1986; Lafuma et al. 1987).

**Cardiovascular Effects.** Martin et al. (1975) reported that 24 of 81 patients who underwent <sup>60</sup>Co teletherapy for Hodgkin's disease, using an upper mantle treatment regimen of 4,000 rad (40 Gy) over 22–35 days, developed radiation-related pericarditis. In 14 of these patients, the condition was transient, while it persisted in the other 10 patients. Llena et al. (1976) presented a case wherein a 51-year-old woman who had received 13,150 rad (131.5 Gy) of <sup>60</sup>Co radiation between the nasopharynx and cervical lymph nodes as part of radiotherapy developed severe alterations in the endothelial cells of the brain, including proliferation, increased cytoplasmic organelles, and infoldings of the plasma membrane.

Whole-body exposure of Rhesus monkeys to 10,000 rad (100 Gy) over a 90-second period resulted in dramatic decreases in mean systemic arterial blood pressure, as well as in mean blood flow in the pons and pre-central gyrus, beginning at 10 minutes post-irradiation and persisting throughout the 60-minute observation period (Cockerham et al. 1986). Bruner (1977) examined cardiovascular parameters in

Rhesus monkeys exposed to 1,000 rad (10 Gy) at rates of 129–164 rad/minute (1.29–1.64 Gy/minute). Heart rate was elevated post-exposure, blood pressure was reduced near the end of exposure and thereafter, cardiac output increased at the end of exposure, but thereafter fell to below control levels, and total peripheral resistance decreased at early times post-exposure, but thereafter rose to above control levels. Ten of 12 dogs irradiated with 4,355–5,655 rad (43.6–56.6 Gy), focused on the interatrial septum of the heart, developed cardiac arrhythmias (Dick et al. 1979). The permeability of the blood-brain barrier was significantly increased, particularly for hydrophillic compounds, in rats exposed to 2,500 rad (25 Gy) from a <sup>60</sup>Co source (Bezek et al. 1990).

**Gastrointestinal Effects.** A worker accidentally exposed to an acute dose of 2,250 rad (22.5 Gy) showed slight atrophy of the stomach glands, marked atrophy in the small intestine, and total atrophy of the glands in the large intestine (Stavem et al. 1985). Two years after a woman received <sup>60</sup>Co radiation therapy amounting to 4,000 rads (40 Gy) anteriorly and 3,500 rads (35 Gy) posteriorly over a 6-week period, she reported severe gastrointestinal difficulties, including epigastric pain, vomiting, bloody stools, and weight loss (Roschler and Woodard 1969), eventually resulting in death. Autopsy revealed dense fibrous layers around the sacrum, with severe fibrosis confirmed by microscopic examination. Cobalt radiotherapy for carcinoma of the bladder (~3,100–3,600 rad, 31–36 Gy, over 18 days) resulted in loose bowel movements and a decreased absorption of vitamin B12 following oral exposure in 8 of 14 patients (McBrien 1973). No gastrointestinal symptoms were reported in three workers who were accidentally exposed to much lower exposure levels, ranging from 2.24 to 12.7 rad (0.022–0.127 Gy) (House et al. 1992).

Exposure of male Sprague-Dawley rats to 850 rad (8.5 Gy) of <sup>60</sup>Co gamma radiation resulted in marked alterations in drug absorption, primarily due to a decrease in gastric emptying rate (Brady and Hayton 1977b). Exposure of young adult beagle dogs to 800 rad (8 Gy) of <sup>60</sup>Co radiation at a rate of 177.5 rad/minute (1.775 Gy/minute) resulted in a 100% emesis rate within 10 hours post-irradiation, with an average of 2.4 episodes per animal and an average time to emesis of 82 minutes (Gomez-d-Segura et al. 1998). King (1988a) reported a NOAEL of 49 rad (0.49 Gy) and an EC<sub>50</sub> of 77 rad (0.77 Gy) for emesis and wretching following exposure of male ferrets to <sup>60</sup>Co gamma radiation. Exposure of male Swiss mice to 1,000 rad (10 Gy) of <sup>60</sup>Co radiation resulted in necrosis of the intestinal crypt cells (Devi et al. 1979).

Hematological Effects. No changes in hematologic parameters were reported in three workers who were accidentally exposed to levels ranging from 2.24 to 12.7 rad (0.022–0.127 Gy) (House et al. 1992). Hashimoto and Mitsuyasu (1967) reported that in 50 of 58 patients receiving local radiotherapy, irradiated bone marrow was more hypoplastic in the hematopoietic elements than in non-irradiated marrow in the same individual. A male worker exposed to 159 rad (1.59 Gy) showed minor reductions in leukocytes, neutrophils, and lymphocytes (Klener et al. 1986). Stavem et al. (1985) reported that a male worker exposed to 2,250 rad (22.5 Gy) showed a progressive decrease in hemoglobin and circulating thrombocytes prior to death. Autopsy showed a pronounced hypocellularity of the bone marrow.

Seed et al. (1989) exposed male Beagle dogs to 7.5 rad/day (0.075 Gy/day) gamma radiation for 150–700 days from a <sup>60</sup>Co source. The irradiated dogs initially showed a significant suppression, compared with levels from the control animals, of the five circulating types of cells studied (granulocytes, monocytes, platelets, erythrocytes, and lymphocytes), which lasted ~250 days; this was followed by a recovery phase for the remainder of the study period. Hashimoto and Mitsuyasu (1967) exposed guinea pigs to whole-body <sup>60</sup>Co radiation, and reported an initial hypoplasia of the bone marrow followed by recovery of hematopoietic activity by 3 weeks post-irradiation. Robbins et al. (1989b) reported significant reductions in erythrocyte count, hematocrit, and hemoglobin levels within 6–8 weeks of irradiation of the kidneys of female pigs with 980–1,400 rad (9.8–14 Gy) of <sup>60</sup>Co gamma rays.

**Musculoskeletal Effects.** No studies were located regarding musculoskeletal effects in humans or animals following external exposure to <sup>60</sup>Co radiation. These tissues are among the most radioresistant in both humans and animals.

**Hepatic Effects.** No studies were located regarding hepatic effects in humans following external exposure to <sup>60</sup>Co radiation.

No changes in liver weight were seen in male Swiss mice exposed to 1,000 rad (10 Gy) of <sup>60</sup>Co radiation and examined every 4 hours for 24 hours post-irradiation (Mazur et al. 1991). Andrzejewski et al. (1980) reported increased respiration rates in rat liver mitochondria after whole-body exposure to 1,000 or 3,000 rad (10 or 30 Gy) of <sup>60</sup>Co radiation; the increase was greater and more persistent at the higher exposure level.

**Renal Effects.** Stavem et al. (1985) reported that a 64-year-old man who accidentally received a fatal dose (2,250 rad) of cobalt radiation developed enlarged kidneys. No other studies were located regarding renal effects in humans after external exposure to cobalt radiation.

Robbins et al. (1989a, 1989b, 1989c, 1991a, 1991b) performed a series of studies in female White pigs wherein the kidneys of the animals were exposed to single doses of 780–1,400 rad (7.8–14 Gy) of <sup>60</sup>Co radiation and examined for periods up to 24 weeks postirradiation. Irradiation resulted in an initial increase in glomerular filtration rate (GFR), followed by a dose-related decrease in the GFR, beginning at 4 weeks postexposure. Effective renal plasma flow (ERPF) was also decreased in a dose-related manner beginning at 4 weeks postexposure, but did not show the initial increase seen in GFR. Some recovery of GFR and ERPF occurred by 24 weeks postirradiation, though values were still significantly reduced below controls in all groups but the 780 rad (7.8 Gy) group. Histology was performed on animals exposed to 980 rad (9.8 Gy) and killed between 2 and 24 weeks after exposure. Beginning at 2 weeks postirradiation, increased numbers of inflammatory cells were present within the glomerulus, and there was an increase in mesangial matrix and number of mesangial cells. The glomerular changes continued to progress in severity throughout the observation period, with generalized thickening of the capillary walls, extensive duplication of the basement membrane, and progressive inflammation. Tubular changes appeared to be maximal at 6 weeks, including focal degeneration and necrosis, with partial recovery at later timepoints.

**Dermal Effects.** Klener et al. (1986) described the accidental irradiation of a worker who was attempting to bring under control a sealed <sup>60</sup>Co source. The patient's left palm (the patient was left-handed) developed an irregular oval defect 3x4 cm with whitish edges and bleeding, as well as superficial lesions on the third and fourth finger. Considerable spontaneous pain required the administration of analgesics. The lesions showed no tendency to heal, instead spreading to the adjacent digits. After several failed skin graft attempts, the condition worsened, necessitating the amputation of fingers five through two. Walter (1980) reported that a patient who had undergone <sup>60</sup>Co radiotherapy (dose not reported) of the forehead and scalp developed a pronounced acneform reaction, characterized primarily by alopecia with multiple open comedones on the scalp and forehead, and hair loss. With treatment, the comedones were 80% cleared at 9 months post-diagnosis (13 months post-treatment), but no hair regrowth was noted. Myskowski and Safai (1981) have likewise reported localized comedones in a patient following 4,056 rad (40.6 Gy) of <sup>60</sup>Co radiotherapy. Van Oort et al. (1984) reported that patients receiving 4,700–6,000 rad (47–60 Gy) of <sup>60</sup>Co radiotherapy over a 7-week period showed significant

differences in baseline color of the skin, primarily erythema and pigmentation, beginning the third week of exposure and persisting throughout the fifth week postirradiation (study week 12).

Cox et al. (1981) reported a dose-related loss of hair in rabbits exposed to 1,730–3,210 rad (17.3–32.1 Gy) <sup>60</sup>Co gamma rays, with recovery initially noted in animals exposed to 2,140 rad (<21.4 Gy) by day 200 postirradiation. Beginning at day 500 postirradiation, a substantial loss of hair again was seen, persisting throughout the end of the study. Mice exposed to 1,800 rad (18 Gy) of <sup>60</sup>Co radiation showed a slight increase in epilation score (Down et al. 1986).

**Ocular Effects.** Augsburger and Shields (1985) described 13 patients who developed cataracts following <sup>60</sup>Co plaque radiotherapy; estimated doses to the eyes ranged from 2,000 to 10,000 rad (20–100 Gy). Fishman et al. (1976) reported on two patients who received head-only <sup>60</sup>Co radiotherapy, in combination with chemotherapy, for the treatment of acute lymphocytic leukemia. Both patients, who received 2,400 rads (24 Gy) over an initial 16 day course of treatment followed later by either 2,400 or 2,500 rads (24 or 25 Gy) in followup therapy, developed progressively severe vision disorders, resulting in partial or total blindness. Exposure of a male worker to a whole-body dose of 159 rad (1.59 Gy) of <sup>60</sup>Co radiation resulted in a progressive deterioration of visual acuity, due to cataract development, in the left eye (which was more exposed than the right) over time (Klener et al. 1986).

**Other Systemic Effects.** Thibadoux et al. (1980) reported that of 61 children receiving a course of 2,400 rad (24 Gy) of cranial radiotherapy, none developed significant reductions in hearing levels by the end of the third year after irradiation.

Sweeney et al. (1977) examined the effects of <sup>60</sup>Co radiation on the teeth of rats exposed to 0, 2,400, 4,800, or 7,200 rads (0, 24, 48, or 72 Gy). Animals exposed to 4,800 rad (48 Gy) showed transient effects on the incisors only, while at 7,200 rad (72 Gy) the effects lasted throughout the 10-week study period.

# 3.2.4.3 Immunological and Lymphoreticular Effects

A worker accidentally exposed to an acute dose of 2,250 rad (22.5 Gy) showed a rapid fall in circulating lymphocytes and granulocytes prior to death (Stavem et al. 1985). Chronic exposure to low amounts of <sup>60</sup>Co radiation in people living in a contaminated building significantly reduced the numbers of circulating

CD4<sup>+</sup> lymphocytes in the blood (Chang et al. 1997, 1999b); mean total radiation dose was estimated to be 0.169 Gy (16.9 rad) over a 2–13-year period. Similarly, children chronically-exposed to low levels (estimated dose of 0.002–0.085 Gy [0.2–8.5 rad]) of <sup>60</sup>Co radiation in a contaminated kindergarten building showed significant decreases in total leucocytes and neutrophils, but an increase in eosinophils, 5–7 years after exposure had ceased (Chang et al. 1999a).

In male Swiss mice exposed to 1,000 rad (10 Gy) of <sup>60</sup>Co radiation, significant decreases in weight of the spleen were seen as early as 1 hour post-exposure, and persisting throughout the following 24 hours (Mazur et al. 1991). Spleen acid phosphatase activity, expressed as activity per gram of protein, was significantly increased in irradiated animals beginning at 13 hours post-exposure.

# 3.2.4.4 Neurological Effects

Llena et al. (1976) presented a case wherein a 51-year-old woman who had received 13,150 rad (131.5 Gy) of <sup>60</sup>Co radiation between the nasopharynx and cervical lymph nodes as part of radiotherapy developed focal necrosis of the brain in the frontal lobe, as confirmed by gross and microscopic examination. Fishman et al. (1976) reported on two patients who received head-only <sup>60</sup>Co radiotherapy, in combination with chemotherapy, for the treatment of acute lymphocytic leukemia. Both patients, who received 2,400 rad (24 Gy) over an initial 16-day course of treatment followed later by either 2,400 or 2,500 rad (24 or 25 Gy) in followup therapy, developed progressively severe vision disorders, resulting in partial or total blindness. Histopathology from one patient demonstrated severe alterations in the optic nerve, including severe atrophy, terminal beading, lack of myelin, and calcification. Sanyal et al. (1979) reported on five patients, exposed to 4,500–6,000 rad (45–60 Gy) <sup>60</sup>Co radiation as radiotherapy, who developed varying degrees of myelopathy, resulting in minimal to mild paralysis.

Mele et al. (1988) exposed male rats to 50, 150, or 450 rad (0.5, 1.5, or 4.5 Gy) 3 times, separated by 43-day intervals, and examined for changes in behavior daily for 30 days following each exposure. Rats exposed to 450 rad (4.5 Gy), but not those exposed to 150 rad (1.5 Gy) or 50 rad (0.5 Gy), showed significant deficits in fixed-ratio response rates and running rates after each exposure, beginning the day after exposure and persisting for 4–5 days, after which both rates returned to normal. After the third exposure, all rats were exposed to 650 rad (6.5 Gy), which resulted in similar performance decrements as were seen in the 450 rad (4.5 Gy) animals, again beginning 24 hours after exposure, with previous exposure resulting in no differences in behavioral parameters. Maier and Landauer (1989) reported

significant decreases in offensive behavior in mice acutely exposed to 500 or 700 rad (5 or 7 Gy), but not those exposed to 300 rad (3 Gy), with changes occurring in the second week postirradiation and responses returning to normal by day 19 postirradiation. Bassant and Court (1978) reported that rabbits exposed to 450 rad (4.5 Gy) of <sup>60</sup>Co radiation whole-body showed an altered activity of hippocampal cells, with a slowed mean discharge rate and increased interspike variability persisting for at least 12 hours postirradiation.

### 3.2.4.5 Reproductive Effects

Ionizing radiation in general, and gamma-emitting isotopes in particular, is known to have profound effects on reproductive tissues, with effects seen primarily in rapidly-dividing germ cells resulting in temporary or permanent sterility in both sexes, as well as other effects (ATSDR 1999). These effects are usually observed only at high radiation doses. Keys and Reed (1980) reported a case of a man who, as treatment for a prostate tumor, received an estimated dose of 6,600 rad (66 Gy) to the prostate over a 47-day period, and who later developed a severe prostatic calcification necessitating surgical correction.

<sup>60</sup>Co radiation at high doses has been shown to elicit profound decrements in reproductive ability in animal species. Whole-body acute exposure of rats to 330 rad (3.3 Gy) decreased testicular weights beginning at 22 days postirradiation, with recovery of testicular weight beginning about day 65 (Cunningham and Huckins 1978). Histologic examination of the testes revealed destruction of the spermatogonial population, with a slow recovery as the spermatogonial population was rebuilt from the surviving stem cells. Searl et al. (1976) reported that exposure of male mice to 1,128 rad (11.3 Gy) over a 28-week period resulted in significant reductions of testis mass and epididymal sperm count. Male Wistar rats exposed to a single dose of 80 rad (0.8 Gy) of testicular radiation showed increased tubular fluid production and decreased testicular weight at 30 and 45 days postirradiation, but not at later time points (Laporte et al. 1985). Single doses of >100 rad (1 Gy) of <sup>60</sup>Co radiation caused decreased fertility in exposed female mice (Philippe 1975). Continuous exposure of female mice to an average daily dose of 8 or 16 rad/day (0.08 or 0.16 Gy/day) caused a decreased number of offspring per litter and decreased reproductive performance, with 100% sterility occurring at 32 weeks of exposure at 8 rad/day (0.08 Gy/day) or 20 weeks of exposure at 16 rad/day (0.16 Gy/day) (Searl et al. 1980). Female rabbits exposed to 400 rad (4 Gy) prior to implantation showed dramatic decreases in implantation (Chang et al. 1963).

#### 3.2.4.6 Developmental Effects

No studies were located regarding developmental effects in humans after external exposure to cobalt radiation.

In utero exposure to cobalt radiation has been extensively studied in animal species, and may elicit substantial effects across many organ systems of the developing organism. Effects have been noted following single-dose exposures as low as 10 rad (0.10 Gy) in mice (Devi et al. 1994; Wang et al. 1993), 50 rad (0.5 Gy) in rats (Bruni et al. 1994), 200 rad (2 Gy) in hamsters (Harvey and Chang 1962), 250 rad in rabbits (Chang et al. 1963), 15.6 rad (0.16 Gy) in dogs (Benjamin et al. 1998a, 1998b), and 100 rad (1 Gy) in monkeys (Brizzee et al. 1978). Organs known to be affected include the brain (Brizzee et al. 1978; Bruni et al. 1994; Devi et al. 1994; Hamilton et al. 1989; Reyners et al. 1992; Schmidt and Lent 1987), eyes (Brizzee et al. 1978; Bruni et al. 1994; Schweitzer et al. 1987), hair (Hirobe 1994; Hirobe and Zhou 1990), kidney (Benjamin et al. 1998a; Brizzee et al. 1978), liver (Devi et al. 1998), ovaries (Inano et al. 1989), pituitary (Brizzee et al. 1978), skeleton (including cleft palate, shortened digits, fused digits, and other gross abnormalities) (Bruni et al. 1994; Chang et al. 1963; Harvey and Chang 1962), spleen (Devi et al. 1998), teeth (Lee et al. 1989), testes (Inano et al. 1989; Suzuki et al. 1990), and thyroid (Benjamin et al. 1997). <sup>60</sup>Co radiation *in utero* has also shown to cause functional alterations, including postnatal growth retardation (Wang et al. 1993; Zhong et al. 1996), neurobehavioral changes (Brizzee et al. 1978; Wang et al. 1993), hormonal production (Brizzee et al. 1978; Inano et al. 1989; Suzuki et al. 1990), alterations in hepatic enzymes (Inano et al. 1990), and diabetes mellitus (Benjamin et al. 1998a). *In utero* irradiation with cobalt also leads to increased tumor incidence later in life (Benjamin et al. 1991, 1997, 1998b; Nitta et al. 1992).

Devi et al. (1994) exposed pregnant mice to a single dose of 0–50 rad (0–0.50 Gy) of <sup>60</sup>Co radiation on day 11.5 of gestation. A significant decrease in pup brain weight and increase in the incidence of microphthalmia was seen at 10 rad (0.10 Gy), with decreases in head width, head length, body length, and body weight occurring at higher doses. A later study (Devi et al. 1998) found decreases in body weight, liver weight, and spleen weight in pups 72 hours after irradiation with 25 rad (0.25 Gy) of <sup>60</sup>Co radiation on day 17 of gestation. Male offspring, but not female offspring, of mice exposed to 50 rad (0.5 Gy) on gestation day 9 showed decreased body weights on postnatal days 0, 3, and 7, while offspring of both sexes showed delays in pinna detachment, incisor eruption, eye opening, and testes descent (Zhong et al. 1996). Wang et al. (1993) reported that mice exposed to a cumulative *in utero* dose of 10 rad (0.10 Gy)

showed alterations in visual placing reflex tests, while those exposed to 20 or 40 rad (0.20 or 0.40 Gy) showed decreased mean body weight, delayed eye opening, and alterations in the air righting reflex.

Rats exposed to 50 rad (0.50 Gy) of <sup>60</sup>Co radiation on gestational day 9.5 showed histologic damage to the neuro-epithelium 4 hours post-exposure, with abnormal flexion of the embryo and abnormal flexion of the head at 48 hours post-exposure (Bruni et al. 1994). At birth, rats showed increased incidence of defective eye development, spinal curvature, and visceral anomalies. Reyners et al. (1992) reported decreased brain weight in 3-month-old rats that had been exposed to cumulative doses of 160 rad (1.6 Gy) over gestation days 12–16 or 170 rad (1.7 Gy) over gestation days 14–20. Male rats exposed to 210 rad (2.1 Gy) on day 20 of gestation showed atrophy of the testes, prostates, and seminal vesicles, as well as a complete disappearance of germinal cells within the testes, on postnatal day 70 (Suzuki et al. 1990). Inano et al. (1989) exposed rats on gestation day 20 to 260 rad (2.6 Gy) of <sup>60</sup>Co radiation. Seminiferous tubules of male offspring and ovaries of female offspring showed pronounced atrophy, and steroid hormone production was significantly altered.

Benjamin et al. (1997, 1998a, 1998b) exposed groups of pregnant Beagle dogs to 15.6–17.5 or 80.8–88.3 rad (0.15–0.175 or 0.8–0.88 Gy) of <sup>60</sup>Co radiation on day 8, 28, or 55 post-breeding. Animals were allowed to live their full life span and were observed for radiation-related illnesses and cause of death. No change in the mean age at death was seen as a result of exposure. Males exposed to either exposure level at day 55 post-breeding, but not females at any time or males exposed at days 8 or 28, showed an increase in deaths due to renal disease. High-dose females exposed on days 28 or 55 showed an increase in the frequency of diabetes mellitus. Both sexes showed an increase in malignant neoplasias in general when exposed to radiation at 8 or 55 days postcoitus, but not at 28 days, while females exposed on day 55 also showed an increase in lymphoid neoplasia. A similar exposure on day 28 or 55 postcoitus also resulted in a dose-dependent decrease in brain weight (Hamilton et al. 1989). *In utero* radiation of dogs to higher doses (100–380 rad [1–3.8 Gy]) resulted in retinal dysplasia and atrophy (Schweitzer et al. 1987).

#### 3.2.4.7 Cancer

The carcinogenic effects of high doses of ionizing radiation have been well documented (ATSDR 1999), though the effects of lower doses are less clearly defined. Duncan et al. (1977) reported on a cohort of patients who had received radiotherapy for carcinoma of the cervix. Eight of 2,674 patients developed bladder tumors within 6 months to 20 years following irradiation; the incidence rate was over 57 times greater than the general female population. All eight patients had received <sup>60</sup>Co irradiation, though five of the eight also received radium therapy in conjunction with <sup>60</sup>Co irradiation. Wollenberg et al. (1995) presented a case of a 55-year-old farmer who received a total of 25,150 rad (251.5 Gy) distributed over six areas of the body over an 8-month period as a <sup>60</sup>Co teletherapy treatment regimen. Twenty years after irradiation, the patient developed a total of 43 basal cell carcinomas over the treated areas, all of which were successfully removed with cryosurgery. A 2-year-old girl exposed to 1,800 rad (18 Gy) of <sup>60</sup>Co radiation as part of a treatment regimen for acute lymphoblastic leukemia L1 developed, at age 12, a basal cell carcinoma of the scalp (Garcia-Silva et al. 1996). Three patients receiving cobalt irradiation as part of a chemotherapy/radiation treatment developed basal cell carcinoma of the scalp 8–15 years after treatment in the area of radiation treatment (Dinehart et al. 1991).

#### 3.3 GENOTOXICITY

*Stable Cobalt.* No studies were located regarding genotoxic effects in humans following oral or dermal exposure to cobalt. No studies were located regarding genotoxic effects in animals following inhalation exposure to cobalt.

Gennart et al. (1993) examined a cohort of 26 male workers who were occupationally-exposed to cobalt, chromium, nickel, and iron. Analysis of variance on sister-chromatid exchange rank values revealed that exposure status (exposed vs. controls) and smoking habits had statistically significant effects.

Single oral exposure of male Swiss mice to 0, 4.96, 9.92, or 19.8 mg cobalt/kg as cobalt chloride resulted in significantly increased percentages of both chromosomal breaks and chromosomal aberrations in bone marrow cells, with significant linear trends toward increasing aberrations with increased exposure (Palit et al. 1991a, 1991b, 1991c, 1991d).

Results of genetic testing of cobalt are presented in Table 3-5. Several different forms of cobalt, including cobalt chloride and cobalt sulfide, were tested. No profound differences were found among the various forms.

Cobalt was found to be generally nonmutagenic in bacteria (Salmonella typhimurium, Escherichia coli) and yeast when compounds with a valence state of II were tested (Arlauskas et al. 1985; Fukunaga et al. 1982; Kanematsu et al. 1980; Kharab and Singh 1985; Ogawa et al. 1986; Singh 1983; Tso and Fung 1981). A very weak mutagenic response was found with *Bacillus subtilis* (Kanematsu et al. 1980). A mutagenic response to cobalt was found, however, when compounds with a valence state of III were tested in S. typhimurium and E. coli (Schultz et al. 1982). The authors suggested that this may be due to the formation of cobalt(III) complexes that are inert to ligand substitution, allowing optimal interaction of cobalt with genetic material (Schultz et al. 1982). Other studies have shown cobalt to be a comutagen in combination with 4-substituted pyridines in S. typhimurium (Ogawa et al. 1988). It has been reported that cobalt acts as an antimutagen in bacterial (S. typhimurium, B. subtilis, E. coli) and yeast test systems (Saccharomyces cerevisiae) (Inoue et al. 1981; Kada et al. 1986; Kuroda and Inoue 1988). A possible explanation was that cobalt acts by correcting the error-proneness of deoxyribonucleic acid (DNA) replicating enzymes by improving their performance in DNA synthesis (Inoue et al. 1981; Kada et al. 1986; Kuroda and Inoue 1988). However, cobalt has also been shown to increase the frequency of genetic conversions in S. cerevisiae (Kharab and Singh 1985; Singh 1983). The reasons for this apparent dichotomy in yeast cells is not known.

In contrast to the results seen in bacteria, stable cobalt compounds were generally found to be genotoxic or mutagenic in mammalian assay systems. Exposure to cobalt compounds (metal, salts, or hard metal) has been shown to produce clastogenic effects in mammalian cells, including human lymphocytes (Anard et al. 1997; Hamilton-Koch et al. 1986; Painter and Howard 1982); transformation in hamster cells (Costa et al. 1982); sister chromatid exchanges in human lymphocytes (Andersen 1983); and micronucleus formation in mouse bone marrow cells (Suzuki et al. 1993) and human lymphocytes (Capomazza and Botta 1991; Olivero et al. 1995; Van Goethem et al. 1997). Hard metal is generally more genotoxic in *in vitro* tests than other cobalt compounds. Cobalt ions are also thought to inhibit DNA repair in mammalian cells by interaction with zinc-finger proteins involved in DNA excision repair (Asmuß et al. 2000; De Boeck et al. 1998; Hartwig et al. 1991; Kasten et al. 1997; Nackerdien et al. 1991; Sarkar 1995).

Table 3-5. Genotoxicity of Cobalt In Vitro

		Re	esults	_	
Species (test system)	End point	With activatio	Without activatio n	Reference	Valenc e state
Stable Cobalt					
Prokaryotic organisms:					
Salmonella typhimurium (plate incorporation)	Gene mutations	No data	-	Tso and Fung 1981	II
S. typhimurium (plate incorporation)	Gene mutations	No data	_	Arlauskas et al. 1985	II
S. typhimurium plate incorporation)	Gene mutations	No data	_	Ogawa et al. 1986	II
S. typhimurium (plate incorporation)	Gene mutations	No data	+	Schultz et al. 1982	III
Bacillus subtilis (rec assay)	Gene mutations	No data	(+)	Kanematsu et al. 1980	II
Escherichia coli (reversion assay)	Gene mutations	No data	-	Kanematsu et al. 1980	II
E. coli (repair assay)	DNA damage	No data	+	Schultz et al. 1982	III
Eukaryotic organisms:					
Fungi: Saccharomyces cerevisiae (plate assay)	Reversion	No data	_	Kharab and Singh 1985	II
S. cerevisiae (plate assay)	Reversion	No data	_	Fukunaga et al. 1982	II
S. cerevisiae (plate assay)	Reversion	No data	_	Singh 1983	II
S. cerevisiae (plate assay)	Conversion	No data	+	Kharab and Singh 1985	II
S. cerevisiae (plate assay)	Conversion	No data	+	Fukunaga et al. 1982	II
S. cerevisiae (plate assay)	Conversion	No data	+	Singh 1983	II

Table 3-5. Genotoxicity of Cobalt In Vitro (continued)

		Re	esults	_	
Species (test system)	End point	With activatio n	Without activatio n	Reference	Valenc estate
Mammalian cells:					
Hamster ovary cells	Clastogenic effects	No data	+	Hamilton-Koch et al. 1986	II
Hamster embryo cells	Transformation	No data	+	Costa et al. 1982	II
Human lymphocytes	Sister chromatid exchange	No data	+	Andersen 1983	II
Human HeLa cells	Inhibition of DNA synthesis	No data	+	Painter and Howard 1982	II
Human diploid fibroblasts	DNA damage	No data	+	Hamilton-Koch et al. 1986	II
Radioactive Cobalt					
Mammalian cells:					
Chinese hamster ovary cells	DNA amplification	No data	+	Luecke-Huhle et al. 1986	N/A
Hamster embryo cells	DNA amplification	No data	+	Luecke-Huhle et al. 1990	N/A
Mouse lymphosarcoma cells	Chromosomal aberrations	No data	+	Juraskova and Drasil 1987	N/A
Mouse lymphosarcoma cells	Sister- chromatid exchanges	No data	+	Juraskova and Drasil 1987	N/A
Human lymphocytes	Chromosomal aberrations	No data	+	Koksal et al. 1995	N/A

Table 3-5. Genotoxicity of Cobalt In Vitro (continued)

		Re	esults	_	
Species (test system)	End point	With activatio	Without activatio n	Reference	Valend estate
Human lymphocytes	Micronucleus formation	No data	+	Koksal et al. 1996	N/A
Human leukocytes	DNA strand breaks	No data	+	Rueff et al. 1993	N/A
Human leukocytes	Chromosomal aberrations	No data	+	Rueff et al. 1993	N/A
Human leukocytes	Chromosome breaks	No data	+	Lindahl-Kiessling et al. 1970	N/A
Human fibroblasts	Transformation	No data	+	Namba et al. 1981	N/A
Human fibroblasts	Transformation	No data	+	Namba et al. 1985	N/A
Human fibroblasts	DNA strand breaks	No data	+	Coquerelle et al. 1987	N/A
Human fibroblasts	Transformation	No data	+	Namba et al. 1988	N/A
Human fibroblasts	Retinoblastoma gene alterations	No data	+	Endo et al. 1993	N/A
Human fibroblasts	DNA strand breaks	No data	+	Dolling et al. 1998	N/A
Human kidney cells	DNA strand breaks	No data	+	Feinendegen et al. 1977	N/A
Human kidney cells	DNA strand breaks	No data	+	Feinendegen et al. 1978	N/A

Thirty hours following single intraperitoneal injection of cobalt(II) chloride in BALB/c mice, an increase in micronucleus formation was seen at 12.4 or 22.3 mg cobalt/kg (as cobalt chloride), but not at 6.19 mg/kg (Suzuki et al. 1993). Single injection of mg cobalt/kg (as cobalt chloride) resulted in significantly increased micronucleus formation at 24 hours post-injection, but not at 12, 48, 72, or 96 hours. Two or 10 days following intraperitoneal injection of male and female F344 rats with 3 or 6 mg cobalt/kg, increased levels of oxidatively-damaged DNA bases were noted in the liver, kidney, and to a lesser extent the lung (Kasprzak et al. 1994).

Radioactive Cobalt. The ability of ionizing radiation to induce genotoxic damage is well-documented (ATSDR 1999). Chang et al. (1999c) reported increased micronucleus frequency, both of single and multiple nucleates, in 48 people who had been exposed to 12–1,600 rad (0.12–16 Gy) over a 2–10-year period as a result of a building contaminated with <sup>60</sup>Co rods. Subjects who had left the building showed a decrease in micronucleus formation that correlated with time since cessation of exposure. Three workers accidentally exposed to 2.2–12.7 rad (0.022–0.127 Gy) showed no elevation in frequency of chromosome alterations (House et al. 1992). Ten children who received chemotherapy and 1,725–2,405 rad (17.25–24.05 Gy) as cobalt radiotherapy for acute lymphatic leukemia showed no clastogenic changes after chemotherapy but before irradiation, but showed significant dose-related increases in chromosomal aberrations after irradiation (Rauscher and Bauchinger 1983).

Radiation from cobalt isotopes has been shown to induce numerous genetic changes, including translocations (Gilot-Delhalle et al. 1988; Grahn and Carnes 1988; Grahn et al. 1983; Searl et al. 1976), decreased DNA synthesis (Lohmann et al. 1966), dominant lethal mutations (Grahn et al. 1986; Searl et al. 1976; Zhou et al. 1986), chromosome deletions (Brooks et al. 1971b, 1974), polycentrics (Brooks et al. 1971a, 1974), and aberrations (Brooks et al. 1971b) in exposed animals.

Radiation from cobalt isotopes was genotoxic in several assay systems in mammalian cells: DNA amplification in hamster cells (Lucke-Huhle et al. 1986, 1990); chromosomal aberrations and sister-chromatid exchanges in mouse lymphosarcoma cells (Juraskova and Drasil 1987); chromosomal aberrations and micronucleus formation in human lymphocytes (Koksal et al. 1995, 1996); DNA breakage in human leukocytes (Lindahl-Kiessling et al. 1970; Reuff et al. 1993), kidney cells (Feinendegen et al. 1977), and fibroblasts (Coquerelle et al. 1987; Dolling et al. 1998); chromosomal aberrations in human leukocytes (Reuff et al. 1993); transformation of human fibroblasts (Namba et al. 1981, 1985, 1988); and retinoblastoma gene alterations in human fibroblasts (Endo et al. 1993).

#### 3.4 TOXICOKINETICS

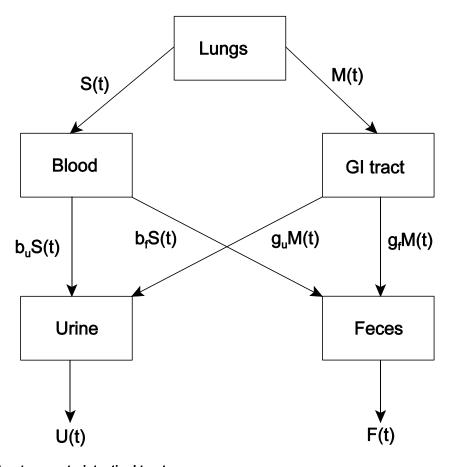
### 3.4.1 Absorption

# 3.4.1.1 Inhalation Exposure

Inhaled cobalt particles are deposited in the upper and lower respiratory tract and cobalt is subsequently absorbed by several mechanisms (Casarett and Doull 1986). The deposition pattern in the respiratory tract is related to particle size, which determines the degree to which particles are affected by inertial impaction, sedimentation, diffusion, and electrostatic precipitation. Large particles (diameter  $>2 \mu m$ ) tend to deposit in the upper respiratory tract where high airstream velocities and airway geometry promote inertial impaction of larger particles. Smaller particles escape inertial impaction and enter the lower respiratory tract where lower airstream velocities and airway geometry favor the process of sedimentation, diffusion, and electrostatic precipitation of small particles. Fractional deposition of inhaled cobalt oxide particles in humans varied from approximately 50% for particles with a mean geometric diameter of 0.8  $\mu$ m to approximately 75% for particles with a mean diameter of 1.7  $\mu$ m (Foster et al. 1989). Fractional deposition can be expected to vary considerably with age, particle size, and breathing patterns (see Table 3-10).

The transfer pathways of cobalt oxide (<sup>57</sup>Co used as a tracer) from the lungs in humans and animals are shown in Figure 3-4. Particles of cobalt deposited in the respiratory tract can be translocated into the blood after dissolution (S(t)) or mechanically transferred to the gastrointestinal tract by mucociliary action of the respiratory tract and swallowing action (M(t)). Only a portion (probably <50%) of the cobalt that enters the gastrointestinal tract will be absorbed. The relative magnitude of the translocation and mechanical clearance pathways depends on the size and solubility of the cobalt particles that are inhaled. Large particles (>2 μm) will tend to deposit in the middle and upper airways where mechanical clearance mechanisms predominate over translocation. Smaller particles that enter the lower respiratory tract will tend to remain until dissolved or phagocytized by macrophages and translocation occurs. The sum of the activities of translocation and mechanical clearance determine the kinetics of absorption of inhaled cobalt. In humans, the ratio of translocation (S(t)) to mechanical clearance (M(t)) is approximately 5–1 for particle sizes ranging from 0.8 to 1.7 μm (mean geometric diameter) (Foster et al. 1989).

Figure 3-4. Transfer Parameters for Cobalt Following Inhalation of Cobalt Oxide\* (CO<sub>3</sub>O<sub>4</sub>) Particles, Showing the Fractions of the Lung Content, L(t), and Time, t, Cleared Per Day by Each Route\*\*



GI tract = gastrointestinal tract;

b<sub>r</sub>S(t) = fraction of cobalt excreted in the feces after translocation; b<sub>r</sub>S(t) = fraction of cobalt excreted in the urine after translocation;

b<sub>u</sub>S(t) = fraction of cobalt exercise F(t) = fecal excretion rate;

g<sub>f</sub>M(t) = fraction of cobalt excreted in the feces after mechanical clearance to the gastrointestinal tract;

g<sub>u</sub>M(t) = fraction of cobalt excreted in the urine after mechanic clearance to the gastrointestinal tract;

M(t) = rate of mechanical transport of cobalt particles from the lungs to the gastrointestinal tract;

S(t) = rate of translocation of cobalt from the lugns to the blood;

U(t) = urinary excretion rate

\*Cobalt-57 tracer used

\*\*Derived from Bailey et al. 1989

Data on retention of cobalt oxide (<sup>57</sup>Co used as a tracer) in the respiratory tracts of humans and several animal species are summarized in Table 3-6. Considerable variability exists between species. In humans, almost one-half of the original lung burden persisted 6 months after exposure; in rats, clearance of cobalt from the lungs was nearly complete after 6 months. The elimination half-time for cobalt in the human lung increased with increasing time after exposure (Foster et al. 1989; Sedlet et al. 1958). This may reflect slower clearance of cobalt that is bound to cellular components in the lung (Kreyling et al. 1985, 1986).

# 3.4.1.2 Oral Exposure

Gastrointestinal absorption of cobalt in humans varies considerably (18–97% of the given dose) based on the type and dose of cobalt compound given and the nutritional status of the subjects (Harp and Scoular 1952; Smith et al. 1972; Sorbie et al. 1971; Valberg et al. 1969). More cobalt was absorbed through the gastrointestinal tract of humans when the body was deficient in iron (31–71% in iron deficiency; 18–44% in controls) (Sorbie et al. 1971; Valberg et al. 1969). One study in humans has showed that oral exposure to cobalt resulted in significantly higher urinary excretion in females relative to males (Christensen et al. 1993).

In animal studies, many factors have been shown to influence the absorption of cobalt compounds following oral exposure. In several studies in rats (Ayala-Fierro et al. 1999; Barnaby et al. 1968; Hollins and McCullough 1971; Kirchgessner et al. 1994; Schade et al. 1970; Taylor 1962), soluble cobalt chloride was absorbed in the range of 13–34%, whereas physiologically insoluble cobalt oxide particles have been shown to be poorly absorbed, in the range of 1–3% (Bailey et al. 1989; Collier et al. 1989; Patrick et al. 1989). The particle size of the given dose of cobalt oxide had no significant effect on absorption (see Table 3-7). Administration of cobalt chloride labeled with radioactive <sup>58</sup>Co and complexed with histidine, lysine, glycylglycine, ethylenediaminetetraacetic acid (EDTA), casein, or glycine resulted in decreased gastrointestinal absorption of cobalt; administration of cobalt chloride (with <sup>58</sup>Co tracer) in cows' milk permitted a significantly greater (about 40%) absorption through the gastrointestinal tract (Taylor 1962). The same study found that while there was no difference in the chlorides of cobalt(II) and cobalt(III), a cobalt(II) glycine complex was absorbed in greater quantities than a cobalt(III) glycine complex. Other studies have also demonstrated that the chemical form of the cobalt compound can affect the absorption of cobalt following oral exposure (Deka et al. 1981; Firriolo et al. 1999; Inaba et al. 1980; Kinoshita and Fujita 1972).

Table 3-6. Initial (Day 3) Lung Deposits of Cobalt Oxide and Summary of Lung Retention at 90 and 180 Days<sup>a,b</sup>

Species (strain)	Mean initial <sup>57</sup> Co activity in lung L(3) (kBq)		Lung retention L(90)/L(3) (%)		Lung retention L(180)/L(3) (%)	
	0.8 µm	1.7 µm	0.8 µm	1.7 µm	0.8 µm	1.7 µm
Human	53	42	64	75	45	56
Baboon	2,100	1,700	55	55	26	37
Beagle dog	1,150	1,450	27	45	5.5	12
Guinea pig (Harwell)	8.4	1.4	49	46	8.3	15
Rat (HMT, 1985)	10.8	4.7	5.2	20	1.3	8.0
Rat (HMT, 1986)	3.2	0.7	5.3	18	1.2	9.2
Rat (F344, SPF)	8.8	4.4	14	25	4.7	9.2
Rat (Sprague- Dawley)	0.9	0.10	8	39	1	15
Syrian hamster	4.0	1.2	21	35	3.4	12
Mouse (CBA/H)	1.8	No data	15	No data	2.8	No data

<sup>&</sup>lt;sup>a</sup>Derived from Bailey et al. 1989

<sup>&</sup>lt;sup>b</sup>Cobalt-57 used as tracer

Table 3-7. Summary of Measurements of Retention and Excretion After Intragastric Administration of Cobalt Oxide (Co<sub>3</sub>O<sub>4</sub>) Particles (Mean Percentage of Recovered Activity at 7 Days After Administration)<sup>a,b</sup>

Species (strain)	Cumulative fecal excretion		Whole bo	Whole body retention		Cumulative urinary excretion		Absorption	
	0.8 µm	1.7 µm	0.8 µm	1.7 µm	0.8 µm	1.7 µm	0.8 µm	1.7 µm	
Baboon	97.8	98.4	0.12	0.20	2.0	1.4	2.6	1.9	
Guinea pig	98.7	97.6	0.16	0.66	1.1	1.9	1.3	2.3	
Rat (HMT)	96.3	99.4	0.09	0.02	2.8	0.6	3.9	1.0	
Rat (F-344)	99.6	99.7	0.04	0.03	0.4	0.3	0.4	0.3	
Hamster	96.0	96.3	0.50	0.18	3.5	3.5	5.1	5.1	
Mouse (CBA/H)	99.1	No data	0.3	No data	0.6	No data	0.8	No data	

<sup>&</sup>lt;sup>a</sup>Derived from Bailey et al. 1989

\*\*\*DRAFT FOR PUBLIC COMMENT\*\*\*

<sup>&</sup>lt;sup>b</sup>Cobalt 57 used as tracer

Iron deficiency led to increased absorption of cobalt from the gastrointestinal tract, and simultaneous administration of cobalt and iron reduced the amount of cobalt absorbed (Reuber et al. 1994; Schade et al. 1970). Increasing oral doses of cobalt resulted in decreased fractional absorption (Houk et al. 1946; Kirchgessner et al. 1994; Taylor 1962), and more soluble forms of cobalt were better absorbed than less soluble compounds (Kreyling et al. 1986). Absorption is 3- to 15-fold greater in younger animals (rats and guinea pigs examined from days 1–60 of life) than in adult (200 days of age) animals (Naylor and Harrison 1995). Species differences in absorption of cobalt oxide do not appear to exist (Bailey et al. 1989), but absorption of soluble cobalt compounds is greater in rats (13–34%) than in dairy cows (1–2%) and guinea pigs (4–5%) following oral exposure (Ayala-Fierro et al. 1999; Barnaby et al. 1968; Hollins and McCullough 1971; Kirchgessner et al. 1994; Naylor and Harrison 1995; Schade et al. 1970; Taylor 1962; van Bruwaene et al. 1984).

### 3.4.1.3 Dermal Exposure

Four volunteers who placed their right hands in a box filled with hard metal dust (~5–15% cobalt metal, 95–85% tungsten carbide) for 90 minutes showed an increase in urinary cobalt levels by an order of magnitude in the post-exposure samples, remaining elevated for as long as 48–60 hours (Scansetti et al. 1994).

The absorption of 2.2x10<sup>-5</sup> mg <sup>60</sup>Co/kg as cobalt chloride in 1.4N HCl through the intact or abraded skin of guinea pigs was examined by Inaba and Suzuki-Yasumoto (1979). Absorption through intact skin was very small (<1%), while absorption through abraded skin was almost 80% 3 hours after exposure. A study in hamsters (Lacy et al. 1996) also reported a low amount of absorption of cobalt through unabraded skin.

#### 3.4.2 Distribution

As a component of vitamin B12, cobalt is an essential element and, therefore, is found in most body tissues. It has been identified in liver, muscle, lung, lymph nodes, heart, skin, bone, hair, stomach, brain, pancreatic juice, kidneys, plasma, and urinary bladder of nonexposed subjects, with the highest cobalt concentration found in the liver (Collecchi et al. 1986; Forbes et al. 1954; Hewitt 1988; Ishihara et al. 1987; Muramatsu and Parr 1988; Teraoka 1981; Yamagata et al. 1962; Yukawa et al. 1980) (see Chapter 6 for more information). Tissue levels reflected exposure from all routes. The total body content

of cobalt has been estimated at 1.1–1.5 mg (ICRP 1979; Yamagata et al. 1962); about 0.11 mg was found in the liver (ICRP 1979).

In patients with laryngeal carcinoma, levels of cobalt in the tumor were significantly higher (p<0.001) than levels in the nonmalignant tissues around the tumor (68.7 ng/g tissue versus 39.6 ng/g) (Collecchi et al. 1986). The mean cobalt concentrations in plasma (18.3 ng/mL) were also significantly higher in these patients than in the comparison population (0.73 ng/mL). The clinical significance of these findings is not known.

# 3.4.2.1 Inhalation Exposure

In workers occupationally exposed to cobalt, increased cobalt levels were found in tissues at death. Significant increases in cobalt in the lung have been found in copper smelter and metal workers and coal miners occupationally exposed to cobalt (Gerhardsson et al. 1984; Hewitt 1988; Hillerdal and Hartung 1983; Teraoka 1981). No increase in liver or kidney cobalt levels were found in the copper smelter workers as compared to controls (Gerhardsson et al. 1984). In metal workers, increased cobalt levels were also found in the lymph nodes, liver, spleen, and kidneys (Hillerdal and Hartung 1983; Teraoka 1981).

The tissue distribution of cobalt in animals is similar to that in humans, with marked increases in the concentration of cobalt in the lungs following inhalation exposure (Barnes et al. 1976; Brune et al. 1980; Collier et al. 1991; Kreyling et al. 1986; Kyono et al. 1992; Patrick et al. 1989; Talbot and Morgan 1989). Histologically, the particles of cobalt in the lung are found in macrophages within the bronchial wall or in the interstitium close to the terminal bronchioli (Brune et al. 1980). Significant concentrations of cobalt have been found in the liver, kidney, trachea, spleen, bones, and heart (Barnes et al. 1976; Brune et al. 1980; Kerfoot 1975; Kreyling et al. 1986; Wehner and Craig 1972), with the greatest concentrations in the liver and the kidney (Kerfoot 1975; Wehner and Craig 1972).

#### 3.4.2.2 Oral Exposure

No studies were located regarding distribution in humans after oral exposure to cobalt.

In animals, the cobalt absorbed through the gastrointestinal tract was primarily retained in the liver (Ayala-Fierro et al. 1999; Greenberg et al. 1943; Simesen 1939). Appreciable levels were also found in the kidneys, heart, stomach, and intestines (Ayala-Fierro et al. 1999; Persson et al. 1992; Simesen 1939). Following a single oral dose of cobalt napthenate, appreciable levels of cobalt were found in heart, liver, and kidney but not in spleen or testes (Firriolo et al. 1999).

Following longer-term exposure (8 weeks) to cobalt sulfate in the diet, exposed rats showed a 30-fold increase in the cobalt concentration in the myocardium, a 26-fold increase in the concentration in the soleus muscle, and a 100-fold increase in the concentration in serum compared with nonexposed controls (Clyne et al. 1988; Pehrsson et al. 1991). Long-term oral exposure of rats to cobalt chloride resulted in significantly increased levels of cobalt in the liver, kidney, muscle, brain, and testes of treated rats (Barnaby et al. 1968; Bourg et al. 1985; Thomas et al. 1976).

# 3.4.2.3 Dermal Exposure

No studies were located regarding distribution in humans or animals after dermal exposure to cobalt.

# 3.4.2.4 Other Routes of Exposure

Following intravenous injection of cobalt chloride (as a combination of radioactive <sup>55</sup>CoCl<sub>2</sub> and <sup>56</sup>CoCl<sub>2</sub>) in two volunteers, the liver and bladder contained the highest portions of cobalt (Jansen et al. 1996).

Two hours after intravenous injection of cobalt chloride (with a radioactive <sup>57</sup>Co tracer) in rats, accumulation was found in the liver (22.8% of the dose), kidneys (10.2%), and intestines (3.16%) (Gregus and Klaassen 1986). Similar results (29% liver, 10% kidneys, 4.6% intestines) were found following intracardiac injection of cobalt nitrate in rats (Patrick et al. 1989), or intravenous injection of a combination of radioactive <sup>55</sup>CoCl<sub>2</sub> and <sup>56</sup>CoCl<sub>2</sub> in rats (exact percentages not given in manuscript) (Jansen et al. 1996). One hundred days after intravenous injection of <sup>60</sup>CoCl<sub>2</sub> in rats, the greatest concentrations were found in spleen > heart > bone. Liver and kidney, initially the highest in cobalt, containing comparatively low amounts of cobalt (Thomas et al. 1976). Similar results were seen 132 days following an intraperitoneal injection of <sup>60</sup>CoCl<sub>2</sub> in rats (Barnaby et al. 1968). Intramuscular injection of cobalt mesoporphyrin in rats yielded the greatest levels of cobalt in liver and blood, followed by kidney, lung, spleen, adrenal glands, and heart at 7 days post-injection and later (Feng et al. 1998). Four weeks

after subcutaneous administration of cobalt protoporphyrin, the greatest tissue levels of cobalt occurred in the kidney, followed by spleen, liver, lung, thymus, and gonads (Rosenberg 1993). When cobalt (with a <sup>57</sup>Co tracer) encapsulated in liposomes was intravenously injected into rats, decreased distribution to the heart (40% less than animals receiving cobalt chloride), kidneys, and carcass, and increased distribution to the spleen and bones were found (Szebeni et al. 1989).

#### 3.4.3 Metabolism

Cobalt is essential in the body in that it is a component of cyanocobalamin (vitamin  $B_{12}$ ) (Vouk 1986). Vitamin  $B_{12}$  acts as coenzyme in many enzymatic reactions, most notably a methyl transfer reaction that converts homocysteine to methionine and for a separate reaction that converts L-methylmalonylcoenzyme A (CoA) to succinyl-CoA (Institute of Medicine 2000). Vitamin  $B_{12}$  is also involved in some enzymes involved in hematopoiesis; deficiency can lead to pernicious anemia (Domingo 1989). No other essential function of cobalt has been reported. The Recommended Dietary Allowance (RDA) for vitamin  $B_{12}$  for adults is 2.4  $\mu$ g/day, which contains 0.1  $\mu$ g of cobalt (Institute of Medicine 2000).

#### 3.4.4 Elimination

### 3.4.4.1 Inhalation Exposure

No data are available on the clearance of soluble cobalt particles in humans. Following exposure of humans to physiologically insoluble cobalt compounds (cobalt metal, cobalt oxides), clearance from the body, assessed by both urinary/fecal clearance as well as a reduction in whole-body retention, appears to follow three-phase kinetics. The first phase, likely representing mucociliary clearance of particles deposited in the tracheobronchial region, has a half-time on the order of 2–44 hours (Apostoli et al. 1994; Mosconi et al. 1994b). The second phase, with a half-time on the order of 10–78 days, may represent macrophage-mediated clearance of cobalt particles from the lung (Beleznay and Osvay 1994; Mosconi et al. 1994b). The third clearance phase, representing long-term clearance from the lungs, has a half-time on the order of years (Bailey et al. 1989; Beleznay and Osvay 1994; Mosconi et al. 1994b; Newton and Rundo 1971). Following a controlled aerosol exposure in humans, about 40% of the initial lung burden of inhaled cobalt oxide (with a <sup>57</sup>Co tracer) was retained for a period of 6 months after exposure (Foster et al. 1989). Within the first week, about 17% of the initial lung burden was eliminated, with the

majority (about 90%) mechanically cleared to the gastrointestinal tract and excreted in the feces (Foster et al. 1989). Six months after exposure, a cumulative elimination of 33% of the initial lung burden was found in the urine and 28% was found in the feces (Foster et al. 1989). The ratio of peak translocation rate to average mechanical clearance rate (Figure 3-4 and Table 3-8) was about 5 to 1. The elimination of cobalt following inhalation exposure was affected by the time after exposure (urinary excretion increases as time increases) and particle size (more cobalt is initially mechanically cleared to the gastrointestinal tract when the aerosol consists of bigger particles) (Bailey et al. 1989; Foster et al. 1989).

In animals, the solubility of the cobalt compound appears to greatly affect its long-term clearance. Studies with cobalt oxides have shown that the more soluble CoO is cleared from the lungs at a greater rate than the less soluble Co<sub>3</sub>O<sub>4</sub> (Barnes et al. 1976; Kreyling 1984a). More soluble cobalt compounds are absorbed into the blood at a greater rate, and excreted in the urine and, to a lesser extent, the feces (Barnes et al. 1976). The rate of urinary excretion appears to correlate with the rate of translocation of cobalt from the lungs to the blood, and the rate of fecal clearance with the rate of mechanical clearance of cobalt from the lungs to the gastrointestinal tract (Andre et al. 1989; Bailey et al. 1989; Collier et al. 1989; Kreyling et al. 1986, 1989; Patrick et al. 1989; Talbot and Morgan 1989). Following an initial high rate of fecal clearance, urinary excretion was the primary route of cobalt excretion after a single inhalation exposure (2 weeks of exposure) (Palmes et al. 1959) or 3 months of exposure (Kerfoot 1975; Palmes et al. 1959). In several species of animals, most of the inhaled Co<sub>3</sub>O<sub>4</sub> (with a <sup>57</sup>Co tracer) following a single exposure was cleared from the lungs by 6 months after exposure (Table 3-6) (Andre et al. 1989; Bailey et al. 1989; Collier et al. 1989; Kreyling et al. 1989; Patrick et al. 1989; Talbot and Morgan 1989). The peak translocation and average mechanical clearance of cobalt from the lungs for different species are reported in Table 3-8, with the rate (high to low) following as mouse > rat > hamster > guinea pig > baboon, human > beagle dog.

#### 3.4.4.2 Oral Exposure

In humans orally exposed to cobalt, fecal elimination, which is the primary route of excretion, varies considerably (3–99% of the dose) and depends on the amount and type of cobalt given and on the nutritional status of the subjects (Section 3.4.1.2) (Harp and Scoular 1952; Paley et al. 1958; Smith et al. 1972; Sorbie et al. 1971; Valberg et al. 1969). Within days after oral exposure, 10 times more cobalt is excreted in feces than in the urine (Paley et al. 1958). Less cobalt is eliminated in the feces (more was absorbed) in subjects with an iron deficiency (Sorbie et al. 1971; Valberg et al. 1969).

Table 3-8. Peak Translocation and Average Mechanical Clearance Rates After Inhalation of Cobalt Oxide<sup>a,b</sup>

		Perd	cent of lung	g content clea	red per day
	Transloca	ation at peak			Mechanical clearance <sup>c</sup>
Species (strain)	0.8 µm	Peak day	1.7 µm	Peak day	Average
Human	0.45	180	0.5	180	0.1
Baboon	0.6	180	0.2	d	0.1
Beagle dog	2.1	85	1.7	180	0.03
Guinea pig	2.1	180	1.0	75	0.3
Rat HMT	2.4	40	0.6	d	0.9
Rat (F-344)	1.1	10	0.4	d	1.0
Hamster	1.8	180	0.7	180	0.8
Mouse	1.7	180	No data	No data	1.05

<sup>&</sup>lt;sup>a</sup>Derived from Bailey et al. 1989 <sup>b</sup>Cobalt-57 used as tracer

<sup>°</sup>Clearance rates were virtually identical in both particle size groups

dConstant value over 180 days

Fecal elimination of cobalt is the primary route of elimination in animals following oral exposure and depends upon the particle size (increasing fecal clearance with increasing size) and solubility (decreasing fecal clearance with increasing solubility) of the cobalt compound. The cumulative urinary and fecal elimination in several species following oral administration of Co<sub>3</sub>O<sub>4</sub> (with a <sup>57</sup>Co tracer) is reported in Table 3-7 (Bailey et al. 1989). Following oral administration, very little Co<sub>3</sub>O<sub>4</sub> was absorbed through the gastrointestinal tract and most (>96%) was quickly elimination in the feces. No significant differences in elimination of Co<sub>3</sub>O<sub>4</sub> were found between species of animals (Andre et al. 1989; Bailey et al. 1989; Collier et al. 1989; Patrick et al. 1989; Talbot and Morgan 1989). For the more soluble cobalt(II) chloride, reported fecal elimination levels have ranged from 70 to 83% of the administered dose for rats, with urinary excretion accounting for the majority of the remainder of the dose (Ayala-Fierro et al. 1999; Barnaby et al. 1968; Hollins and McCullough 1971). In lactating dairy cows, about 97% of an oral dose of cobalt chloride was recovered in the feces by day 70 post-exposure, while the urine and milk contained 0.26 and 0.012% of the dose, respectively (van Bruwaene et al. 1984). In beagle dogs, more Co<sub>3</sub>O<sub>4</sub> (physiologically insoluble) was elimination in the feces (90% in the feces and 5% in the urine) than cobalt nitrate (soluble) (70% in the feces and 25% in the urine) following a single oral exposure (Kreyling et al. 1986).

As is the case for absorption of cobalt compounds, the iron status of the animal also appears to affect the elimination of cobalt compounds. Following oral exposure, iron-deficient animals elimination less of a given dose in the feces than normal rats, while co-administration of iron compounds resulted in an increased fecal excretion of cobalt compounds (Reuber et al. 1994; Schade et al. 1970)

# 3.4.4.3 Dermal Exposure

No studies were located regarding excretion in humans after dermal exposure to cobalt.

Lacy et al. (1996) reported that the majority of the absorbed dose of CoCl<sub>2</sub> was excreted in the urine 48 hours after a single dermal exposure in Syrian hamsters. No other studies were located regarding excretion in animals after dermal exposure to cobalt.

## 3.4.4.4 Other Routes of Exposure

Following intravenous injection of cobalt chloride in humans, about 30% of the dose was excreted in the urine within 24 hours (Smith et al. 1972), 56–73% was excreted within 48 hours (Paley et al. 1958), or 57% within 2 weeks (Kent and McCance 1941).

Following intravenous injection of cobalt nitrate (with a <sup>57</sup>Co tracer) in various species of animals, most of the injected dose was excreted in the urine; about 80% of the given dose was excreted in the urine within 21 days (Table 3-9) (Andre et al. 1989; Bailey et al. 1989; Collier et al. 1989; Patrick et al. 1989; Talbot and Morgan 1989). Other investigators have also found that the urine is the primary route of cobalt excretion following intravenous administration (Ayala-Fierro et al. 1999; Barnaby et al. 1968; Gregus and Klaassen 1986; Kreyling et al. 1986; Onkelinx 1976; Thomas et al. 1976). Most (5–30%) of the remaining cobalt after intravenous exposure was excreted in the feces, with the majority of studies reporting very little long-term retention. Excretion of cobalt (about 2–7% of the injected dose) in the bile was also reported (Cikrt and Tich 1981; Gregus and Klaasen 1986; Sheline et al. 1945). Elimination following intraperitoneal injection is similar to that seen following intravenous exposure, with urinary excretion being the major route of elimination, and fecal excretion accounting for the majority of the remainder of the dose (Barnaby et al. 1968; Hollins and McCullough 1971; Talbot and Morgan 1989), though long-term clearance may be more balanced between the two (Hollins and McCullough 1971). Following subcutaneous injection, both CoCl<sub>2</sub> and Co(NO<sub>3</sub>)<sub>2</sub> were cleared rapidly from the body (Rosenberg 1993; Talbot and Morgan 1989), with the urine being the major route of clearance (Talbot and Morgan 1989).

Following injection, studies have shown that the chemical form of the cobalt compound can affect its elimination. Subcutaneous injection of cobalt protoporphyrin in rats, in which the cobalt atom is chelated within the porphyrin ring, resulted in a slower elimination from the body than cobalt chloride, with significant cobalt levels (~20% of initial injection) still present in the body 14 days after exposure (Rosenberg 1993). Likewise, intramuscular injection of cobalt mesoporphyrin resulted in primarily in fecal excretion, with a high systemic retention (Feng et al. 1998).

#### 3. HEALTH EFFECTS

Table 3-9. Summary of Measurements of Retention and Excretion of Cobalt Following Injection of Cobalt Nitrate Co(NO<sub>3</sub>)<sub>2</sub> Solution (Mean Percent Recovery)<sup>a,b</sup>

	Whole body retention on day				Cumulative urinary excretion on day			Cumulative fecal excretion on day		
Species (strain)	1	7	21	1	7	21	1	7	21	
Baboon	ND	ND	ND	57	74	80	5	17	20	
Beagle dog	ND	ND	ND	71	86	87	3.4	4.4	4.9	
Guinea pig	34	8	3.5	64	82	85	2.2	10	12	
Rat (HMT)	18	4.2	1.9	64	72	74	18	24	24	
Rat (F-344)	ND	ND	2.9	ND	ND	80	ND	ND	18	
Hamster	27	4.3	1.9	55	68	69	17	28	29	
Mouse	23	2.9	1.1	59	71	72	18	26	27	

<sup>&</sup>lt;sup>a</sup>Derived from Bailey et al. 1989 <sup>b</sup>Cobalt-57 used as tracer

ND = No data

## 3.4.5 Physiologically Based Pharmacokinetic (PBPK)/Pharmacodynamic (PD) Models

Physiologically based pharmacokinetic (PBPK) models use mathematical descriptions of the uptake and disposition of chemical substances to quantitatively describe the relationships among critical biological processes (Krishnan et al. 1994). PBPK models are also called biologically based tissue dosimetry models. PBPK models are increasingly used in risk assessments, primarily to predict the concentration of potentially toxic moieties of a chemical that will be delivered to any given target tissue following various combinations of route, dose level, and test species (Clewell and Andersen 1985). Physiologically based pharmacodynamic (PBPD) models use mathematical descriptions of the dose-response function to quantitatively describe the relationship between target tissue dose and toxic end points.

PBPK/PD models refine our understanding of complex quantitative dose behaviors by helping to delineate and characterize the relationships between: (1) the external/exposure concentration and target tissue dose of the toxic moiety, and (2) the target tissue dose and observed responses (Andersen et al. 1987; Andersen and Krishnan 1994). These models are biologically and mechanistically based and can be used to extrapolate the pharmacokinetic behavior of chemical substances from high to low dose, from route to route, between species, and between subpopulations within a species. The biological basis of PBPK models results in more meaningful extrapolations than those generated with the more conventional use of uncertainty factors.

The PBPK model for a chemical substance is developed in four interconnected steps: (1) model representation, (2) model parametrization, (3) model simulation, and (4) model validation (Krishnan and Andersen 1994). In the early 1990s, validated PBPK models were developed for a number of toxicologically important chemical substances, both volatile and nonvolatile (Krishnan and Andersen 1994; Leung 1993). PBPK models for a particular substance require estimates of the chemical substance-specific physicochemical parameters, and species-specific physiological and biological parameters. The numerical estimates of these model parameters are incorporated within a set of differential and algebraic equations that describe the pharmacokinetic processes. Solving these differential and algebraic equations provides the predictions of tissue dose. Computers then provide process simulations based on these solutions.

The structure and mathematical expressions used in PBPK models significantly simplify the true complexities of biological systems. If the uptake and disposition of the chemical substance(s) is

adequately described, however, this simplification is desirable because data are often unavailable for many biological processes. A simplified scheme reduces the magnitude of cumulative uncertainty. The adequacy of the model is, therefore, of great importance, and model validation is essential to the use of PBPK models in risk assessment.

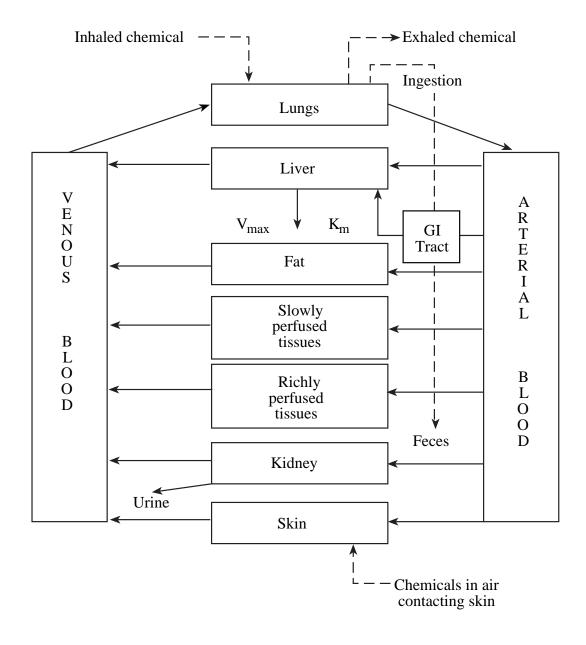
PBPK models improve the pharmacokinetic extrapolations used in risk assessments that identify the maximal (i.e., the safe) levels for human exposure to chemical substances (Andersen and Krishnan 1994). PBPK models provide a scientifically sound means to predict the target tissue dose of chemicals in humans who are exposed to environmental levels (for example, levels that might occur at hazardous waste sites) based on the results of studies where doses were higher or were administered in different species. Figure 3-5 shows a conceptualized representation of a PBPK model.

The ICRP (1995) developed a Human Respiratory Tract Model for Radiological Protection which contains respiratory tract deposition and clearance compartmental models for inhalation exposure that may be applied to particulate aerosols of cobalt compounds. The ICRP (1993) also developed a 3-compartment biokinetic model for human oral exposure that applies to cobalt. EPA (1998) has adopted the ICRP (1993, 1995) models for assessment of radiologic cancer risks from cobalt exposures. The National Council on Radiation Protection and Measurement (NCRP) has also developed a respiratory tract model for inhaled radionuclides (NCRP 1997). At this time, the NCRP recommends the use of the ICRP model for calculating exposures for radiation workers and the general public. Readers interested in this topic are referred to NCRP Report No. 125; *Deposition, Retention and Dosimetry of Inhaled Radioactive Substances* (NCRP 1997). In the appendix to the report, NCRP provides the animal testing clearance data and equations fitting the data which supported the development of the human model for cobalt.

### Human Respiratory Tract Model for Radiological Protection (ICRP 1994).

**Respiratory Tract Deposition.** The ICRP (1994) has developed a deposition model for behavior of aerosols and vapors in the respiratory tract. ICRP (1994) provides inhalation dose coefficients which can be used to estimate the committed equivalent and effective doses to organs and tissues throughout the body based on a unit intake of radioactive material and anticipated distribution and retention of the

Figure 3-5. Conceptual Representation of a Physiologically Based Pharmacokinetic (PBPK) Model for a Hypothetical Chemical Substance



Source: adapted from Krishnan et al. 1994

Note: This is a conceptual representation of a physiologically based pharmacokinetic (PBPK) model for a hypothetical chemical substance. The chemical substance is shown to be absorbed via the skin, by inhalation, or by ingestion, metabolized in the liver, and excreted in the urine or by exhalation.

material, its radioactive decay, and the energy of the radiation emitted from the material and absorbed by tissues. The model applies to three levels of particle solubility, a wide range of particle sizes (approximately 0.0005–100 µm in diameter), and parameter values that can be adjusted for various segments of the population (e.g., sex, age, level of physical exertion). This model also allows one to evaluate the bounds of uncertainty in deposition estimates. Uncertainties arise from natural biological variability among individuals and the need to interpret some experimental evidence that remains inconclusive. It is applicable to particulate aerosols containing cobalt, and was developed for a wide variety of radionuclides and their chemical forms.

The ICRP deposition model estimates the fraction of inhaled particle mass that initially deposits in each compartment (see Figure 3-6). The model was developed with 5 compartments: (1) the anterior nasal passages (ET<sub>1</sub>); (2) all other extrathoracic airways (ET<sub>2</sub>) (posterior nasal passages, the naso- and oropharynx, and the larynx); (3) the bronchi (BB); (4) the bronchioles (bb); and (5) the alveolar interstitium (AI). Particles deposited in each of the regions may be removed from each region and redistributed either upward into the respiratory tree or to the lymphatic system and blood by different particle removal mechanisms.

For extrathoracic deposition of particles, the model uses experimental data, where deposition is related to particle size and airflow parameters, and scales deposition for women and children from adult male data. Similarly to the extrathoracic region, experimental data served as the basis for lung (bronchi, bronchioles, and alveoli) aerosol transport and deposition. A theoretical model of gas transport and particle deposition was used to interpret data and to predict deposition for compartments and subpopulations other than adult males. Table 3-10 provides reference respiratory values for the general Caucasian population under several levels of activity.

Deposition of inhaled gases and vapors is modeled as a partitioning process, which depends on the physiological parameters noted above as well as the solubility and reactivity of compound in the respiratory tract (see Figure 3-7). The ICRP (1994) model defines three categories of solubility and reactivity: SR-0, SR-1, and SR-2:

• Type SR-0 compounds include insoluble and nonreactive gases (e.g., inert gases such as H<sub>2</sub>, He). These compounds do not significantly interact with the respiratory tract tissues and essentially all compound inhaled is exhaled. Radiation doses from inhalation of SR-0 compounds are assumed to result from the irradiation of the respiratory tract from the air spaces.

Table 3-10. Reference Respiratory Values for a General Caucasian Population at Different Levels of Activity

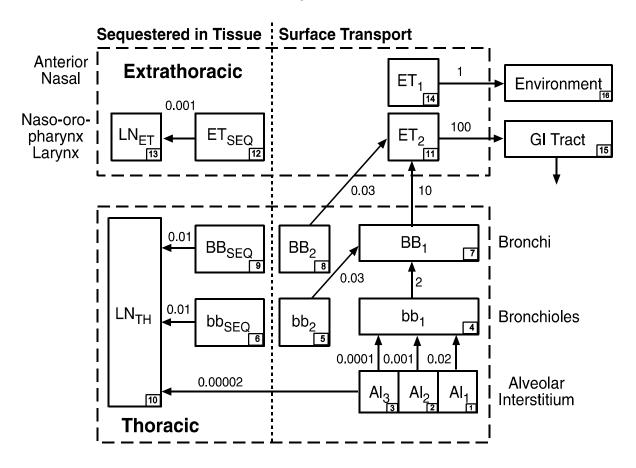
Activity:		Re	Resting (sleeping)		;	Sitting awake		Light exercise			Heavy exercise		
Maximal workload (%):		8		12		32			64				
Breathing p	parameters <sup>b</sup>	ν <sub>τ</sub> (L)	<i>B</i> (m³h <sup>-1</sup> )	f <sub>R</sub> (min <sup>-1</sup> )	<i>V</i> <sub>⊤</sub> (L)	<i>B</i> (m³h <sup>-1</sup> )	f <sub>R</sub> (min <sup>-1</sup> )	<i>V</i> <sub>⊤</sub> (L)	<i>B</i> (m³h <sup>-1</sup> )	<b>f</b> <sub>R</sub> (min <sup>-1</sup> )	<i>V</i> <sub>⊤</sub> (L)	<i>B</i> (m³h <sup>-1</sup> )	<b>f</b> <sub>R</sub> (min <sup>-1</sup> )
Age	Sex	_											
3 months		0.04	0.09	38	N/A	N/A	N/A	0.07	0.19	48	N/A	N/A	N/A
1 years		0.07	0.15	34	0.1	0.22	36	0.13	0.35	46	N/A	N/A	N/A
5 years		0.17	0.24	23	0.21	0.32	25	0.24	0.57	39	N/A	N/A	N/A
10 years	Male: Both:	0.3	0.31	17	0.33	0.38	19	0.58	1.12	32	0.841	2.22	44
	Female:										0.667	1.84	46
15 years	Male:	0.500	0.42	14	0.533	0.48	15	1.0	1.38	23	1.352	2.92	36
	Female:	0.417	0.35	14	0.417	0.40	16	0.903	1.30	24	1.127	2.57	38
Adult	Male:	0.625	0.45	12	0.750	0.54	12	1.25	1.5	20	1.923	3.0	26
	Female:	0.444	0.32	12	0.464	0.39	14	0.992	1.25	21	1.364	2.7	33

<sup>&</sup>lt;sup>a</sup>See Annexe B (ICRP 1994) for data from which these reference values were derived.

 $<sup>{}^{</sup>b}V_{T}$  = Tidal volume, B = ventilation rate,  $f_{R}$  = respiration frequency

h = hour; L = liter(s); min = minute(s); N/A = not applicable

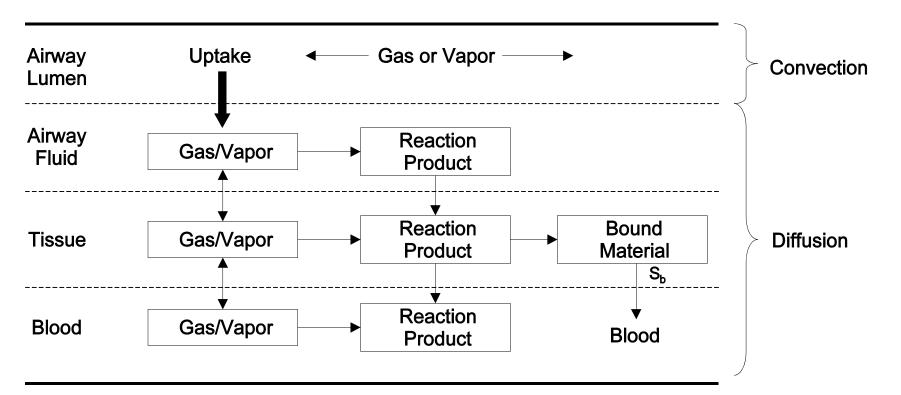
Figure 3-6. Respiratory Tract Compartments\* in Which Particles May be Deposited



<sup>\*</sup>Compartment numbers shown in lower right corners are used to define clearance pathways. The clearance pathways as well as the compartment abbreviations are presented in Table 3-11.

Source: ICRP 1994

Figure 3-7. Reaction of Gases or Vapors at Various Levels of the Gas-Blood Interface



From ICRP (1994)

- Type SR-1 compounds include soluble or reactive gases and vapors which are expected to be taken up by the respiratory tract tissues and may deposit in any or all of the regions of the respiratory tract, depending on the dynamics of the airways and properties of the surface mucous and airway tissues, as well as the solubility and reactivity of the compound.
- Type SR-2 compounds include soluble and reactive gases and vapors which are completely retained in the extrathoracic regions of the respiratory tract. SR-2 type compounds include sulfur dioxide (SO<sub>2</sub>) and hydrogen fluoride (HF).

Respiratory Tract Clearance. This portion of the model identifies the principal clearance pathways within the respiratory tract. The model was developed to predict the retention of various chemical materials. Figure 3-8 presents the compartmental model and is linked to the deposition model (Figure 3-6) and to reference values presented in Table 3-10. Table 3-11 provides deposition fractions and clearance rates for each compartment for insoluble particles. The table provides rates of insoluble particle transport for each of the compartments, expressed as a fraction per day and also as half-time. ICRP (1994) also developed modifying factors for some of the parameters, such as age, smoking and disease status. Parameters of the clearance model are based on human evidence for the most part, although particle retention in airway walls is based on experimental data from animal experiments.

The clearance of deposited particles from the respiratory tract is a dynamic process. The rate of clearance generally changes with time from each region and by each route. Following deposition of large numbers of particles (acute exposure), transport rates change as particles are cleared from the various regions. Physical and chemical properties of deposited material determine the rate of dissolution and as particles dissolve; absorption rates tend to change over time. By creating a model with compartments of different clearance rates within each region (e.g., BB<sub>1</sub>, BB<sub>2</sub>, BB<sub>seq</sub>), the ICRP model overcomes problems associated with time-dependent functions. Each compartment clears to other compartments by constant rates for each pathway.

Particle transport from all regions is toward both the lymph nodes and the pharynx, and a majority of deposited particles end up being swallowed. In the front part of the nasal passages (ET<sub>1</sub>), nose blowing, sneezing, and wiping remove most of the deposited particles. Particles remain here for about a day. For particles with AMADs a few micrometers or greater, the ET<sub>1</sub> compartment is probably the largest deposition site. A majority of particles deposited at the back of the nasal passages and in the larynx (ET<sub>2</sub>) are removed quickly by the fluids that cover the airways. In this region particle clearance is completed within 15 minutes.

#### 3. HEALTH EFFECTS

Table 3-11. Reference Values of Parameters for the Compartment Model to Represent Time-dependent Particle Transport from the Human Respiratory Tract

Part A

		Clearance Rates for Insolu	ble Particles	
Pathway	From	То	Rate (d <sup>-1</sup> )	Half-time <sup>a</sup>
m <sub>1,4</sub>	$AI_1$	bb <sub>1</sub>	0.02	35 days
m <sub>2,4</sub>	$Al_2$	bb <sub>1</sub>	0.001	700 days
m <sub>3,4</sub>	$Al_3$	bb <sub>1</sub>	0.0001	7,000 days
m <sub>3,10</sub>	$AI_3$	$LN_TH$	0.00002	_
m <sub>4,7</sub>	bb <sub>1</sub>	$BB_1$	2	8 hours
m <sub>5,7</sub>	$bb_2$	$BB_1$	0.03	23 days
m <sub>6,10</sub>	$bb_seq$	$LN_TH$	0.01	70 days
m <sub>7,11</sub>	$BB_1$	ET <sub>2</sub>	10	100 minutes
m <sub>8,11</sub>	$BB_2$	ET <sub>2</sub>	0.03	23 days
m <sub>9,10</sub>	$BB_seq$	$LN_TH$	0.01	70 days
m <sub>11,15</sub>	ET <sub>2</sub>	GI tract	100	10 minutes
m <sub>12,13</sub>	$ET_seq$	$LN_{ET}$	0.001	700 days
m <sub>14,16</sub>	ET <sub>1</sub>	Environment	1	17 hours

See next page for Part B

Table 3-11. Reference Values of Parameters for the Compartment Model to Represent Time-dependent Particle Transport from the Human Respiratory Tract (continued)

Part B

Partition of	Partition of deposit in each region between compartments <sup>b</sup> Fraction of deposit in region assigned to compartment <sup>c</sup> ET <sub>2</sub> 0.9995  ET <sub>seq</sub> 0.0005					
Region or deposition site	Compartment					
ET <sub>2</sub>	ET <sub>2</sub>	0.9995				
	ET <sub>seq</sub>	0.0005				
ВВ	$BB_1$	0.993 <i>-f</i> <sub>s</sub>				
	$BB_2$	$f_{ m s}$				
	$BB_seq$	0.007				
bb	bb <sub>1</sub>	0.993- <i>f</i> <sub>s</sub>				
	$bb_2$	$f_{ m s}$				
	$bb_{seq}$	0.007				
Al	Al <sub>1</sub>	0.3				
	$Al_2$	0.6				
	$Al_3$	0.1				

<sup>&</sup>lt;sup>a</sup>The half-times are approximate since the reference values are specified for the particle transport rates and are rounded in units of day<sup>1</sup>. A half-time is not given for the transport rate from  $Al_3$  to  $LN_{TH}$ , since this rate was chosen to direct the required amount of material to the lymph nodes. The clearance half-time of compartment  $Al_3$  is determined by the sum of the clearance rates from it.

$$f_S = 0.5 \text{ for } d_{ae} \le 2.5\sqrt{\rho/\chi} \text{ } \mu m \text{ } and$$
  
 $f_S = 0.5e^{0.63(d_{ae}\sqrt{\rho/\chi} - 2.5)} \text{ } for d_{ae} > 2.5\sqrt{\rho/\chi} \text{ } \mu m$ 

where

 $f_s$  = fraction subject to slow clearance d<sub>ae</sub> = aerodynamic particle diameter/( $\mu$ m)

p = particle density (g/cm³) χ = particle shape factor

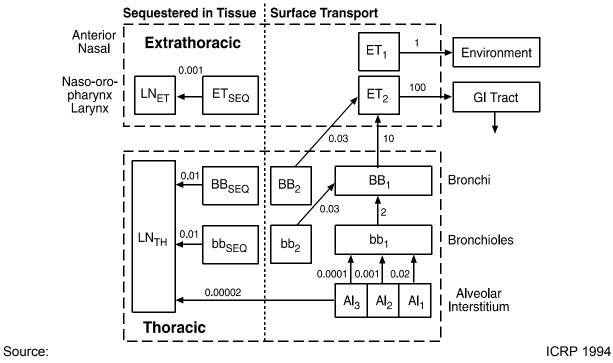
Al = alveolar-interstitial region; BB = bronchial region; bb = bronchiolar region; BB<sub>seq</sub> = compartment representing prolonged retention in airway walls of small fraction of particles deposited in the bronchial region; bb<sub>seq</sub> = compartment representing prolonged retention in airway walls of small fraction of particles deposited in the bronchiolar region; ET = extrathoracic region; ET<sub>seq</sub> = compartment representing prolonged retention in airway tissue of small fraction of particles deposited in the nasal passages; LN<sub>ET</sub> = lymphatics and lymph nodes that drain the extrathoracic region; LN<sub>TH</sub> = lymphatics and lymph nodes that drain the thoracic region

Source: ICRP 1994

bSee paragraph 181, Chapter 5 (ICRP 1994) for default values used for relating  $f_s$  to  $d_{ae}$ .

<sup>°</sup>It is assumed that  $f_s$  is size-dependent. For modeling purposes,  $f_s$  is taken to be:

Figure 3-8. Compartment Model to Represent Time-Dependent Particle
Transport in the Respiratory Tract



See Table 3-11 for abbreviations, rates, half-lives, and fractions by compartment

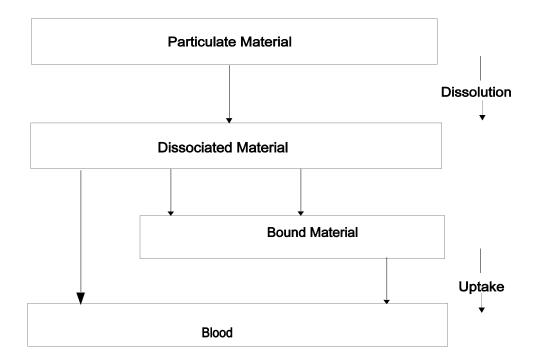
Ciliary action removes deposited particles from both the bronchi and bronchioles. Though it is generally thought that mucocilliary action rapidly transports most particles deposited here toward the pharynx, a fraction of these particles are cleared more slowly. Evidence for this is found in human studies. For humans, retention of particles deposited in the lungs (BB and bb) is apparently biphasic. The "slow" action of the cilia may remove as many as half of the bronchi- and bronchiole-deposited particles. In human bronchi and bronchiole regions, mucus moves more slowly the closer to the alveoli it is. For the faster compartment it has been estimated that it takes about 2 days for particles to travel from the bronchioles to the bronchi and 10 days from the bronchi to the pharynx. The second (slower) compartment is assumed to have approximately equal fractions deposited between BB<sub>2</sub> and bb<sub>2</sub> and both with clearance half-times estimated at 20 days. Particle size is a primary determinant of the fraction deposited in this slow thoracic compartment. A small fraction of particles deposited in the BB and bb regions is retained in the airway wall for even longer periods (BB<sub>seq</sub> and bb<sub>seq</sub>).

If particles reach and become deposited in the alveoli, they tend to stay imbedded in the fluid on the alveolar surface or move into the lymph nodes. The one mechanism by which particles are physically resuspended and removed from the AI region is coughing. For modeling purposes, the AI region is divided into 3 subcompartments to represent different clearance rates, all of which are slow.

In the alveolar-interstitial region, human lung clearance has been measured. The ICRP model uses 2 half-times to represent clearance: about 30% of the particles have a 30-day half-time, and the remaining 70% are given a half-time of several hundred days. Over time, the AI particle transport rate falls and some compounds have been found in lungs 10–50 years after exposure.

Absorption into Blood. The ICRP model assumes that absorption into blood occurs at equivalent rates in all parts of the respiratory tract, except in the anterior nasal passages (ET<sub>1</sub>), where no absorption occurs. It is essentially a 2-stage process, as shown in Figure 3-9. First, there is a dissociation (dissolution) of particles; then the dissolved molecules or ions diffuse across capillary walls and are taken up by the blood. Immediately following dissolution, rapid absorption is observed. For some elements, rapid absorption does not occur because of binding to respiratory-tract components. In the absence of specific data for specific compounds, the model uses the following default absorption rate values for those specific compounds that are classified as Types F (fast), M (medium), S (slow), and V (instantaneous):

Figure 3-9. The Human Respiratory Tract Model: Absorption into Blood



Source: ICRP 1994

- For Type F, there is rapid 100% absorption within 10 minutes of the material deposited in the BB, bb, and AI regions, and 50% of material deposited in ET<sub>2</sub>. Thus, for nose breathing, there is rapid absorption of approximately 25% of the deposit in ET and 50% for mouth breathing.
- C For Type M, about 70% of the deposit in AI reaches the blood eventually. There is rapid absorption of about 10% of the deposit in BB and bb, and 5% of material deposited in ET<sub>2</sub>. Thus, there is rapid absorption of approximately 2.5% of the deposit in ET for nose breathing, and 5% for mouth breathing.
- C For Type S, 0.1% is absorbed within 10 minutes and 99.9% is absorbed within 7,000 days, so there is little absorption from ET, BB, or bb, and about 10% of the deposit in AI reaches the blood eventually.
- C For Type V, complete absorption (100%) is considered to occur instantaneously.

ICRP (1995) considers the experimental and human data to support the following classifications: cobalt chloride and nitrate, Type F; cobalt oxides, Type M or S; cobalt in fused aluminosilicate or polystyrene, Type S; cobalt in mineral dusts such as fly ash and volcanic ash, Type M; cobalt metal and metal alloys, M or S. ICRP (1995) recommends assigning all cobalt aerosols to Type M in the absence of specific information supporting an alternative classification.

# ICRP (1993) Cobalt Biokinetics Model.

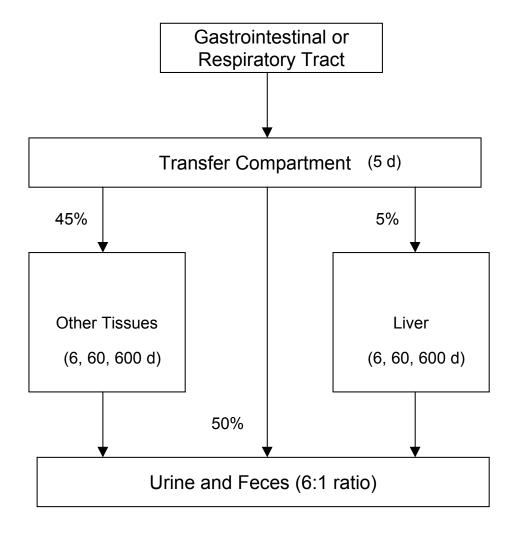
### Description of the model.

ICRP (1979,1993) developed a 3-compartment model of the kinetics of ingested cobalt in humans that is applicable to infants, children, adolescents, and adults. Absorption of ingested cobalt is assumed to be 60% in infants up to 3 months of age, 30% from 3 months to 15 years of age, and 10% after age 15 years. Absorbed cobalt is assumed to distribute as follows: 50% is excreted (urine and feces combined in a 6:1 ratio), 5% is transferred to the liver, and 45% is transferred to other tissues (see Figure 3-10). Elimination from tissue compartments is described by three first order rate constants representing slow, medium, and fast elimination pools with half-times of 6, 60, and 800 days, respectively. The elimination half-times are assumed to be constant with age.

#### Validation of the model.

The extent to which the ICRP model has been validated is not described in ICRP (1993).

Figure 3-10. ICRP Biokinetics Model for Cobalt



Absorbed cobalt enters a virtual transfer compartment from which unidirectional transfer to tissues is assumed to occur. Percentages shown are of the initial amounts absorbed. Numbers in parentheses are elimination half-times to urine and feces combined (d=days. Liver other tissues are assumed to have fast, medium, and slow elimination pools.

#### Risk assessment.

The model has been used to establish radiation dose equivalents (Sv/Bq) of ingested <sup>57</sup>Co, <sup>58</sup>Co, and <sup>60</sup>Co for ages 3 months to 70 years (ICRP 1993).

# Target tissues.

The model can be used to estimate the radiation dose from cobalt radionuclides to all major organs and can be applied to environmental and occupational exposures.

### Species extrapolation.

The model is designed for applications to human dosimetry and cannot be applied to other species without modification.

### Interroute extrapolation.

The model is designed to simulate oral exposures to cobalt and cannot be applied to other routes of exposure without modification.

#### 3.5 MECHANISMS OF ACTION

# 3.5.1 Pharmacokinetic Mechanisms

**Absorption.** Following inhalation exposure, the absorption of deposited cobalt compounds seems to be related to their biological solubility. Cobalt compounds deposit in the lungs based on their aerosol characteristics. Physiologically insoluble cobalt particles are generally cleared by phagocytosis and/or mucociliary transport and thus have a low systemic absorption. To some extent, cobalt particles may be dissolved within alveolar macrophages (Kreyling et al. 1990). More soluble forms of cobalt may enter the bloodstream through the alveolar or bronchial walls.

Following oral exposure, the absorption of cobalt varies with the amount given, with a greater dose leading to 4- to 20-fold greater fractional absorption (Smith et al. 1972). Nutritional status also seems to

be an important factor in cobalt absorption, with both overnight fasting and iron deficiency resulting in increased cobalt absorption (Smith et al. 1972; Sorbie et al. 1971; Valberg et al. 1969). It has been suggested that cobalt and iron share a common absorptive pathway in the intestines, though the cobalt absorption takes place without ferritin (Reuber et al. 1994; Schade et al. 1970; Thomson et al. 1971). Solubility of the cobalt compound is also an important factor regarding the absorption following oral exposure, with increasing solubility resulting in increasing absorption (Christensen et al. 1993). One study in humans showed that oral exposure to cobalt resulted in significantly higher urinary excretion in females relative to males (Christensen et al. 1993), but these results have not been verified by other studies. A complex, specific pathway exists for the absorption of vitamin B<sub>12</sub>, whereby the molecule interacts with several factors in the stomach and intestine to facilitate absorption (for review, see Russel-Jones and Alpers 1999).

Dermal absorption of cobalt compounds depends greatly on whether the skin is intact or damaged. Absorption through intact skin is very low, while absorption through damaged skin is much higher (Inaba and Suzuki-Yasumoto 1979; Lacy et al. 1996).

**Distribution.** As a component of vitamin B<sub>12</sub>, cobalt is found in most body tissues. Absorbed cobalt is transported throughout the body in the blood, with greatest levels found in liver, followed by the kidney (Ayala-Fierro et al. 1999; Greenberg et al. 1943; Gregus and Klaassen 1986; Patrick et al. 1989). Following inhalation exposure, significant levels of cobalt are found in the lungs of exposed humans and animals (Barnes et al. 1976; Brune et al. 1980; Collier et al. 1991; Gerhardsson et al. 1984; Hewitt 1988; Hillerdal and Hartung 1983; Kreyling et al. 1986; Kyono et al. 1992; Patrick et al. 1989; Talbot and Morgan 1989; Teraoka 1981). Within the lung, physiologically insoluble cobalt particles tend to be located within macrophages within the bronchial wall or in the interstitium close to the terminal bronchioli (Brune et al. 1980).

**Excretion.** Following inhalation exposure, the rate of urinary excretion appears to correlate with the rate of translocation of cobalt from the lungs to the blood, and the rate of fecal clearance with the rate of mechanical clearance of cobalt from the lungs to the gastrointestinal tract (Andre et al. 1989; Bailey et al. 1989; Collier et al. 1989; Kerfoot 1975; Kreyling et al. 1986, 1989; Palmes et al. 1959; Patrick et al. 1989; Talbot and Morgan 1989). Likewise, absorbed cobalt following oral exposure is rapidly removed from the body by excretion in the urine, and to a lesser extent the bile and feces, with fecal excretion being the primary method of excretion for physiologically insoluble cobalt compounds in both humans

and animals (Andre et al. 1989; Bailey et al. 1989; Collier et al. 1989; Harp and Scoular 1952; Paley et al. 1958; Patrick et al. 1989; Smith et al. 1972; Sorbie et al. 1971; Talbot and Morgan 1989; Valberg et al. 1969). The primary route for excretion following dermal exposure is the urine (Lacy et al. 1996).

## 3.5.2 Mechanisms of Toxicity

Stable Cobalt. The exact mechanisms by which cobalt exerts its effects on cells are not completely understood. However, a number of potential mechanisms have been identified. Several studies have demonstrated that hard metal, a metal alloy with a tungsten carbide and cobalt matrix, is considerably more toxic than either cobalt or tungsten carbide alone. A mechanism by which hard metal may exert its effects has been proposed by a group of Belgian researchers (Lasfargues et al. 1995; Lison et al. 1995, 1996). In this proposed mechanism, tungsten carbide, which is a very good conductor of electrons, facilitates the oxidation of cobalt metal to ionic cobalt (presumably Co<sup>2+</sup>) by transferring electrons from the cobalt atom to molecular oxygen adjacent to the tungsten carbide molecule. The result is an increased solubility of cobalt, relative to cobalt metal alone, and the generation of active oxygen species, both of which have been shown to occur following *in vivo* exposure to hard metal. The cobalt ions formed may be absorbed into the blood and transported throughout the body, where they may elicit effects by the above mechanisms. *In vitro* evidence for this mechanism includes the ability of hard metal particles, but neither cobalt nor tungsten carbide alone, to generate oxidant species and cause lipid peroxidation (Lison et al. 1995; Zanetti and Fubini 1997). Hard metal particles have also been shown to increase the levels of inducible nitric oxide synthase (iNOS), a gene responsive to oxidant stress (Rengasamy et al. 1999).

Another potential mechanism for cobalt toxicity is though oxidant-based and free radical-based processes. Exposure to soluble cobalt increases indices of oxidative stress, including diminished levels of reduced glutathione, increased levels of oxidized glutathione, and free-radical-induced DNA damage (Kasprzak et al. 1994; Lewis et al. 1991; Zhang et al. 1998a). Cobalt has been shown to generate oxygen radicals, including superoxide, both *in vitro* and *in vivo* (Kadiiska et al. 1989; Kawanishi et al. 1994; Moorhouse et al. 1985), through what may be a Fenton-type mechanism (Lloyd et al. 1997). *In vivo* exposure to cobalt in rats resulted in increased lipid peroxidation in the liver (Sunderman and Zaharia 1988). Exposure to cobalt results in accumulation in cardiac tissues, and is thought to stimulate carotid-body chemoreceptors, mimicking the action of hypoxia (Di Giulio et al. 1990, 1991; Hatori et al. 1993; Morelli et al. 1994). Cobalt administration to a neuroblastoma/glioma cell line resulted in an upregulation of opioid delta

receptors, through a mechanism similar to that of hypoxia (Mayfield et al. 1994). Exposure to cobalt also elicits effects on a number of genes known to be sensitive to oxidant status, including hypoxia-inducible factor 1, erythropoietin, vascular endothelial growth factor, catalase, and monooxygenase enzymes (Dalvi and Robbins 1978; Di Giulio et al. 1991; Goldberg et al. 1988, 1994; Ladoux and Frelin 1994; Legrum et al. 1979; Semenza et al. 1994; Yasukochi et al. 1974).

Soluble cobalt has also been shown to alter calcium influx into cells, functioning as a blocker of inorganic calcium channels (Henquin et al. 1983; Moger 1983; Yamatani et al. 1998). This mechanism has been linked to a reduction of steroidogenesis in isolated mouse Leydig cells (Moger 1983). Additionally, soluble cobalt has been shown to alter the inorganic calcium influx in liver cells after exposure to glucagon (Yamatani et al. 1998), and calcium influx into pancreatic  $\beta$  cells (Henquin et al. 1983) and isolated rat islets (Henquin and Lambert 1975). Cobalt may also affect neuromuscular transmission though antagonism with calcium (Weakly 1973).

Another potential mechanism of cobalt toxicity is relevant to cobalt cardiomyopathy. As mentioned previously, cobalt accumulated in the heart of beer drinkers. Microscopic analysis revealed fragmentation and degeneration of myofibers and aggregates of abnormal mitochondria (Ferrans et al. 1964). These mitochondrial changes are indicative of disturbances in energy production or utilization possibly related to cobalt effects on lipoic acid. Cobalt irreversibly chelates lipoic acids under aerobic conditions (Webb 1982). Lipoic acid is a required cofactor for oxidative decarboxylation of pyruvate to acetyl CoA and of α-ketoglutarate to succinate (Lehninger 1982). In the myocadrium of rats treated with cobalt, oxidation of pyruvate or fatty acids is impaired (Wiberg 1968).

A number of investigators have shown that cobalt ions can result in increased damage to DNA when coexposed with oxidants, such as UV radiation or  $H_2O_2$  (De Boeck et al. 1998; Hartwig et al. 1991; Nackerdien et al. 1991). It is believed that cobalt acts by inhibition of DNA repair, particularly the incision and polymerization steps (Asmuß et al. 2000; Kasten et al. 1997), accomplishing this through interaction with zinc finger DNA repair proteins (Asmuß et al. 2000; Sarkar 1995).

Another potentially important mechanism by which cobalt may exert effects is through its effects on heme and heme-containing enzymes. Cobalt is thought to inhibit heme synthesis *in vivo* by acting upon at least two different sites in the biosynthetic pathway: synthesis of 5-aminolevulinate and conversion of 5-aminolevulinate into heme (de Matteis and Gibbs 1977). This inhibitory activity might result in the

formation of cobalt protoporphyrin rather than heme (Sinclair et al. 1979). Cobalt treatment also stimulates heme oxidation in many organs, due to the induction of heme oxygenase (for review, see Sunderman 1987). Effects on heme synthesis may potentially affect a wide variety of heme-containing proteins, including monooxygenase enzymes (i.e., cytochromes P450), and catalase (Legrum et al. 1979; Yasukochi et al. 1974). Conversely, cobalt acts, through a mechanism believed to involve a heme-containing protein, to increase erythropoietin, which stimulates the production of red blood cells (Di Giulio et al. 1991; Goldberg et al. 1988). The regulatory mechanisms behind this apparent dichotomy have not been fully elucidated.

Another potential mechanism by which cobalt may exert its effects is through interactions with the immune system. Exposure of humans to cobalt by the inhalation and dermal routes have resulted in sensitization to cobalt (Alomar et al. 1985; Bencko et al. 1983; Dooms-Goossens et al. 1980; Fischer and Rystedt 1983; Goh et al. 1986; Kanerva et al. 1988; Marcussen 1963; Shirakawa et al. 1988, 1989; Valer et al. 1967). Exposure to inhaled cobalt chloride aerosols can precipitate an asthmatic attack in sensitized individuals (Shirakawa et al. 1989), suggesting cobalt sensitization as one mechanism by which cobaltinduced asthma may be produced. IgE and IgA antibodies specific to cobalt have been reported in humans (Bencko et al. 1983; Shirakawa et al. 1988, 1989). There is evidence that cobalt sensitivity in humans may to be regulated by T-lymphocytes (Katsarou et al. 1997). A human helper T-lymphocyte cell line specific for cobalt (CoCl<sub>2</sub>) has been established (Löfström and Wigzell 1986). Cobalt may also interact directly with immunologic proteins, such as antibodies or Fc receptors, to result in immunosensitization (Cirla 1994). In vitro, cobalt(III) has been shown to reduce the proliferation of both B and T lymphocytes, as well as the release of the cytokines IL-2, IL-6, and IFN-Gamma (Wang et al. 1996). Interrelationships exist between nickel and cobalt sensitization (Bencko et al. 1983; Rystedt and Fisher 1983). In guinea pigs, nickel and cobalt sensitization appear to be interrelated and mutually enhancing (Lammintausta et al. 1985), though cross-reactivity was not reported to occur.

Cobalt has been shown to have a number of effects on glucose metabolism. Treatment of animals with cobalt results in a depression of serum (Eaton and Pommer 1973; Ybarra et al. 1997) or tissue (Wiberg 1968) glucose levels. In rats made diabetic by pretreatment with streptozotocin this depression was persistent, whereas it was transient in normal rats (Ybarra et al. 1997). Many of the effects of cobalt on glucose metabolism are thought to result from alterations in the expression of the *glut* family of glucose transport proteins, a family of facilitative Na<sup>+</sup>-independent transport proteins thought to mediate non-insulin-dependent transport of glucose. Exposure to soluble cobalt results in increased expression of

these genes, particularly GLUT1, in cells of the liver, kidney cortex, myocardium, skeletal muscle, and cerebrum (Behrooz and Ismail-Beigi 1997; Ybarra et al. 1997). Cobalt also reduces the amount of glucose produced in liver cells following stimulation with glucagon (Eaton and Pommer 1973; Yamatani et al. 1998), as well as reducing insulin release in isolated rat islets (Henquin and Lambert 1975).

**Radioactive Cobalt.** Due to the nature of its ionizing radiation, radioactive cobalt can present a health hazard. Highly-penetrating gamma emissions are the major source of damage to tissues and internal organs following external exposure to radioactive cobalt isotopes. If radioactive cobalt is internalized, nearby tissues are at highest risk for damage due to the release of beta particles. In either case, exposure to ionizing radiation results in an increased risk of cellular damage. Both beta and gamma radiations are capable of producing ionization events when they hit cellular molecules, including DNA, RNA, or lipids. Ionized molecules within irradiated cells may be repaired quickly to prevent further damage. On the other hand, irreparable damage may be imposed on cellular materials, such as DNA, which might ultimately result in either cell death or the formation of cancerous tumors. Very large acute radiation doses can damage or kill enough cells to cause the disruption of organ systems, resulting in acute radiation syndrome or even death. Human and animal data indicate that sufficiently high exposures to cobalt radiation can result in adverse effects such as reduced fertility, abnormal development, genotoxicity, pulmonary fibrosis, gastrointestinal atrophy and fibrosis, hematological and lymphoreticular disorders, cancer, and death (Chang et al. 1999b; Davis et al. 1992; Dinehart et al. 1991; Hashimoto and Mitsuyasu 1967; Klener et al. 1986; Libshitz 1993; Myskowski and Safai 1981; Rauscher and Bauchinger, 1983; Roschler and Woodard 1969; Roswit and White 1977; Stavem et al. 1985; Van Oort et al. 1984). For a more complete discussion of the mechanisms associated with the toxic effects of ionizing radiation, refer to Chapter 5 of the Toxicological Profile for Ionizing Radiation (ATSDR 1999).

#### 3.5.3 Animal-to-Human Extrapolations

Bailey et al. (1989) reported a wide variation across species, including man, in the retention and clearance of inhaled physiologically insoluble <sup>57</sup>Co particles (see Table 3-8), noting that this variation illustrates the potential difficulty of extrapolating the results of animal lung retention experiments to man even qualitatively. Species differences in absorption of physiologically insoluble cobalt oxide following oral exposure do not appear to exist (Bailey et al. 1989), though humans were not examined. Absorption of soluble cobalt compounds is greater in rats (13–34%) than in dairy cows (1–2%) and guinea pigs (4–5%) following oral exposure (Ayala-Fierro et al. 1999; Barnaby et al. 1968; Hollins and McCullough 1971;

Kirchgessner et al. 1994; Naylor and Harrison 1995; Schade et al. 1970; Taylor 1962; van Bruwaene et al. 1984).

### 3.6 ENDOCRINE DISRUPTION

Recently, attention has focused on the potential hazardous effects of certain chemicals on the endocrine system because of the ability of these chemicals to mimic or block endogenous hormones, or otherwise interfere with the normal function of the endocrine system. Chemicals with this type of activity are most commonly referred to as endocrine disruptors. Some scientists believe that chemicals with the ability to disrupt the endocrine system are a potential threat to the health of humans, aquatic animals, and wildlife. Others believe that endocrine disrupting chemicals do not pose a significant health risk, particularly in light of the fact that hormone mimics exist in the natural environment. Examples of natural hormone mimics are the isoflavinoid phytoestrogens (Adlercreutz 1995; Livingston 1978; Mayr et al. 1992). These compounds are derived from plants and are similar in structure and action as endogenous estrogen. While there is some controversy over the public health significance of endocrine disrupting chemicals, it is agreed that the potential exists for these compounds to affect the synthesis, secretion, transport, binding, action, or elimination of natural hormones in the body that are responsible for the maintenance of homeostasis, reproduction, development, and/or behavior (EPA 1997a). As a result, endocrine disruptors may play a role in the disruption of sexual function, immune suppression, and neurobehavioral function. Endocrine disruption is also thought to be involved in the induction of breast, testicular, and prostate cancers, as well as endometriosis (Berger 1994; Giwercman et al. 1993; Hoel et al. 1992).

Stable Cobalt. A group of female workers occupationally exposed to a semisoluble cobalt glaze (cobalt-zinc silicate) showed significantly elevated levels of serum thyroxine ( $T_4$ ) and free thyroxine, but no change in  $T_3$  levels (Prescott et al. 1992). In contrast to this, Swennen et al. (1993) reported no significant change in serum  $T_4$  levels, but a significant reduction in serum  $T_3$  in workers occupationally exposed to cobalt oxides, cobalt salts, and cobalt metal.

Patients (n=12) injected with a single dose of radioactive iodine, and then treated 48 hours later with 1 mg cobalt/kg/day as cobalt chloride for 2 weeks resulted in a greatly reduced uptake of radioactive iodine by the thyroid in 1 week, with uptake nearing 0 by the second week (Roche and Layrisse 1956). When the cobalt treatment ended, the uptake values returned to normal. The decrease of radioactive iodine uptake

found in patients administered 0.54 mg cobalt/kg/day for 10–25 days prior to iodine injection was found to result from cobalt blocking the organic binding of iodine (Paley et al. 1958).

Roy et al. (1968) reported on 20 Québécois patients who died of beer drinkers' myocardosis. Of these, 14 thyroids were available for examination. Three of those were normal, and the other 11 formed the basis of the study. "Abnormal" thyroids did not show gross changes, but upon histologic examination, they showed irregular follicle morphology and decreased follicular size.

Kriss et al. (1955) reported on five patients who had been receiving cobalt therapy for sickle-cell anemia or renal amyloidosis. Three of five developed goiter, one severe, while four of five showed microscopic alterations of the thyroid gland. Two of the patients died from non-cobalt-related causes, while the other three recovered once cobalt treatment was removed. No other studies examining the endocrine effects of stable cobalt in humans were located.

In various species of animals, parenteral administration of cobalt resulted in cytotoxic effects on the alpha cells of the pancreas (Beskid 1963; Goldner et al. 1952; Hakanson et al. 1974; Lacy and Cardeza 1958; Lazarus et al. 1953; Van Campenhout 1955). Because this effect has never been reported in humans or animals following inhalation, oral, or dermal exposure to cobalt, the relevance of the effect to humans is not known. NTP (1998; Bucher et al. 1999) reported increased incidence of pheochromocytoma, a tumor of the adrenal medulla, in female rats exposed to 1.14 mg cobalt/m³ for 2 years, but did not measure any other endocrine effects. Female mice exposed to 26 mg cobalt/kg-day in the drinking water for up to 45 days showed histopathological changes to the thyroid gland (Shrivastava et al. 1996). Cobalt significantly stimulated serum testosterone in mice treated orally with 23 mg cobalt/kg as cobalt chloride, though no dose-response relationship was present (Pedigo et al. 1988).

No other studies examining the endocrine effects of stable cobalt in animals were located.

Moger (1983) exposed primary cultures of mouse Leydig cells to 0–2.5 mM cobalt as cobalt for 3 hours, and measured the effects on androgen production. Cobalt exposure caused a dose-related decrease in both basal and LH-stimulated androgen production, with no effects on protein synthesis. The author suggested that these effects are the result of cobalt inhibition of calcium influx across the plasma membrane.

**Radioactive Cobalt.** Prager et al. (1972) reported that 5 of 23 patients receiving cobalt radiotherapy (3,900-4,600 rad, 39-46 Gy) for Hodgkin's disease developed hypothyroidism, with substantial decreases in levels of  $T_4$  relative to patients with normal thyroids. No other studies examining the endocrine effects of radioactive cobalt exposure, either internal or external, in humans were located.

Whole-body acute exposure of rats to 330 rad (3.3 Gy) did not effect FSH, LH, or testosterone levels (Cunningham and Huckins 1978). Similarly, male Wistar rats exposed to a single dose of 80 rad (0.8 Gy) of testicular radiation showed no changes in FSH, LH, prolactin, or testosterone (Laporte et al. 1985). No other studies examining the endocrine effects of radioactive cobalt exposure, either internal or external, in animals were located.

#### 3.7 CHILDREN'S SUSCEPTIBILITY

This section discusses potential health effects from exposures during the period from conception to maturity at 18 years of age in humans, when all biological systems will have fully developed. Potential effects on offspring resulting from exposures of parental germ cells are considered, as well as any indirect effects on the fetus and neonate resulting from maternal exposure during gestation and lactation. Relevant animal and *in vitro* models are also discussed.

Children are not small adults. They differ from adults in their exposures and may differ in their susceptibility to hazardous chemicals. Children's unique physiology and behavior can influence the extent of their exposure. Exposures of children are discussed in Section 6.6 Exposures of Children.

Children sometimes differ from adults in their susceptibility to hazardous chemicals, but whether there is a difference depends on the chemical (Guzelian et al. 1992; NRC 1993). Children may be more or less susceptible than adults to health effects, and the relationship may change with developmental age (Guzelian et al. 1992; NRC 1993). Vulnerability often depends on developmental stage. There are critical periods of structural and functional development during both prenatal and postnatal life and a particular structure or function will be most sensitive to disruption during its critical period(s). Damage may not be evident until a later stage of development. There are often differences in pharmacokinetics and metabolism between children and adults. For example, absorption may be different in neonates because of the immaturity of their gastrointestinal tract and their larger skin surface area in proportion to body weight (Morselli et al. 1980; NRC 1993); the gastrointestinal absorption of lead is greatest in infants

and young children (Ziegler et al. 1978). Distribution of xenobiotics may be different; for example, infants have a larger proportion of their bodies as extracellular water and their brains and livers are proportionately larger (Altman and Dittmer 1974; Fomon 1966; Fomon et al. 1982; Owen and Brozek 1966; Widdowson and Dickerson 1964). The infant also has an immature blood-brain barrier (Adinolfi 1985; Johanson 1980) and probably an immature blood-testis barrier (Setchell and Waites 1975). Many xenobiotic metabolizing enzymes have distinctive developmental patterns. At various stages of growth and development, levels of particular enzymes may be higher or lower than those of adults, and sometimes unique enzymes may exist at particular developmental stages (Komori et al. 1990; Leeder and Kearns 1997; NRC 1993; Vieira et al. 1996). Whether differences in xenobiotic metabolism make the child more or less susceptible also depends on whether the relevant enzymes are involved in activation of the parent compound to its toxic form or in detoxification. There may also be differences in excretion, particularly in newborns who all have a low glomerular filtration rate and have not developed efficient tubular secretion and resorption capacities (Altman and Dittmer 1974; NRC 1993; West et al. 1948). Children and adults may differ in their capacity to repair damage from chemical insults. Children also have a longer remaining lifetime in which to express damage from chemicals; this potential is particularly relevant to cancer.

Certain characteristics of the developing human may increase exposure or susceptibility, whereas others may decrease susceptibility to the same chemical. For example, although infants breathe more air per kilogram of body weight than adults breathe, this difference might be somewhat counterbalanced by their alveoli being less developed, which results in a disproportionately smaller surface area for alveolar absorption (NRC 1993).

Though human data are lacking, animal studies have suggested several differences in pharmacokinetic behavior of cobalt compounds between children and adults. Following inhalation exposure to Co<sub>3</sub>O<sub>4</sub>, deposition tended to increase with age, though no significant differences were reported (Collier et al. 1991). The youngest animals exposed (3 weeks postnatal) had the lowest fractional retention 182 days postexposure, though no differences were seen at day 7 or 83. The authors attributed this to an faster rate of translocation of cobalt from the lung to the blood, which could enhance subsequent excretion. Naylor and Harrison (1995) reported that in rats and guinea pigs, fractional absorption of cobalt following oral exposure was highest at 1 day after birth, and diminished rapidly with time thereafter. Collier et al. (1991) reported no difference in absorption of cobalt nitrate following oral exposure to animals aged

3–46 weeks, which is in agreement with the results of the later portion of the Naylor and Harrison (1995) study. No PBPK models specific for cobalt exposures to children were located.

Once in the bloodstream, soluble cobalt compounds have been shown, in animal studies, to cross the placenta and enter the fetus. Twenty-four hours after intravenous injection of cobalt chloride in rats, 0.14% of the dose was found in the fetus, 0.19% in the chorioallantoic placenta, and 0.22% in the yolk sac (Zylicz et al. 1975). Several studies (Nishimura et al. 1978; Zylicz et al. 1975, 1976) have demonstrated that the amount of cobalt crossing the placenta following intravenous injection is greater later gestation stages, though the percent of the maternal dose reaching the fetus is still relatively low (in <1% of the maternal dose). The fetal uptake of cobalt following intravenous administration to the mother was increased when the cobalt was given as cyanocobalmin, relative to cobalt chloride (~5% of the maternal dose for cyanocobalmin, compared to <1% for cobalt chloride) (Nishimura et al. 1978), indicating that the form of the cobalt compound may affect its availability to the fetus.

Cobalt has been detected in human breast milk (Byczkowski et al. 1994; Kratchler et al. 1998). In general, physiological concentrations of cobalt in breast milk are very low, on the order of parts per billion (Byczkowski et al. 1994). Animals studies are in agreement with this observation. By day 70 post-exposure in lactating dairy cows orally exposed to cobalt chloride, the milk contained 0.012% of the dose (van Bruwaene et al. 1984). One to two percent of cobalt given intravenously to mother rats as cyanocobalmin was transferred to offspring via the breast milk (Nishimura et al. 1978).

Health Effects from Exposure to Stable Cobalt. Available data have not clearly defined whether children are at greater risk from exposure to stable cobalt than adults. Studies in adult humans have identified several health effects of cobalt compounds following inhalation, oral, or dermal exposure. Data on effects of cobalt in children following inhalation exposures are lacking. Jacobziner and Raybin (1961) reported on two cases of children who had accidentally ingested unknown amounts of cobalt chloride; a 19-month-old male died approximately 6.5 hours after ingestion, whereas a 3-year-old male was given medical treatment and showed no symptoms after ingestion. Patch testing of children aged 4–14 years revealed a 13.3% dermal sensitization rate to cobalt chloride (Romaguera and Vilaplana 1998). More girls reacted positively than boys, which the authors attributed to the wearing of costume jewelry, which often contains cobalt, and the resulting exposure.

Offspring of mice intravenously injected with approximately 1.2 mg cobalt/kg at day 8 of gestation, but not at day 3, showed a significant increase in the number of skeletons with delayed ossification (Wilde 1984). Other studies, however, have not shown developmental effects of stable cobalt compounds, or have shown effects only at maternally toxic doses (Domingo et al. 1985b; Paternian et al. 1988; Seidenberg 1986).

Health Effects from Exposure to Radioactive Cobalt. No studies of human children exposed to radioactive cobalt or cobalt radiation were located. As rapidly-dividing cells are more sensitive to radiation, the developing fetus and growing children are expected to be more sensitive to cobalt radiation than adults.

Animal studies have shown that exposures to external radiation from cobalt isotopes (as low as 10 rad [0.1 Gy] in mice) may have a dramatic effect on the developing fetus (see Section 3.2.4.6 and ATSDR 1999). Exposure duration, gestational day, and dose all influence the effect of cobalt radiation on the developing organism. Radiation exposure to very young dogs (80 rad [0.8 Gy] on day 2 or 70 postpartum) has resulted in an increased incidence of diabetes mellitus, renal disease, and cancer (Benjamin et al. 1998a, 1998b).

### 3.8 BIOMARKERS OF EXPOSURE AND EFFECT

Biomarkers are broadly defined as indicators signaling events in biologic systems or samples. They have been classified as markers of exposure, markers of effect, and markers of susceptibility (NAS/NRC 1989).

Due to a nascent understanding of the use and interpretation of biomarkers, implementation of biomarkers as tools of exposure in the general population is very limited. A biomarker of exposure is a xenobiotic substance or its metabolite(s) or the product of an interaction between a xenobiotic agent and some target molecule(s) or cell(s) that is measured within a compartment of an organism (NAS/NRC 1989). The preferred biomarkers of exposure are generally the substance itself or substance-specific metabolites in readily obtainable body fluid(s), or excreta. However, several factors can confound the use and interpretation of biomarkers of exposure. The body burden of a substance may be the result of exposures from more than one source. The substance being measured may be a metabolite of another xenobiotic substance (e.g., high urinary levels of phenol can result from exposure to several different aromatic

compounds). Depending on the properties of the substance (e.g., biologic half-life) and environmental conditions (e.g., duration and route of exposure), the substance and all of its metabolites may have left the body by the time samples can be taken. It may be difficult to identify individuals exposed to hazardous substances that are commonly found in body tissues and fluids (e.g., essential mineral nutrients such as copper, zinc, and selenium). Biomarkers of exposure to cobalt are discussed in Section 3.8.2.

Biomarkers of effect are defined as any measurable biochemical, physiologic, or other alteration within an organism that, depending on magnitude, can be recognized as an established or potential health impairment or disease (NAS/NRC 1989). This definition encompasses biochemical or cellular signals of tissue dysfunction (e.g., increased liver enzyme activity or pathologic changes in female genital epithelial cells), as well as physiologic signs of dysfunction such as increased blood pressure or decreased lung capacity. Note that these markers are not often substance specific. They also may not be directly adverse, but can indicate potential health impairment (e.g., DNA adducts). Biomarkers of effects caused by cobalt are discussed in Section 3.8.2.

A biomarker of susceptibility is an indicator of an inherent or acquired limitation of an organism's ability to respond to the challenge of exposure to a specific xenobiotic substance. It can be an intrinsic genetic or other characteristic or a preexisting disease that results in an increase in absorbed dose, a decrease in the biologically effective dose, or a target tissue response. If biomarkers of susceptibility exist, they are discussed in Section 3.10 "Populations That Are Unusually Susceptible".

# 3.8.1 Biomarkers Used to Identify or Quantify Exposure to Cobalt

Biomonitoring data exist that demonstrate a positive correlation between occupational exposure levels of cobalt and the levels of cobalt in both the urine and blood (Table 3-12) (Alexandersson 1988; Ichikawa et al. 1985; Lison et al. 1994; Nemery et al. 1992; Scansetti et al. 1985). Available studies of unexposed humans have reported cobalt blood levels of 0.05–0.19 μg/dL, and urinary cobalt levels of 0.04–2 μg/dL (Alexandersson 1988; Ichikawa et al. 1985). Figure 3-11 graphically presents the cobalt exposure data and cobalt in blood data presented in Table 3-12 (Ichikawa et al. 1985). The highest excretion rate of cobalt in urine occurs during the first 24 hours after short-term exposure; therefore, subjects should be tested quickly to assess if cobalt exposure has occurred (Alexandersson 1988). Occupational exposure to 0.1 mg/m³ cobalt resulted in blood levels of cobalt ranging (95% CI) from 0.57 to 0.79 μg/dL, compared

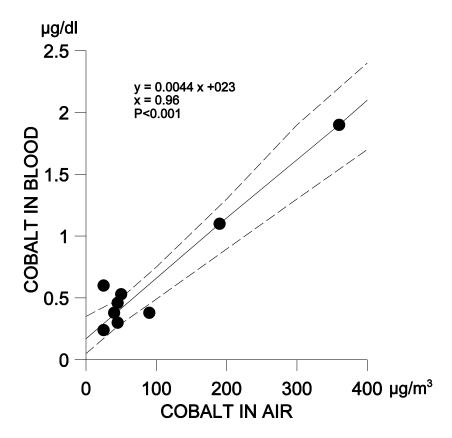
Table 3-12. Cobalt Exposure Concentrations and Amounts in the Blood and Urine of Subjects Examined<sup>a</sup>

Subjects		Numb er	Cobalt in air <sup>b</sup> mean±SD μg/m³			Cobalt in blood <sup>b</sup> mean±SD μg/dL		Cobalt in urine <sup>b</sup> mean±SD μg/L	
Powder handle	rs	2	186±10 8	(110–262)	1.08±0. 28	(0.88–1.28)	148±13	(138–158)	
Rubber press of	operators	6	367±32 4	(92–859)	1.87±1. 96	(0.40–5.30)	235±18 2	(41–392)	
Automatic pres	s operators	11	56±60	(9–210)	0.57±0. 53	(0.10–0.95)	34±43	(4–73)	
Shapers (lathin	g)	7	33±15	(15–62)	0.67±0. 44	(0.14–1.34)	33±30	(11–95)	
Shapers (sawir	ng)	21	50±35	(8–144)	0.52±0. 31	(0.15–1.15)	41±60	(6–266)	
Sintering worke	ers	21	28±30	(4–145)	0.26±0. 10	(0.09–0.45)	10±10	(2–46)	
Wet grinders	Α	27	44±48	(4–227)	0.42±0. 31	(0.10–1.30)	35±34	(2–180)	
	В	18	45±50	(3–161)	0.33±0. 10	(0.16–0.52)	19±15	(2–67)	
	С	12	92±92	(15–291)	0.43±0. 39	(0.12–1.90)	68±87	(3–265)	
	D	25	44±54	(3–205)	0.35±0. 20	(0.10–1.00)	17±16	(1–69)	
Workers using	respirators	25	317±30 7	(7–1,203)	0.65±0. 86	(0.20–3.90)	26±30	(1–119)	
Office workers		20	No data		0.19±0. 11	(0.08–0.40)	2±1	(1–4)	

<sup>&</sup>lt;sup>a</sup>Adapted from Ichikawa et al. 1985

<sup>&</sup>lt;sup>b</sup>The range of each value is given in parentheses.

Figure 3-11. Relation Between Mean Cobalt Exposure and Mean Blood Concentration of Cobalt in Exposed Workers\*



<sup>\*</sup>Adapted from Ichikawa et al. 1985

to  $0.19 \,\mu\text{g/dL}$  in unexposed workers, and urinary levels from 59 to 78  $\mu\text{g/L}$ , compared to  $2 \,\mu\text{g/dL}$  in unexposed workers (Ichikawa et al. 1985). Correlations between recent exposure and cobalt levels in the blood or urine are more consistent for soluble cobalt compounds (metal, salts, and hard metals), while blood and/or urinary cobalt levels are less reflective of recent exposure for less soluble compounds (cobalt oxides) (Lison et al. 1994).

Sensitive serum protein responses were found in animals exposed to cobalt at levels below those necessary to produce hematopoietic effects (Stokinger and Wagner 1958). These serum protein responses included an increase in alpha globulin fractions of serum proteins and associated serum neuraminic acid. The responses were observed in rabbits and dogs following both inhalation and injection of cobalt. The authors indicated that this increase was a unique response to cobalt exposure. The characteristics of the response were similar to those of the erythropoietic response found following exposure to higher levels of cobalt; the response is delayed, does not occur in all animals within a given exposure group, is not of great magnitude, and is not persistent (Stokinger and Wagner 1958).

Biomarkers specific for exposure to cobalt radioisotopes have not been reported.

# 3.8.2 Biomarkers Used to Characterize Effects Caused by Cobalt

Sensitization to cobalt results in cobalt-specific changes in serum antibodies (IgE and IgA) that may be monitored to determine if exposure to cobalt has occurred (Bencko et al. 1983; Shirakawa et al. 1988, 1989).

No biomarkers specific for effects of radioactive cobalt isotopes have been reported. Biomarkers for response to ionizing radiation are discussed in ATSDR (1999).

#### 3.9 INTERACTIONS WITH OTHER CHEMICALS

A major medical use of cobalt is in combination with bleomycin, an antineoplastic antibiotic, as a tumor-localizing and therapeutic agent (Goodwin and Meares 1976; Hansen et al. 1976; Kapstad 1978, 1979). The anti-tumor effects of the two agents are amplified when given in combination with each other. The complex, wherein cobalt is coordinately bound to the bleomycin molecule, is intravenously injected and acts by binding to and cleaving the DNA in the tumor cells (Kakinuma and Orii 1982).

The interaction of cobalt with various chelators has been investigated in animals for mitigation of the toxicity of cobalt (Baker et al. 1987; Domingo et al. 1983; Llobet et al. 1988). Glutathione, N-acetyl-L-cysteine (NAC), and diethylenetriaminepentaacetic acid (DTPA), administered to rats previously exposed to cobalt, significantly increased urinary excretion of cobalt, while EDTA, NAC, and 2,3-dimercaptosuccinic acid (DMSA) increased fecal excretion. NAC was the most effective chelator because it increased both urinary and fecal excretion of cobalt while decreasing its levels in liver and spleen (Llobet et al. 1988). Cysteine, also acting as a chelator, mitigated the toxicity of cobalt when both chemicals were given to chicks in the feed (Baker et al. 1987).

A number of studies have suggested an association between cobalt ions and calcium ions. Soluble cobalt has also been shown to alter calcium influx into cells, functioning as a blocker of inorganic calcium channels (Henquin et al. 1983; Moger 1983; Yamatani et al. 1998). This mechanism has been linked to a reduction of steroidogenesis in isolated mouse Leydig cells (Moger 1983). Additionally, soluble cobalt has been shown to alter the inorganic calcium influx in liver cells after exposure to glucagon (Yamatani et al. 1998), and calcium influx into pancreatic  $\beta$  cells (Henquin et al. 1983) and isolated rat islets (Henquin and Lambert 1975). Cobalt may also affect neuromuscular transmission though antagonism with calcium (Weakly 1973).

Hard metal, consisting of 5–10% cobalt with the balance being tungsten carbide, has been shown to be considerably more toxic than cobalt alone, resulting from interactions between particles of cobalt metal and tungsten carbide particles. The mechanisms responsible for this interaction are discussed in Section 3.5.2.

An interrelationship between cobalt and nickel sensitization has been reported in individuals exposed to the two metals (Rystedt and Fisher 1983; Veien et al. 1987). It was concluded that the combination of nickel sensitivity and irritant eczema resulted in a high risk for developing an allergy to cobalt.

#### 3.10 POPULATIONS THAT ARE UNUSUALLY SUSCEPTIBLE

A susceptible population will exhibit a different or enhanced response to cobalt than will most persons exposed to the same level of cobalt in the environment. Reasons may include genetic makeup, age, health and nutritional status, and exposure to other toxic substances (e.g., cigarette smoke). These parameters result in reduced detoxification or excretion of cobalt, or compromised function of organs affected by

cobalt. Populations who are at greater risk due to their unusually high exposure to cobalt are discussed in Section 6.7, Populations with Potentially High Exposures.

Individuals that are already sensitized to cobalt may be unusually susceptible because cobalt exposure may trigger asthmatic attacks (Shirakawa et al. 1988, 1989). Sensitization to cobalt results in cobalt-specific changes in serum antibodies (IgE and IgA) (Bencko et al. 1983; Shirakawa et al. 1988, 1989). Potolicchio et al. (1997, 1999) have suggested that individuals with a polymorphism in the HLA-DP gene (presence of glutamate 69 in the β chain) may be more susceptible to hard metal lung disease. Individuals with ongoing respiratory illness may also be more susceptible to the effects of inhaled cobalt. Following oral exposure, individuals with iron deficiency may be more at risk, as animal studies have shown an increased absorption of cobalt compounds in iron-deficient animals (Reuber et al. 1994; Schade et al. 1970). Studies of beer-cobalt cardiomyopathy have suggested that individuals with high alcohol consumption may be more susceptible to health effects of cobalt (Alexander 1969, 1972; Morin et al. 1971).

Ionizing radiation has greater effects on rapidly-dividing cells. The most sensitive population to exposure to cobalt radiation is likely to be the developing fetus, as even moderate exposures to cobalt radiation have been shown to cause dramatic effects on the developing fetus in animal studies (see Section 3.2.4.6), Likewise, growing children are likely to be more susceptible to cobalt radiation than adults, and people who are immunocompromised, have existing lung diseases, or who have defects in genetic repair enzymes would be expected to show an increased susceptibility to cobalt radiation.

#### 3.11 METHODS FOR REDUCING TOXIC EFFECTS

This section will describe clinical practice and research concerning methods for reducing toxic effects of exposure to cobalt. However, because some of the treatments discussed may be experimental and unproven, this section should not be used as a guide for treatment of exposures to cobalt. When specific exposures have occurred, poison control centers and medical toxicologists should be consulted for medical advice. The following texts provide specific information about treatment following exposures to cobalt:

Ellenhorn MJ, Schonwald S, Ordog G, et al., eds. 1997. Medical toxicology: Diagnosis and treatment of human poisoning. 2<sup>nd</sup> edition. Baltimore, MD: Williams & Wilkins. 1682-1723.

Goldfrank, LR, Flomenbaum, NE, Lewin, NA, et al. eds. 1998. Toxicological emergencies. 6<sup>th</sup> edition. Connecticut: Appleton & Lange. 481t, 489, 490t, 1338-1339.

REAC/TS. Radiation Emergency Assistance Center/Training Site. Online at www.orau.gov/reacts/.

# 3.11.1 Reducing Peak Absorption Following Exposure

General management and treatment of patients following acute exposure to cobalt includes removal of the victim from the contaminated area, and removal and isolation of contaminated clothing, jewelry, and shoes (Bronstein and Currance 1988; Stutz and Janusz 1988). The excess solid contaminant is gently brushed away, and excess liquids blotted with absorbent material. If the victim is in respiratory distress, ventilation assistance is provided and oxygen is administered. Measures that are appropriate to the route of exposure are then taken to remove cobalt from the body. Following ocular exposure, the eyes are immediately flushed thoroughly with water. Skin is washed immediately with soap or mild detergent and water. Following ingestion of cobalt, two conflicting forms of treatment have been recommended. Stutz and Janusz (1988) recommend that victims over 1-year-old be given ipecac, followed by activated charcoal (after vomiting). A cathartic, such as magnesium sulfate in water, is then administered to adults and children. Bronstein and Currance (1988) recommend that the victim be given water for dilution of the cobalt; however, they recommend that emetics not be administered. Following all routes of exposure, victims are monitored for pulmonary edema, circulatory collapse, and shock, and treated as necessary.

# 3.11.2 Reducing Body Burden

Chelation therapy with EDTA or dimercaprol can be effectively used if necessary (Goldfrank et al. 1990; Haddad and Winchester 1990; Stutz and Janusz 1988). Animal studies have investigated the effectiveness of various chelating agents for mitigating the toxicity of cobalt (Chapter 2) (Baker et al. 1987; Domingo et al. 1983; Llobet et al. 1988). NAC was found to be the most effective chelator because it increased both urinary and fecal excretion of cobalt as well as decreased the levels of cobalt in the liver and spleen (Llobet et al. 1988). For more complete information on treatment of specific symptoms, refer to Bronstein and Currance (1988) and Stutz and Janusz (1988). These chelators react chemically with cobalt, so they are effective for both stable and radioactive cobalt isotopes.

#### 3.11.3 Interfering with the Mechanism of Action for Toxic Effects

No studies were located in humans or animals regarding interfering with the mechanism of action of stable or radioactive cobalt compounds.

#### 3.12 ADEQUACY OF THE DATABASE

Section 104(i)(5) of CERCLA, as amended, directs the Administrator of ATSDR (in consultation with the Administrator of EPA and agencies and programs of the Public Health Service) to assess whether adequate information on the health effects of cobalt is available. Where adequate information is not available, ATSDR, in conjunction with the National Toxicology Program (NTP), is required to assure the initiation of a program of research designed to determine the health effects (and techniques for developing methods to determine such health effects) of cobalt.

The following categories of possible data needs have been identified by a joint team of scientists from ATSDR, NTP, and EPA. They are defined as substance-specific informational needs that if met would reduce the uncertainties of human health assessment. This definition should not be interpreted to mean that all data needs discussed in this section must be filled. In the future, the identified data needs will be evaluated and prioritized, and a substance-specific research agenda will be proposed.

# 3.12.1 Existing Information on Health Effects of Cobalt

The existing data on health effects of inhalation, oral, and dermal exposure of humans and animals to cobalt are summarized in Figure 3-12 for stable cobalt and Figure 3-13 for radioactive cobalt. The purpose of these figure is to illustrate the existing information concerning the health effects of cobalt. Each dot in the figure indicates that one or more studies provide information associated with that particular effect. The dot does not necessarily imply anything about the quality of the study or studies, nor should missing information in this figure be interpreted as a "data need". A data need, as defined in ATSDR's *Decision Guide for Identifying Substance-Specific Data Needs Related to Toxicological Profiles* (ATSDR 1989), is substance-specific information necessary to conduct comprehensive public health assessments. Generally, ATSDR defines a data gap more broadly as any substance-specific information missing from the scientific literature.

Figure 3-12. Existing Information on Health Effects of Stable Cobalt

162

	Qe de	in ko	je lite	S. The dide	system onic line	ic ic	And Section 1	Solutive Oe	a dormaria	doic	
Inhalation	•	•		•	•	•				•	
Oral	•		•		•			•			
Dermal				•	•						

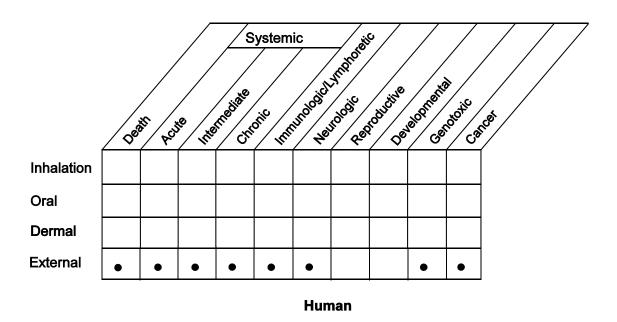
Human

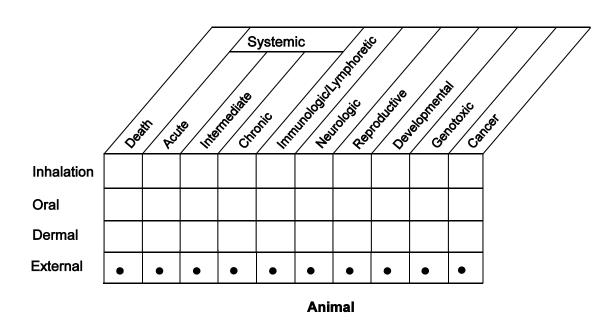
	Qe?	in bou		S Chi	bystem Oric cut	ic ic	red die	roducine De	a doment	doic	, coi
Inhalation	•	•	•	•	•	•	•	/ 🐧		•	
Oral	•	•	•		•	•	•	•			
Dermal	•	•			•						

**Animal** 

Existing Studies

Figure 3-13. Existing Information on Health Effects of Radioactive Cobalt





Existing Studies

Figures 3-12 and 3-13 represent studies conducted with all forms of cobalt. The effects of cobalt have been studied in humans following both inhalation and oral exposure. Human dermal studies designed to investigate nondermal systemic effects of cobalt have been reported. Similarly, the effects of cobalt in animals have been studied following inhalation and oral exposure. Few dermal studies are available.

#### 3.12.2 Identification of Data Needs

# **Acute-Duration Exposure.**

Stable Cobalt. Effects in humans following acute inhalation and oral exposure to cobalt have been reported; no studies of acute dermal exposure were reported. In humans, the primary targets following acute exposure to cobalt include the respiratory system following inhalation exposure (Kusaka et al. 1986a), the thymus following oral exposure (Roche and Layrisse 1956), and the immunological system following dermal exposure (Alomar et al. 1985; Fischer and Rystedt 1983; Kanerva et al. 1988). Acute oral studies in animals have also identified the cardiovascular and hematopoietic systems as targets of cobalt toxicity (Domingo and Llobet 1984; Speijers et al. 1982). Although acute exposure levels associated with some of these effects in humans have been reported, the minimal acute exposure levels required to produce these effects are not known because few acute human studies exist. The results of animal studies of the acute toxicity of cobalt have been used to determine dose levels that produce death and respiratory effects following inhalation exposure, death and various systemic effects following oral exposure, and dermal and immunological effects following dermal exposure. There were insufficient data for derivation of inhalation or oral acute MRLs because reported effects were severe and occurred at levels above those reported in the few human studies. Animal studies that identify minimally effective inhalation and oral exposure levels for the various cobalt compounds would be useful in estimating acute MRLs for each cobalt compound. Acute dermal studies would enable determination of hazardous levels for this route of exposure. Because a small portion of the cobalt taken into the body is retained for a relatively long time, studies on the long-term consequences of acute exposure on the heart, respiratory tract, hematological system, and immune response could provide information about the potential for chronic effects of acute exposures in humans. Knowledge about the acute toxicity of cobalt is important because people living near hazardous waste sites might be exposed for brief periods.

Radioactive Cobalt. Data on health effects following acute exposure to radioactive cobalt by the inhalation, oral, or dermal routes are lacking. Because the most common cobalt radioisotopes are manmade (see Chapter 5), only low-level exposure to radiocobalt in the environment by these routes is likely to occur. A number of health effects have been seen following cases of accidental acute exposure to high levels of external cobalt radiation in humans, including death, gastrointestinal disorders, hematological alterations, and dermal lesions (Klener et al. 1986; Stavem et al. 1985). Acute-exposure animal studies have shown pronounced effects, including death, cardiovascular changes, gastrointestinal effects, kidney effects, and neurobehavioral changes (Brady and Hayton 1977b; Bruner 1977; Cockerham et al. 1986; Darwezah et al. 1988; Down et al. 1986; Gomez-d-Segura et al. 1998; Hanks et al. 1966; King 1988a; Mele et al. 1988; Page et al. 1968; Robbins 1989a, 1989b, 1989c, 1991a). The most pronounced effects in animals following acute exposure to cobalt radiation have been reproductive and developmental effects (see Sections 3.2.4.5 and 3.2.4.6). ATSDR (1999) has derived an acute MRL for external exposure to ionizing radiation, which is applicable to external exposures to cobalt radiation, so additional data for the derivation of an MRL are not needed.

#### Intermediate-Duration Exposure.

Stable Cobalt. Information on oral exposure of humans to cobalt, in the form of cobalt chloride added to beer as a foam stabilizer, provides the only human data available for exposure of intermediate duration (Alexander 1969, 1972; Morin et al. 1971). Inhalation and dermal data in humans were not located for this duration of exposure. The cardiac and hematopoietic systems are the primary targets in humans following oral exposure to cobalt. Some exposure levels associated with cardiomyopathy have been reported following oral exposure, but the minimal exposure level required to produce this effect in humans is not known (Alexander 1969, 1972; Morin et al. 1971). Oral studies in animals reported dose levels associated with death, various systemic and neurological effects, and effects on reproduction and development (Domingo et al. 1984, 1985b; Krasovskii and Fridlyand 1971; Mohiuddin 1970; Mollenhauer et al. 1985; Pedigo et al. 1988). Intermediate-duration inhalation studies in animals reported that the respiratory tract is the target of the toxicity of inhaled cobalt (Bucher et al. 1990; Johansson et al. 1987; Kerfoot 1975; NTP 1991; Palmes et al. 1959). Animal studies were insufficient for derivation of an intermediate-duration MRL for oral exposure, since the reported effects were severe and the effects occurred at levels above those reported in the few human studies. Dermal data in animals were not located. Animal studies that investigate the possible toxic interaction between cobalt and alcohol may be helpful in understanding the role of cobalt in the cardiomyopathy reported in the heavy beer drinkers

(Alexander 1969, 1972; Morin et al. 1971). One such study in guinea pigs already exists (Mohiuddin et al. 1970), but this study used a single, high dose of cobalt. Studies using a series of lower doses, both with and without alcohol preexposure, would be helpful in determining the threshold for the cardiac effects. Intermediate-duration dermal studies would enable determination of hazardous levels for this route of exposure. Intermediate-duration toxicity information is important because people living near hazardous waste sites might be exposed for corresponding time periods.

Radioactive Cobalt. Data on health effects following acute exposure to radioactive cobalt by the inhalation, oral, or dermal routes are lacking. Because cobalt radioisotopes are man-made (see Chapter 5), only low-level exposure to radiocobalt in the environment by these routes is likely to occur. Substantial human data exist concerning intermediate-duration exposure to external radiation, as radiotherapy treatment regimens fall into this duration category. Animal data from intermediate-duration external exposure also exist, but are less numerous.

# **Chronic-Duration Exposure and Cancer.**

Stable Cobalt. Chronic inhalation exposure levels in humans associated with respiratory effects have been reported (Gennert and Lauwerys 1990; Nemery et al. 1992; Shirakawa et al. 1988; Sprince et al. 1988). In humans, the respiratory system is the primary target following chronic inhalation exposure. A chronic-duration inhalation MRL was derived from a NOAEL for decreased ventilatory function in exposed workers (Nemery et al. 1992). Wehner et al. (1977) reported no adverse effects in hamsters exposed chronically to cobalt oxide. NTP (1998; Bucher et al. 1999) exposed rats and mice to cobalt sulfate for 2 years, reporting pronounced effects on the respiratory tract, including hyperplasia, inflammation, fibrosis, and metaplasia; an increased incidence of cancer was also reported. Chronic oral or dermal studies have not been reported in either humans or animals. Animal studies that identify minimally effective chronic oral exposure levels would be useful for estimating a chronic MRL. Chronic dermal studies would enable determination of hazardous levels for this route of exposure. Chronic toxicity information is important because people living near hazardous waste sites might be exposed to cobalt for many years.

Several studies of hard metal exposure in humans have reported increases in lung cancer mortality from occupational inhalation exposure to hard metal (Lasfargues et al. 1994; Moulin et al. 1998; Wild et al. 2000). In humans, cancer has not been reported following exposure to cobalt by the oral or dermal routes.

An increased incidence of alveolar/bronchiolar neoplasms was noted following lifetime exposure of male rats to 1.14 mg cobalt/m³ and in female rats to 0.38 mg cobalt/m³ as cobalt sulfate, with tumors occurring in both sexes with significantly positive trends (Bucher et al. 1999; NTP 1998). Similarly, mice of both sexes exposed to 1.14 mg cobalt/m³ showed an increase in alveolar/bronchiolar neoplasms, again with lung tumors occurring with significantly positive trends. Parenteral exposure to cobalt has been found to induce tumors (Gilman 1962; Gilman and Ruckerbauer 1962; Heath 1956, 1969; Heath and Daniel 1962; Shabaan et al. 1977). Further chronic exposure studies by the oral and dermal routes may determine the actual carcinogenic potential of cobalt. Also, studies examining the effect of cobalt speciation (i.e., cobalt metal vs. cobalt sulfate) would add to our understanding of the carcinogenic potential of cobalt.

Radioactive Cobalt. Data on health effects following chronic exposure to radioactive cobalt by the inhalation, oral, or dermal routes are lacking. Because cobalt radioisotopes are man-made (see Chapter 5), only low-level exposure to radiocobalt in the environment by these routes is likely to occur. Limited data exist on chronic exposure to cobalt radiation in humans, with genotoxicity, immunologic effects, and cancer being the primary end points examined. Animal data are similarly limited. Additional human or animal data following chronic exposure to external cobalt radiation would be useful in further identifying possible long-term health effects or susceptible populations. ATSDR (1999) has derived a chronic-duration MRL for external radiation exposure, which is applicable to external exposures to cobalt radiation, so additional data for the derivation of an MRL are not needed.

# Genotoxicity.

Stable Cobalt. There are no data available regarding the genotoxicity of cobalt in humans *in vivo*. Data regarding the mutagenic action of cobalt in bacterial cell lines and mammalian cell lines have been reported in the literature (Hamilton-Koch et al. 1986; Kharab and Singh 1985; Ogawa et al. 1986). *In vivo* mutagenicity studies in animals following inhalation, oral, or dermal exposure to cobalt would be helpful in ascertaining its true mutagenic potential. Further studies examining the differences in genotoxicity between different valence states of cobalt would also be useful.

**Radioactive Cobalt.** Data on genotoxic effects following exposure to radioactive cobalt by the inhalation, oral, or dermal routes are lacking. Because cobalt radioisotopes are man-made (see Chapter 5), only low-level exposure to radiocobalt in the environment by these routes is likely to occur.

Several studies have demonstrated genotoxic effects in humans exposed to external cobalt radiation (Chang et al.1999c; House et al. 1992; Rauscher and Bauchinger 1983). Numerous data from animal studies exist demonstrating the genotoxic effects of ionizing radiation, including cobalt radiation.

# Reproductive Toxicity.

Stable Cobalt. No studies were located regarding the reproductive effects of cobalt in humans following exposure by any route. Inhalation and oral studies in male animals have demonstrated adverse effects on reproductive organs (Anderson et al. 1992, 1993; Bucher et al. 1990; Corrier et al. 1985; Domingo et al. 1985b; Mollenhauer et al. 1985; NTP 1991; Pedigo et al. 1988). One study also reported effects on the estrous cycle in mice following inhalation exposure (Bucher et al. 1990; NTP 1991). Multigenerational studies would be helpful in assessing the significance of these effects on reproductive performance.

Radioactive Cobalt. Data on reproductive effects following exposure to radioactive cobalt by the inhalation, oral, or dermal routes are lacking. Because cobalt radioisotopes are man-made (see Chapter 5), only low-level exposure to radiocobalt in the environment by these routes is likely to occur. Human data on reproductive effects following external exposure to cobalt radiation are lacking, but are sufficiently understood for gamma radiation. Available animal studies are limited, but have demonstrated radiation-induced deficits on reproductive ability in both genders (Cunningham and Huckins 1978; Laporte et al. 1985; Searl et al. 1976, 1980). Additional data in humans and animals would be helpful in refining minimal effective doses for radiation effects on reproduction.

# **Developmental Toxicity.**

Stable Cobalt. No developmental effects were observed in the children of 78 women given cobalt chloride orally during pregnancy for treatment of anemia (Holly 1955); however, only a limited examination of offspring was reported, and details of examined end points were not reported. No studies of developmental effects by other routes of exposure in humans were located. Developmental effects in animals following oral exposure during gestation, however, have been observed (Domingo et al. 1985b). Further developmental studies in animals by all relevant routes of exposure (inhalation, oral, dermal) may clarify the potential developmental effects of cobalt in humans.

Radioactive Cobalt. Data on developmental effects following exposure to radioactive cobalt by the inhalation, oral, or dermal routes are lacking. Because cobalt radioisotopes are man-made (see Chapter 5), only low-level exposure to radiocobalt in the environment by these routes is likely to occur. No human studies describing developmental effects of exposure to external cobalt radiation were located. Extensive data from animal studies have shown that even acute exposures to small amounts of cobalt radiation may elicit profound effects on the developing organism (see Section 3.2.4.6). The effects of ionizing radiation on the developing organism are also described in the ATSDR Toxicological Profile for Ionizing Radiation (1999).

# Immunotoxicity.

Stable Cobalt. Humans have been shown to develop sensitivity to cobalt following occupational exposure (Bencko et al. 1983; Shirakawa et al. 1988, 1989). No immunological effects were observed following oral exposure of humans to cobalt. Similar evidence of sensitization has been reported in animals (Lammintausta et al. 1985). Studies examining the mechanism of sensitization might be helpful in fully understanding and treating this effect in humans. A battery of immune function tests would further assess the immunotoxicity of cobalt in humans and animals.

Radioactive Cobalt. Data on immunotoxic effects following exposure to radioactive cobalt by the inhalation, oral, or dermal routes are lacking. Because cobalt radioisotopes are man-made (see Chapter 5), only low-level exposure to radiocobalt in the environment by these routes is likely to occur. Following external exposure to cobalt radiation, above levels normally encountered except for medical procedures, decreases in white blood cell counts have been seen in both humans and animals. Further studies on the immunotoxic effects of external cobalt radiation would be useful in refining the minimum effective dose.

# Neurotoxicity.

Stable Cobalt. No studies were located regarding neurotoxic effects of cobalt in humans following oral or dermal exposure. Two occupational inhalation exposure studies have reported memory deficits, optic atrophy, or nerve deafness in humans exposed to cobalt (Jordan et al. 1990; Meecham and Humphrey 1991). In animals, alterations in several neurologic parameters were found following oral exposure (Bourg et al. 1985; Krasovskii and Fridlyand 1971; Mutafova-Yambolieva et al. 1994; Nation et al. 1983;

Singh and Junnarkar 1991; Vassilev et al. 1993; Wellman et al. 1984). Additional studies in animals would assist in determining whether these neurological effects have any relevance to potential effects in humans.

Radioactive Cobalt. Data on neurotoxic effects following exposure to radioactive cobalt by the inhalation, oral, or dermal routes are lacking. Because cobalt radioisotopes are man-made (see Chapter 5), only low-level exposure to radiocobalt in the environment by these routes is likely to occur. Human data following cobalt radiotherapy have demonstrated effects believed to result from neurological damage, but data are limited, doses were extreme, and effects have not been well-characterized. Several animal studies have shown neurobehavioral or neurophysiological changes following exposure to cobalt radiation (Bassant and Court 1978; Maier and Landauer 1989; Mele et al. 1988).

# **Epidemiological and Human Dosimetry Studies.**

Stable Cobalt. Epidemiological studies relating to cobalt exposure are available in the literature. Studies of persons exposed to cobalt occupationally are available (Kusaka et al. 1986a, 1986b; Shirakawa et al. 1988, 1989; Sprince et al. 1988), dietetically (beer drinkers) (Alexander 1969, 1972; Morin et al. 1971), and medically (cobalt given to alleviate anemia) (Davis and Fields 1958; Holly 1955; Taylor et al. 1977). Further studies assessing the cause/effect relationship between cobalt exposure and human health effects would be helpful in monitoring individuals living near a hazardous waste site to verify that documented exposure levels are not associated with adverse health effects.

Radioactive Cobalt. Epidemiological data on exposure to radioactive cobalt by the inhalation, oral, or dermal routes are lacking. Because cobalt radioisotopes are man-made (see Chapter 5), only low-level exposure to radiocobalt in the environment by these routes is likely to occur. Human external exposures to cobalt radiation have been documented in the literature. Radiotherapy exposures, though to extremely high radiation doses, are generally well-controlled and documented, whereas environmental and accidental workplace exposures are less frequent and less well-documented.

# Biomarkers of Exposure and Effect.

#### Exposure.

Stable Cobalt. Information is available on the monitoring of cobalt exposure by the quantification of cobalt in urine and blood (Alexandersson 1988; Ichikawa et al. 1985; Scansetti et al. 1985). A portion of inhaled cobalt is rapidly excreted in the feces, and the amount retained in the body tends to be steadily excreted over time. Levels in body fluids, therefore, can be monitored up to several days after exposure. Many different methods for the detection of cobalt in body fluids have been reported (Section 7.1).

Radioactive Cobalt. No information is available regarding biomarkers specific for exposure to cobalt radionuclides by the inhalation, oral, dermal, or external exposure routes. Biomarkers for exposure to ionizing radiation are discussed in ATSDR (1999). Personal dosimeters (film or luminescent) are an artificial surrogate to measure the amount of exposure to external beta or gamma radiation, though these are not specific for radiation from cobalt radionuclides.

#### Effect.

Stable Cobalt. Alterations in serum proteins and changes in serum antibodies have been found that are specific for cobalt exposure (Stokinger and Wagner 1958). These changes may be the earliest indication of cobalt exposure. Further studies may reveal other cobalt-specific biomarkers that, in combination with these changes, may alert health professionals to cobalt exposure before serious toxicological effects occur.

**Radioactive Cobalt.** No information is available regarding biomarkers specific for effects of cobalt radionuclides following exposure by the inhalation, oral, dermal, or external exposure routes. Biomarkers for effects of ionizing radiation are discussed in ATSDR (1999), and include changes in levels of formed elements of the blood as some of the most sensitive indicators.

**Absorption, Distribution, Metabolism, and Excretion.** Pharmacokinetic data in humans indicate that cobalt is absorbed through the lungs (Foster et al. 1989) and the gastrointestinal tract (Harp and Scoular 1952; Sorbie et al. 1971; Valberg et al. 1969), that cobalt is well distributed in the body with the highest concentration being found in the lungs following inhalation (Gerhardsson et al. 1984; Hewitt

1988; Hillerdal and Hartung 1983; Teraoka 1981), and that some of the inhaled or ingested cobalt is rapidly excreted in the feces with the amount retained in the body being excreted slowly, primarily in the urine (Foster et al. 1989; Paley et al. 1958; Smith et al. 1972). Pharmacokinetic studies in animals following inhalation and oral exposure have demonstrated similar responses (Andre et al. 1989; Bailey et al. 1989; Collier et al. 1989; Foster et al. 1989; Patrick et al. 1989; Talbot and Morgan 1989). Few data exist regarding the pharmacokinetics of cobalt following dermal exposure, though what data are available demonstrate that cobalt can be absorbed in small quantities through human (Scansetti et al. 1994) and animal (Inaba and Suzuki-Yasumoto 1979; Lacy et al. 1996) skin, with more absorbtion occurring through damaged than intact skin.

**Comparative Toxicokinetics.** Several inhalation and oral studies have compared the toxicokinetics of cobalt in several different species of animals, including humans (Andre et al. 1989; Bailey et al. 1989; Collier et al. 1989; Foster et al. 1989; Patrick et al. 1989; Talbot and Morgan 1989). No comparative pharmacokinetic studies following dermal exposure were located. These studies would be useful because humans are exposed via the skin in the workplace and may potentially be exposed via this route at waste sites.

#### **Methods for Reducing Toxic Effects.**

Stable and Radioactive Cobalt. Chelation therapy is expected to apply equally well to stable and radioactive cobalt isotopes. EDTA or British anti-lewisite (BAL) has been shown to effectively mitigate the toxicity of cobalt in humans (Goldfrank et al. 1990; Haddad and Winchester 1990; Stutz and Janusz 1988). In animal studies examining the effectiveness of various chelators, NAC was shown to be the most effective (Llobet et al. 1988). It would be useful to determine the effective dose of NAC in humans. Studies examining the effectiveness of other chelating agents may be helpful in determining the most effective chelation therapy for humans.

# Children's Susceptibility.

*Stable Cobalt.* Data comparing the susceptibility of children to cobalt compounds are limited. Animal studies have suggested that absorption following inhalation or oral exposure may be greater in very young animals, resulting in increased systemic dose. Data are not available on the differences between children and adults following dermal exposure. Further studies on the susceptibility of young animals relative to

adult animals may be useful in determining whether children are at greater risk from exposure to cobalt in the environment than adults.

**Radioactive Cobalt.** No data are available on whether children are more susceptible to the effects of radiocobalt compounds than adults. Animal studies have shown that exposure *in utero* to even moderate amounts of cobalt radiation can cause dramatic effects in the developing organism. It would be expected that children would be more susceptible to the effects of external cobalt radiation, due to the greater percentage of rapidly-dividing cells during growth.

Child health data needs relating to exposure are discussed in 6.8.1 Identification of Data Needs: Exposures of Children.

# 3.12.3 Ongoing Studies

Relevant ongoing studies were not located for cobalt.

COBALT 175

# 4. CHEMICAL AND PHYSICAL INFORMATION

# 4.1 CHEMICAL IDENTITY

Cobalt is a naturally-occurring element that appears in the first transition series of Group 9 (VIII) of the periodic table along with iron and nickel. There is only one stable isotope of cobalt, <sup>59</sup>Co. There are about 26 known radioactive isotopes of cobalt, of which only two are of commercial importance, Cobalt-60 (<sup>60</sup>Co) and Cobalt-57 (<sup>57</sup>Co). <sup>60</sup>Co, a commonly-used source of gamma radiation, is the most important radionuclide. It is also a frequent low level contaminant of cooling water released by nuclear reactors. Table 4-1 summarizes information on the chemical identity of elemental cobalt and some common cobalt compounds.

# 4.2 PHYSICAL, CHEMICAL, AND RADIOLOGICAL PROPERTIES

Cobalt commonly occurs in the 0, +2, and +3 valence states. Compounds containing cobalt in the -1, +1, +4, and +5 oxidation state are few and uncommon (Cotton and Wilkinson 1980). Cobalt (II) is much more stable than Co(III), and Co<sup>3+</sup> is a sufficiently powerful oxidizing agent to oxidize water, liberating oxygen. Table 4-2 summarizes important physical and chemical properties of elemental cobalt and some common cobalt compounds. These properties are similar to those of its neighbors in Group 9 of the periodic table, iron and nickel. Metallic cobalt, Co(0), occurs as two allotropic forms, hexagonal and cubic; the hexagonal form is stable at room temperature. A biochemically important cobalt compound is vitamin B12, or cyanocobalamin, in which cobalt is complexed with four pyrrole nuclei joined in a ring called the corrinoid ligand system (similar to porphyrin).

The Chemical Abstract Service (CAS) registry numbers, decay modes, half-lives, and specific activity of the three principal radioactive cobalt isotopes, <sup>57</sup>Co, <sup>58</sup>Co, and <sup>60</sup>Co, are presented in Table 4-3. <sup>60</sup>Co (half-life of 5.27 years) decays by beta decay to nickel-60, a stable isotope (ICRP 1983; Lide 1998).

$$^{60}_{27}$$
Co 6  $^{60}_{28}$ Ni +  $^{0}_{-1}e$  +  $\gamma$ 

The decay is accompanied by the emission of 1.173 and 1.332 Mev gamma rays.  $^{57}$ Co (half-life of 271.8 days) and  $^{58}$ Co (half-life of 70.9 days) decay by electron capture and electron capture/position ( $\beta^+$ ) emission to  $^{57}$ Fe and  $^{58}$ Fe, respectively.

**Table 4-1. Chemical Identity of Cobalt and Selected Compounds** 

Characteristic	Cobalt	Cobalt(II) acetate	Cobalt(III) acetate	Cobalt(II) carbonate
Synonym(s)	Cobalt-59, cobalt metal	Cobaltous acetate, cobalt diacetate	Cobaltic acetate, cobalt triacetate	Cobaltous carbonate; carbonic acid; cobalt (+2) salt
Registered trade name(s)	No data	No data	No data	No data
Chemical formula	Со	$Co(C_2H_4O_2)_2$	$Co(C_2H_4O_2)_3$	CoCO <sub>3</sub>
Chemical structure	Со	_c_o_co_o_c—		oco
Identification numbers:				
CAS registry	7440-48-4	71-48-7	917-69-1	513-79-10
NIOSH RTECS	GF8750000	AG3150000	No data	FF9450050
EPA hazardous waste	No data	No data	No data	No data
OHM/TADS	No data	No data	No data	No data
DOT/UN/NA/IMCO shipping <sup>a</sup>	UN1318	No data	No data	No data
HSDB	519	997	No data	No data
NCI	C60311	No data	No data	No data

Table 4-1. Chemical Identity of Cobalt and Selected Compounds (continued)

Characteristic	Cobalt carbonyl	Cobalt(II) chloride	Cobalt(II) hydroxide	Cobalt(II) mesoporphyrin
Synonym(s)	Dicobalt octacarbonyl; cobalt tetracarbonyl	Cobalt dichloride; cobaltous chloride	Cobaltous hydroxide; cobalt dihydroxide	Cobalt mesoporphyrin IX Cobalti protoporphyrin
Registered trade name(s)	No data	No data	No data	No data
Chemical formula	Co <sub>2</sub> (CO) <sub>8</sub>	CoCl <sub>2</sub>	Co(OH) <sub>2</sub>	$C_{34}H_{34}CoN_4O_4$
Chemical structure		COCI	но-со-он	No data
Identification numbers:				
CAS registry	10210-68-1	7646-79-9	21041-93-0	21158-51-0
NIOSH RTECS	GG0300000	GF9800000	No data	No data
EPA hazardous waste	No data	No data	No data	No data
OHM/TADS	No data	7217328	No data	No data
DOT/UN/NA/IMCO shipping	No data	No data	No data	No data
HSDB	6345	1000	No data	No data
NCI	No data	No data	No data	No data

Table 4-1. Chemical Identity of Cobalt and Selected Compounds (continued)

Characteristic	Cobalt(II) naphthenate	Cobalt(II) nitrate	Cobalt(II) oxide	Cobalt(III) oxide
Synonym(s)	Naftolite; naphthenic acid, cobalt salt	Cobaltous nitrate	Black 13; C.I. 77322; cobalt monoxide; cobaltous oxide	Cobalt black; cobaltic oxide; cobalt sesquioxide; cobalt trioxide; C.I. 77323
Registered trade name(s)	No data	No data	C.I. Pigment Black 13; Zaffre	No data
Chemical formula	$Co(C_{11}H_{10}O_2)_2$	Co(No <sub>3</sub> ) <sub>2</sub> •6H <sub>2</sub> 0	CoO	Co <sub>2</sub> O <sub>3</sub>
Chemical structure	, o – co	$Co = \begin{bmatrix} O - N \\ O \end{bmatrix}_{2}$	Co=O	O==Co-OCo=C
Identification numbers:				
CAS registry	61789-51-3	10026-22-9	1307-96-6	1308-04-9
NIOSH RTECS	QK8925000	QU7355500	GG2800000	GG2900000
EPA hazardous waste	No data	No data	No data	No data
OHM/TADS	No data	No data	No data	No data
DOT/UN/NA/IMCO shipping	UN2001 (powder)	No data	No data	No data
HSDB	No data	No data	239	No data
NCI	No data	No data	No data	No data

Table 4-1. Chemical Identity of Cobalt and Selected Compounds (continued)

Characteristic	Cobalt(II, III) oxide	Cobalt(II) sulfate
Synonym(s)	Cobaltic-cobaltous oxide; cobalt tetraoxide, tricobalt tetraoxide, cobaltosic oxide; cobalt black; C.I. Pigment Black 13	Cobalt sulfate; cobaltous sulfate
Registered trade name(s)	No data	No data
Chemical formula	Co <sub>3</sub> O <sub>4</sub>	CoSO <sub>4</sub>
Chemical structure	co=0 0=co-0-co=0	co
Identification numbers:		
CAS registry	1308-06-1	10124-43-3
NIOSH RTECS	No data	GG3100000
EPA hazardous waste	No data	No data
OHM/TADS	No data	7217330
DOT/UN/NA/IMCO shipping	No data	No data
HSDB	No data	240
NCI	No data	No data

Source: Budavari 1996; HSDB 2001; RTECS 1987

CAS = Chemical Abstract Service; DOT/UN/NA/IMCO = Department of Transportation/United Nations/North America/International Maritime Dangerous Goods Code; EPA = Environmental Protection Agency; HSDB = Hazardous Substances Data Bank; NCI = National Cancer Institute; NIOSH = National Institute for Occupational Safety and Health; OHM/TADS = Oil and Hazardous Materials/Technical Assistance Data System; RTECS = Registry of Toxic Effects of Chemical Substances

<sup>&</sup>lt;sup>a</sup> The identification number for radioactive materials is UN2910

Table 4-2. Physical and Chemical Properties of Cobalt and Selected Compounds

Property	Cobalt	Cobalt(II)acetate	Cobalt(III)acetate	Cobalt(II) carbonate
Molecular weight	58.93	177.03	236.07	118.94
Color	Silvery gray	light pink	dark green	Red
Physical state	Solid	solid	solid	Solid
Melting point, EC	1,495	No data	decomposes at 100 EC	Decomposes
Boiling point, EC	2,870	No data	Not relevant	Not relevant
Density, g/cm <sup>3</sup>	8.9 (20 EC)	No data	No data	4.13
Odor	No data	No data	No data	No data
Odor threshold: Water Air	No data No data	No data No data	No data No data	No data No data
Solubility: Water Organic solvent(s)	Insoluble Insoluble	soluble No data	soluble soluble in alcohol, acetic acid	Insoluble No data
Partition coefficients: Log K <sub>ow</sub> Log K <sub>oc</sub>	No data No data	No data No data	No data No data	No data No data
Vapor pressure	1 mmHg at 1,910 EC	No data	No data	No data
Henry's law constant	No data	No data	No data	No data
Autoignition temperature	760 EC for dust cloud	No data	No data	No data
Flashpoint	No data	No data	No data	No data
Flammability limits	No data	No data	No data	No data
Conversion factors	Not relevant <sup>a</sup>	Not relevant <sup>a</sup>	Not relevant <sup>a</sup>	Not relevant <sup>a</sup>
Explosive limits	No data	No data	No data	No data

Table 4-2. Physical and Chemical Properties of Cobalt and Selected Compounds (continued)

Property	Cobalt carbonyl	Cobalt(II) chloride	Cobalt(II) hydroxide	Cobalt(II) mesoporphyrin
Molecular weight	341.9	129.84	92.95	621.2 <sup>b</sup>
Color	orange (white when pure)	Blue	rose red or blue green	No data
Physical state	solid	Solid	solid	No data
Melting point, EC	51	724	No data	No data
Boiling point, EC	decomposes	1,049	No data	No data
Density, g/cm³	1.73 at 18 EC	3.356 (36 EC)	3.597 at 15 EC	No data
Odor	No data	No data	No data	No data
Odor threshold: Water Air	No data No data	No data No data	No data No data	No data No data
Solubility: Water Organic solvent(s)	insoluble soluble in ether insoluble in naphtha	450 g/L at 7 EC 544 g/L in ethanol 86 g/L in acetone	0.0032 g/L No data	No data No data
Partition coefficients: Log K <sub>ow</sub> Log K <sub>oc</sub>	No data No data	No data No data	No data No data	No data No data
Vapor pressure	199.5 at 25 EC	No data	No data	No data
Henry's law constant	No data	No data	No data	No data
Autoignition temperature, EC	No data	No data	No data	No data
Flashpoint, EC	No data	No data	No data	No data
Flammability limits	No data	No data	No data	No data
Conversion factors	Not relevant <sup>a</sup>	Not relevant <sup>a</sup>	Not relevant <sup>a</sup>	Not relevant <sup>a</sup>
Explosive limits	No data	No data	No data	No data

Table 4-2. Physical and Chemical Properties of Cobalt and Selected Compounds (continued)

Property	Cobalt(II) naphthenate	Cobalt(II) nitrate	Cobalt(II) oxide	Cobalt(III) oxide
Molecular weight	407	182.94	74.93	165.86
Color	No data	Red	Pink	Black-gray
Physical state	solid	Solid	Solid	Solid
Melting point, EC	140	Decomposes at 100-105 <sup>b</sup>	1,795	895 (decomposes)
Boiling point, EC	No data	Not relevant	No data	Not relevant
Density g/cm <sup>3</sup>	0.9	2.49 <sup>b</sup>	6.45	5.18
Odor	No data	No data	No data	No data
Odor threshold: Water Air	No data No data	No data No data	No data No data	No data No data
Solubility: Water Organic solvent(s)	Insoluble	133.8 at 0 EC° Soluble in ethanol, acetone	Insoluble Insoluble in alcohol	Insoluble Insoluble in ethanol
Partition coefficients: Log K <sub>ow</sub> Log K <sub>ow</sub>	No data No data	No data No data	No data No data	No data No data
Vapor pressure	No data	No data	No data	No data
Henry's law constant	No data	No data	No data	No data
Autoignition temperature	No data	No data	No data	No data
Flashpoint	No data	No data	No data	No data
Flammability limits	No data	No data	No data	No data
Conversion factors	Not relevant <sup>a</sup>	Not relevant <sup>a</sup>	Not relevant <sup>a</sup>	Not relevant <sup>a</sup>
Explosive limits	No data	No data	No data	No data

Table 4-2. Physical and Chemical Properties of Cobalt and Selected Compounds (continued)

Property	Cobalt(II, III) oxide	Cobalt(II) sulfate
Molecular weight	250.80	154.99
Color	Black	Dark blue
Physical state	Solid	Solid
Melting point, EC	-O <sub>2</sub> at 900–950	Decomposes at 735 EC
Boiling point, EC	Not relevant	Not relevant
Density g/cm <sup>3</sup>	6.07	3.71
Odor	No data	No data
Odor threshold: Water Air	No data No data	No data No data
Solubility: Water Organic solvent(s)	Insoluble No data	soluble Slightly soluble in methanol
Partition coefficients: Log K <sub>ow</sub> Log K <sub>ow</sub>	No data No data	No data No data
Vapor pressure	No data	No data
Henry's law constant	No data	No data
Autoignition temperature	No data	No data
Flashpoint	No data	No data
Flammability limits	No data	No data
Conversion factors	Not relevant <sup>a</sup>	Not relevant <sup>a</sup>
Explosive limits	No data	No data

Source: Budavari 1996; HSDB 2001; Lide 1994; Stockinger 1981; Weast 1985

<sup>&</sup>lt;sup>a</sup>Substances exist in the atmosphere in the particulate state, and the concentration is expressed in weight per cubic meter.

<sup>&</sup>lt;sup>b</sup>CAS Online

<sup>&</sup>lt;sup>c</sup>Hexahydrate

**Table 4-3. Principal Radioactive Cobalt Isotopes** 

		_	Decay mode	Beta	radiation	Gamma	radiation	_
Isotope	CAS registry no.	Decay mode (product)	energy (MeV)	Energy (MeV)	Intensity (percent)	Energy (MeV)	Intensity (percent)	Half-life
<sup>55</sup> Co	13982-25-7	E.C. β <sup>+</sup> ( <sup>55</sup> Fe)	3.452	1.498	46	0.9312	75	17.53 hours
				1.021	25.6	0.4772	20	
				2.043	10.7	1.408	16.88	
<sup>57</sup> Co	13981-50-5	E.C. ( <sup>57</sup> Fe)	0.836	0.700	99.8	0.1221	85.6	271.8 days
						0.1365	10.7	
						0.014	9.2	
<sup>58</sup> Co	13981-38-9	E.C. β <sup>+</sup> ( <sup>58</sup> Fe)	2.30	1.4966	83.9	0.811	99	70.86 days
				0.4746	14.9			
<sup>60</sup> Co	10198-40-0	$\beta^{-}(^{60}Ni)$	2.824	0.3181	99.9	1.173	100	5.271 years
						1.332	100	

 $\beta^-$  = negative beta emission;  $\beta^+$  = positron emission; E.C. = orbital electron capture

Source: ICRP 1983; LBNL 2000; Lide 1998;

COBALT 185

# 5. PRODUCTION, IMPORT/EXPORT, USE, AND DISPOSAL

#### 5.1 PRODUCTION

Cobalt is the 33<sup>rd</sup> most abundant element, comprising approximately 0.0025% of the earth's crust. It is often found in association with nickel, silver, lead, copper, and iron ores and occurs in mineral form as arsenides, sulfides, and oxides. The most import cobalt minerals are: linnaeite, Co<sub>3</sub>S<sub>4</sub>; carrolite, CuCo<sub>2</sub>S<sub>4</sub>; safflorite, CoAs<sub>2</sub>; skutterudite, CoAs<sub>3</sub>; erythrite, Co<sub>3</sub>(AsO<sub>4</sub>)<sub>2</sub>•8H<sub>2</sub>O; and glaucodot, CoAsS (Hodge 1993; IARC 1991; Merian 1985; Smith and Carson 1981). The largest cobalt reserves are in Zaire, Zambia, Morocco, Canada, and Australia; Zaire and Zambia have the richest deposits. Most of the U.S. cobalt deposits are in Minnesota, but other important deposits are in Alaska, California, Idaho, Missouri, Montana, and Oregon. Most U.S. cobalt resources are in subeconomic concentrations that are not expected to be feasible to exploit in the foreseeable future. Cobalt is also found in meteorites and deep sea nodules.

The production of pure metal from these ores depends on the nature of the ore. Sulfide ores are first finely ground and the sulfides are separated by a floatation process with the aid of frothers. The concentrated product is subjected to sulfatizing roasting, and the resulting matte is leached with water. The cobalt sulfate leachate is precipitated as its hydroxide by the addition of lime. The hydroxide is dissolved in sulfuric acid, and the resulting cobalt sulfate is electrolyzed to yield metallic cobalt. For the cobalt-rich mineral cobaltite, a leaching process with either ammonia or acid under pressure and elevated temperatures has been used to extract cobalt. The solution is purified to remove iron and is subsequently reduced by hydrogen in the presence of a catalyst under elevated temperature and pressure to obtain fine cobalt powder (Planinsek and Newkirk 1979).

Except for a negligible amount of byproduct cobalt produced from some mining operations, no cobalt is mined or refined in the United States. In addition to byproduct production, U.S. production is derived from scrap (secondary production). In 1998, 3,080 metric tons of cobalt was recycled from scrap. Since 1993, production has been supplemented by sales of excess cobalt from the National Defense Stockpile (NDS), which the government maintains for military, industrial, and essential civilian use during national emergencies. In fiscal year 1999, 1,960 metric tons of cobalt were released from the NDS. In 1995–1998, between 1,550 and 2,310 metric tons of cobalt were removed from the NDS annually. Cobalt is present in the ores mined for platinum group metals at the Stillwater Complex in southern Montana

where cobalt is produced as a byproduct. The 1998 U.S. consumption of cobalt metal, organic and inorganic cobalt compounds, and purchased scrap (in terms of cobalt content) was 3,780, 1,910, and 2,720 metric tons, respectively (USGS 1998).

Current U.S. manufacturers of selected cobalt compounds are given in Table 5-1. Table 5-2 lists facilities in each state that manufacture, process, or use cobalt or cobalt compounds, the intended use, and the range of maximum amounts of these substances that are stored on site. In 1999, there were 695 reporting facilities that produced, processed, or used cobalt or cobalt compounds in the United States. The data listed in Table 5-2 are derived from the Toxics Release Inventory (TRI99 2001). Only certain types of facilities were required to report. Therefore, this is not an exhaustive list.

<sup>60</sup>Co is produced by irradiating natural cobalt, <sup>59</sup>Co, with thermal neutrons in a nuclear reactor: <sup>59</sup>Co(n,γ)<sup>60</sup>Co. The neutron flux employed is 10<sup>12</sup>–10<sup>15</sup> *n*/cm<sup>2</sup>-sec and the conversion is 99%. The maximum activity obtained is 3.7x10<sup>13</sup> Bq/g (1,000 Ci/g). Commercial <sup>60</sup>Co sources are made into rods with double metal shielding. The individual sources have an activity of about 2x10<sup>14</sup>–6x10<sup>14</sup> Bq (6–15 kCi). The annual output of <sup>60</sup>Co was about 2x10<sup>18</sup>–3x10<sup>18</sup> Bq (50–80 MCi) in the early 1990s. In 1991, there were 170 gamma irradiation systems operating in 45 countries having a total activity of about 6x10<sup>18</sup> Bq (160 MCi) (Zyball 1993). Producers of <sup>60</sup>Co include MDS Nordion in Canada, AEA Technology (formerly Amersham QSA) in the United Kingdom, and Neutron Products in Dickerson, Maryland.

<sup>58</sup>Co is not produced commercially. It can be produced by irradiating <sup>58</sup>Ni, a stable isotope, with neutrons, followed by positron decay:  $^{58}$ Ni(n, $\gamma$ ) $^{58}$ Co. It can be produced in a nuclear reactor or a cyclotron. Both  $^{60}$ Co and  $^{58}$ Co may be produced unintentionally in reactors. These are the dominant source of residual radiation in the primary circuit outside the reactor core of nuclear plants and are formed by neutron absorption of  $^{59}$ Co and  $^{58}$ Ni, both stable isotopes commonly used in plant construction materials (Taylor 1996).  $^{60}$ Co is a frequent major contaminant of cooling water released by nuclear reactors.

The <sup>60</sup>Co activities for a typical pressurized-water reactor (PWR) and boiling water reactor (BWR) fuel assemblies are 150 and 37 Ci, respectively (DOE 1999). There are 78 PWR and 41 BWR reactors in the United States, several of which have ceased operation. The total projected inventory of <sup>60</sup>Co for all

Table 5-1. Current U.S. Manufacturers of Cobalt Metal and Selected Cobalt Compounds<sup>a</sup>

Company	Location
Cobalt metal <sup>b</sup> Kennametal, Inc. OM Group, Inc. Osram Sylvania Products, Inc. Stoody Company	Latrobe, Pennsylvania Cleveland, Ohio Towanda, Pennsylvania St. Louis, Missouri
Cobalt (II) acetate: The Hall Chemical Co.  McGean-Rohco, Inc. OM Group, Inc. The Shepherd Chemical Co.	Arab, Alabama Wickliffe, Ohio Cleveland, Ohio Franklin, Pennsylvania Cincinnati, Ohio
Cobalt (II) carbonate:     The Hall Chemical Co.     IMC/Americhem     McGean-Rohco, Inc.     OMG Apex     OM Group, Inc.     The Prince Manufacturing Company  The Shepherd Chemical Co.	Wickliffe, Ohio Shelby, North Carolina Cleveland, Ohio St. George, Utah Franklin, Pennsylvania Bowmanstown, Pennsylvania Quincy, Illinois Phillipsburg, New Jersey Cincinnati, Ohio
Cobalt (II) chloride:     The Hall Chemical Co.     IMC/Americhem     Johnson Matthey, Inc., Alfa Aesar     McGean-Rohco, Inc.     OM Group, Inc.     The Shepherd Chemical Co.     Union Miniere, Inc., Carolmet Cobalt     Products Division	Wickliffe, Ohio Shelby, North Carolina Ward Hill, Massachusetts Cleveland, Ohio Franklin, Pennsylvania Cincinnati, Ohio Laurinburg, North Carolina
Cobalt (II) hydoxide: The Hall Chemical Co. McGean-Rohco, Inc. OM Group, Inc. The Shepherd Chemical Co.	Wickliffe, Ohio Cleveland, Ohio Franklin, Pennsylvania Cincinnati, Ohio

Cobalt (II) nitrate:

The Hall Chemical Co.

IMC/Americhem

Johnson Matthey, Inc., Alfa Aesar

Arab, Alabama

Shelby, North Carolina

Ward Hill, Massachusetts

McGean-Rohco, Inc
OMG Apex
OM Group, Inc.
Cleveland, Ohio
St. George, Utah
Franklin, Pennsylvania

The Shepherd Chemical Co. Cincinnati, Ohio

Union Miniere, Inc., Carolmet Cobalt Laurinburg, North Carolina

Products. Division

# Table 5-1. Current U.S. Manufacturers of Cobalt Metal and Selected Cobalt Compounds<sup>a</sup> (continued)

Company	Location	
Cobalt (II) oxide: The Hall Chemical Co. IMC/Americhem OMG Apex The Shepherd Chemical Co.	Wickliffe, Ohio Shelby, North Carolina St. George, Utah Cincinnati, Ohio	
Cobalt (III) oxide The Hall Chemical Co. Johnson Matthey, Inc., Alfa Aesar Mallinckrodt Baker, Inc. McGean-Rohco, Inc. OM Group, Inc.	Wickliffe, Ohio Ward Hill, Massachusetts Phillipsburg, New Jersey Cleveland, Ohio Franklin, Pennsylvania	
Cobalt (II) sulfate The Hall Chemical Co.  IMC/Americhem McGean-Rohco, Inc. OMG Apex OM Group, Inc. The Prince Manufacturing Company  The Shepherd Chemical Co.	Arab, Alabama Wickliffe, Ohio Shelby, North Carolina Cleveland, Ohio St. George, Utah Franklin, Pennsylvania Bowmanstown, Pennsylvania Quincy, Illinois Cincinnati, Ohio	

<sup>&</sup>lt;sup>a</sup>Derived from SRI 1999, receipt where otherwise noted. SRI reports production of chemicals produced in commercial quantities (defined as exceeding 5,000 pounds or \$10,000 in value annually) by the companies listed. <sup>b</sup>U.S. members of The Cobalt Development Institute that are listed as producers of cobalt powder or hard metal

products.

COBALT 189 5. PRODUCTION, IMPORT/EXPORT, USE, AND DISPOSAL

Table 5-2. Facilities that Produce, Process, or Use Cobalt or Cobalt Compounds

State	Number of facilities	Minimum amount on site in pounds <sup>b</sup>	Maximum amount on site in pounds <sup>b</sup>	Activities and uses <sup>c</sup>
AK	2	10,000	999,999	1, 5, 8, 13
AL	17	1,000	999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 13
AR	6	1,000	99,999	1, 2, 3, 4, 5, 8, 9, 10
AZ	11	1,000	9,999,999	1, 2, 3, 4, 5, 6, 8, 9, 10, 11, 13
CA	18	1,000	999,999	2, 3, 4, 7, 8, 9, 10, 11, 13
CT	8	100	9,999,999	2, 3, 8, 9, 10, 12
DE	2	100	99,999	1, 5, 6, 11
FL	10	0	99,999	1, 3, 4, 5, 6, 7, 8, 9, 10, 13
GA	13	100	999,999	1, 3, 4, 5, 6, 7, 8, 9, 10, 12, 13
IA	7	1,000	999,999	3, 4, 8, 9, 13
ID	1	100,000	999,999	1, 5
IL	25	0	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13
IN	25	100	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13
KS	5	1,000	99,999	1, 3, 5, 7, 8, 9, 11, 13
KY	16	1,000	999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 12, 13
LA	10	1,000	999,999	1, 2, 3, 5, 6, 7, 9, 11, 13
MA	6	1,000	999,999	1, 5, 9, 10, 13
MD	4	1,000	99,999	1, 2, 3, 5, 6, 7, 8
ME	1	10,000	99,999	9
MI	15	0	999,999	1, 2, 3, 4, 5, 6, 8, 9, 10, 11, 12, 13
MN	2	1,000	99,999	1, 6, 7, 8, 10, 11
MO	6	1,000	999,999	1, 2, 3, 4, 5, 6, 7, 9, 10, 13
MS	7	100	99,999	1, 3, 4, 5, 6, 7, 8, 9, 10, 11
MT	1	10,000	99,999	1, 3, 5, 6, 13
NC	17	1,000	999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 13
ND	2	1,000	9,999	1, 5, 6, 13
NE	1	1,000	9,999	9, 12
NH	1	100	999	9
NJ	7	1,000	999,999	1, 3, 4, 7, 8, 9
NM	10	1,000	10,000,000,000	1, 2, 3, 4, 5, 6, 8, 9, 10, 12, 13
NV	12	1,000	9,999,999	1, 5, 6, 7, 8, 9, 10, 12, 13
NY	10	1,000	999,999	1, 2, 3, 4, 5, 8, 9, 10, 12

Table 5-2. Facilities that Produce, Process, or Use Cobalt or Cobalt Compounds *(continued)* 

State	Number of facilities	Minimum amount on site in pounds <sup>b</sup>	Maximum amount on site in pounds <sup>b</sup>	Activities and uses <sup>c</sup>
ОН	31	0	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 12, 13
OK	8	100	99,999	1, 2, 3, 4, 5, 7, 8, 9
OR	4	1,000	999,999	8, 10, 13
PA	31	0	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 12, 13
PR	2	1,000	99,999	9, 10
RI	1	100,000	999,999	9
SC	20	100	999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13
TN	13	100	999,999	1, 2, 3, 5, 6, 7, 8, 9, 11, 13
TX	22	0	999,999	1, 3, 4, 5, 6, 7, 8, 9, 10, 11, 13
UT	6	1,000	999,999	1, 2, 3, 4, 5, 6, 8, 9, 10, 13
VA	6	10,000	999,999	1, 2, 3, 5, 7, 8, 9
VI	1	10,000	99,999	11
WA	2	10,000	99,999	1, 3, 4, 5, 6, 10, 11, 12, 13
WI	11	0	999,999	1, 2, 3, 4, 5, 6, 8, 9, 10, 13
WV	7	100	999,999	1, 2, 3, 4, 5, 6, 8, 9, 10, 12, 13
WY	2	0	99,999	1, 3, 4, 5, 6, 10, 13

Source: TRI99 2001

- 1. Produce
- 2. Import
- 3. Onsite use/processing
- 4. Sale/Distribution
- 5. Byproduct

- 6. Impurity
- 7. Reactant
- 8. Formulation Component
- 9. Article Component
- 10. Repackaging
- 11. Chemical Processing Aid
- 12. Manufacturing Aid
- 13. Ancillary/Other Uses

<sup>&</sup>lt;sup>a</sup>Post office state abbreviations used

<sup>&</sup>lt;sup>b</sup>Amounts on site reported by facilities in each state

<sup>&</sup>lt;sup>c</sup>Activities/Uses:

reactors is  $1.7x10^7$  Ci. The postirradiation cobalt content of typical PWR and BWR reactor fuel assemblies are 38 g (0.01%) and 26 g (0.01%), respectively.

<sup>55</sup>Co may be produced by applying 12 MeV indirect deuteron energy to <sup>54</sup>Fe (<sup>54</sup>Fe(d,n)<sup>55</sup>Co), 40 MeV protons to natural iron (<sup>56</sup>Fe(p,2n)<sup>55</sup>Co), or 20 MeV protons to natural nickel foil (<sup>58</sup>Ni(p,α)<sup>55</sup>Co) followed by separation of the <sup>55</sup>Co on an ion exchange column (Wolf 1955). <sup>57</sup>Co is produced by AEA Technology (formerly Amersham QSA) in the United Kingdom (Web Research Co. 1999).

#### 5.2 IMPORT/EXPORT

In 1999, 8,150 metric tons of cobalt was imported into the United States compared with 6,440, 6,710, 8,430, and 7,670 metric tons in 1995, 1996, 1997, and 1998; respectively; 8,000 metric tons of cobalt is estimated to be imported in 2000 (USGS 2000, 2001). Between 1995 and 1998, Norway, Finland, Canada, and Zambia supplied 24, 18, 14, and 13% of cobalt. Imports for 1998 by form included (form, metric tons cobalt content): metal, 6,450; oxides and hydroxides, 868; acetates, 55; carbonates, 8; chlorides, 6; and sulfates 281. Cobalt exports for 1995, 1996, 1997, 1998, and 1999 were 1,300, 1,660, 1,570, 1,680, and 1,550 metric tons, respectively. Exports for 2000 are estimated to reach 2,300 metric tons.

<sup>60</sup>Co and <sup>57</sup>Co are produced in Canada and the United Kingdom and are imported from these countries. No import and export quantities for cobalt radioisotopes were available.

#### 5.3 USE

The United States is the world's largest consumer of cobalt. Cobalt is used in a number of essential military and industrial applications. The largest use of metallic cobalt is in superalloys that are used in gas turbines aircraft engines. Superalloys are alloys developed for applications where elevated temperatures and high mechanical stress are encountered. It is also used in magnetic alloys and alloys that are required for purposes requiring hardness, wear resistance, and corrosion resistance. Cobalt is used as a binder for tungsten carbide (cemented carbides) cutting tools to increase impact strength. Cobalt compounds are used as pigments in glass, ceramics, and paints; as catalysts in the petroleum industry; as paint driers; and as trace element additives in agriculture and medicine.

Over 40% of nonmetallic cobalt is used in catalysis and most cobalt catalysts are used in hydrotreating/ desulfurization in the oil and gas industry, the production of terephthalic acid and dimethylterephthalate, and the high pressure OXO process for the production of aldehydes. Cobalt chemicals primarily used as catalyst include cobalt(III) acetate, cobalt(II) bromide, carbonate, manganate, oxalate, and sulphide, cobalt carbonyl, and cobalt naphthenate. Cobalt carbonate and chromate are mainly used as pigments and cobalt(II) acetate, 2-ethylhexanoate, linoleate, naphthenate, nitrate, oleate, and stearate are mainly used as driers. Cobalt has been used for hundreds of years as a blue colorant in glass, ceramics and paints.

A growing use for cobalt is as an addition to the Ni/Cd, Ni-metal hydride battery or as the main component of the lithium ion cell (LiCoO<sub>2</sub>). In 1999, the reported U.S. cobalt consumption was 8,420 metric tons with a use pattern was (end use, metric tons cobalt content, percent): superalloys, 3,830, 45.5%; steel alloys, 154, 1.8%; other alloys, including magnetic alloys, 1,085, 12.9%; cemented carbides, 755, 9.0%; chemical and ceramic use, 2,530, 30.0%; and miscellany, 64, 0.76%. Cobalt is also used a target material in electrical x-ray generators (Cobalt Development Institute 2000; Donaldson 1986; Hodge 1993; IARC 1991; Richardson 1993; USGS 1998).

Gamma rays from <sup>60</sup>Co are used to sterilize medical and consumer products, to cross-link, graft and degrade plastics, and as an external source in radiography and radiotherapy. In addition, <sup>60</sup>Co, along with iridium-192, is the most commonly used isotope in radiography. In this application, <sup>60</sup>Co is used for nondestructive testing of high-stress alloy parts, such as pipeline weld joints, steel structures, boilers, and aircraft and ship parts. Radiography may be conducted at permanent, specially shielded facilities or temporary sites in the field (NRC 1999). <sup>60</sup>Co is used in chemical and metallurgical analysis and as a tracer in biological studies. In 1990, about 95% of installed <sup>60</sup>Co activity was used for the sterilization of medical devices; about 45% of medical devices were sterilized using radiation. <sup>60</sup>Co is also a source of gamma rays used for food irradiation; depending on the dose levels, irradiation may be used to sterilize food, destroy pathogens, extend the shelf-life of food, disinfest fruits and grain, delay ripening, and retard sprouting (e.g., potatoes and onions). Sludge, waste water, and wood may also be treated with gamma rays to kill harmful organisms.

<sup>57</sup>Co decays to an excited state of <sup>57</sup>Fe, the most widely used x-ray source in Mössbauer spectroscopy (Hodge 1993; Richardson 1993). It is also made into standards and sources for dose calibrators, gamma cameras, and gauges, and is used as markers and rulers to help estimate organ size/location. It is also used in *in vitro* diagnostic kits for the study of anemia related to vitamin B<sub>12</sub> deficiency/malabsorption

(MDS Nordion 2000). <sup>55</sup>Co- bleomycin has been used for scanning malignant tumors (e.g., lung and brain cancer) and is a practical isotope for positron emission tomography (PET) studies because it mainly (81%) decays by positron emission.

#### 5.4 DISPOSAL

There is a paucity of data on the methods of disposal of cobalt and its compounds. Due to the lack of natural sources of economically extractable ores in the United States, cobalt is entirely imported in the United States, and it is considered a strategic mineral. It is economical to recycle certain cobalt wastes rather than to dispose of them. Recycling of superalloy scrap is an important method for the recovery of cobalt. About 2,800 metric tons of cobalt were recycled from purchased scrap in 2000. This was about 33% of reported consumption for the year. According to the Toxic Chemicals Release Inventory (TRI98 2000), 4.42 and 9.01 million pounds of cobalt and cobalt compounds were recycled on-site and off-site, respectively, in 1998. Waste containing cobalt dust and, presumably, waste containing cobalt in the solid state may be placed in sealed containers and disposed of in a secured sanitary landfill (HSDB 1989). Waste water containing cobalt can be treated before disposal, for instance, by precipitation of carbonate or hydroxide of cobalt or by passage through an ion-exchange resin (Clifford et al. 1986). According to the Toxic Chemicals Release Inventory (TRI99 2001), 1,296,686 pounds of cobalt and cobalt compounds, was transferred off-site for disposal, including solidification/stabilization and waste water treatment, including publicly operated treatment plants (POTWs). The amount of cobalt so transferred by state is shown in Table 6-1.

In August 1998, EPA issued a final rule listing spent hydrotreated and hydrorefined catalysts as hazardous waste under the Resource Conservation and Recovery Act (USGS 1998). Listing under this act requires that releases of these substances will be subject to certain management and treatment standards and emergency notification requirements. Information regarding effluent guidelines and standards for cobalt may be found in Title 40 of the Code of Federal Regulations, Parts 421.230, 421.310, and 471.30.

<sup>60</sup>Co sources used for irradiation purposes are valuable and would not be discarded. However, some radioactive cobalt isotopes may occur in waste material from nuclear reactors. Radioactive waste is categorized according to origin, type of waste present, and level of activity. Radioactive cobalt isotopes may be commingled with other radioactive isotopes. The first distinction in radioactive waste is between defense waste and commercial waste, the former being generated during and after World War II

principally at the Department of Energy (DOE) facilities at Hanford, Washington; Savannah River, South Carolina; and Idaho Falls, Idaho, where plutonium and other isotopes were separated from production reactor spent fuel or nuclear-powered naval vessels. Commercial wastes are produced predominantly by nuclear power plants as well as the long defunct commercial reprocessing facility at West Valley, New York and manufacturers of radioisotopes used in nuclear medicine for the treatment and diagnosis of disease. Nuclear waste is also classified as high-level waste (HLW), transuranic waste (TRU), and low-level waste (LLW). LLW is further differentiated into three classes, A, B, and C, according to increasing of the level of activity. A forth category, commercial greater-than-class-C LLW (listed in 10 CFR 61.55 Tables 1 and 2 for long and short half-life radionuclides, respectively) are not generally suitable for near-surface disposal. This could include operating and decommissioning waste from nuclear power plant and sealed radioisotope sources. The final disposition for this waste is not known. If LLW also contains nonradioactive hazardous material (i.e., that which is toxic, corrosive, inflammable, or explosive) it is termed mixed waste. Mine tailings from uranium mining is still another category of radioactive waste (DOE 1999; Murray 1994). While radioactive cobalt would not ordinarily be found in HLW or TRU, the definitions of these are included below for completion.

TRUs are those containing isotopes, like plutonium, that are above uranium in the periodic table whose half-lives are >20 years. If their level of activity was <100 nanocuries of alpha-emitters per gram of waste material (up from 10 nanocuries/g in 1982), the waste could be disposed of by shallow burial. Otherwise, the waste had to be placed in retrievable storage for eventual transfer to a permanent repository. The level of radioactivity in TRUs is generally low, they generate very little heat, and can be handled by ordinary means without remote control (Eisenbud 1987, Murray 1994).

HLW includes spent fuels which are contained in fuel rods that have been used in a nuclear reactor. These may contain small amounts of transuranic elements. After removal, these rods are placed in pools adjacent to the commercial nuclear power plants and DOE facilities where they were produced. It was originally intended that the fuel rods remain in these pools for only about 6 months to allow for a reduction in radioactivity and temperature and then be transferred to a reprocessing or storage facility. There are no commercial reprocessing facility or permanent disposal facility for HLW operating in the United States. The Nuclear Regulatory Commission (NRC) has issued standards for the disposal of HLW (10 CFR 60), and the DOE is pursuing the establishment of an HLW facility in Yucca Mountain, Nevada. Efforts to establish an HLW facility, which began over 2 decades ago, have experienced many delays. A facility for the permanent disposal of HLW is not projected to be in operation before 2010 (Eisenbud 1987; Murray 1994).

LLWs are officially defined as wastes other than those previously defined. These wastes come from certain reactor operations, manufacturers of radioisotopes used in nuclear medicine and institutions such as hospitals, universities, and research centers. Most LLW contain very little radioactivity and contain practically no transuranic elements. It requires little or no shielding or special handling and may be disposed of by shallow burial. However, some LLW contains sufficient radioactivity as require special treatment. Although NRC regulations for LLW disposal (10 CFR 61) permit shallow land burial, many states have enacted more stringent regulations that require artificial containment of the waste in addition to natural containment (Eisenbud 1987; Murray 1994). The EPA has proposed regulations for LLW disposal that would apply to DOE facilities (EPA 1998b). The Manifest Information Management System (MIMS) maintained by the Idaho National Engineering and Environmental Laboratory, contains information on low-level radioactive waste shipments received at commercial low-level radioactive waste disposal facilities at Barnwell, South Carolina (1/1/86-present), Beaty, Nevada (4/1/86-12/31/92), Richland, Washington (1/1/86-present), and Envirocare, Utah (1/1/98-12/31/99). In 1999, 17 Ci of <sup>57</sup>Co, 1,300 Ci of <sup>58</sup>Co, 0.02 Ci of <sup>59</sup>Co, and 1,080,000 Ci of <sup>60</sup>Co contained in LLW was received at these facilities from academic, industrial, government, and utility generators throughout the United States (INEL 2000). In addition, 4.26 Ci of <sup>57</sup>Co of NARM ("naturally occurring and accelerator-related waste") was received.

At present, DOE stores most of its spent fuel at three primary locations: the Hanford site, Washington, the Idaho National Engineering Laboratory, Idaho, and the Savannah River site, South Carolina. Some spent fuel is also stored at the dry storage facility at Fort St. Vrain in Colorado. Much smaller amounts of spent nuclear fuel stored at other sites were to be shipped to the three prime sites for storage and preparation for ultimate disposal (DOE 1999). The DOE National Spent Fuel Program maintains a spent nuclear fuel data base that lists the total volume, mass and metric tons heavy metal (MTHM) of 16 DOE categories of spent nuclear fuel stored in each of the three locations. The categories having the highest <sup>60</sup>Co activities per spent nuclear fuel canister (decayed to 2030) are 'naval surface ship fuel' and 'naval submarine fuel'. The <sup>58</sup>Co and <sup>60</sup>Co solid waste stored on the Hanford site in 1998 as LLW was 2,600 and 6,900 Ci, respectively (Hanford 1999). In addition, 40 Ci of <sup>60</sup>Co was included in TRU.

In commercial irradiators, additional quantities of <sup>60</sup>Co are added, usually once a year to maintain preferred energy levels of the source (MDS Nordion 2000). <sup>60</sup>Co sources are removed from the facility at the end of their useful life, which is typically 20 years. In general, manufacturers of <sup>60</sup>Co sources guarantee to accept the sources they originally supplied. These old sources may be reencapsulated, reprocessed, or recycled when technically, environmentally, and economically feasible.

COBALT 197

# 6. POTENTIAL FOR HUMAN EXPOSURE

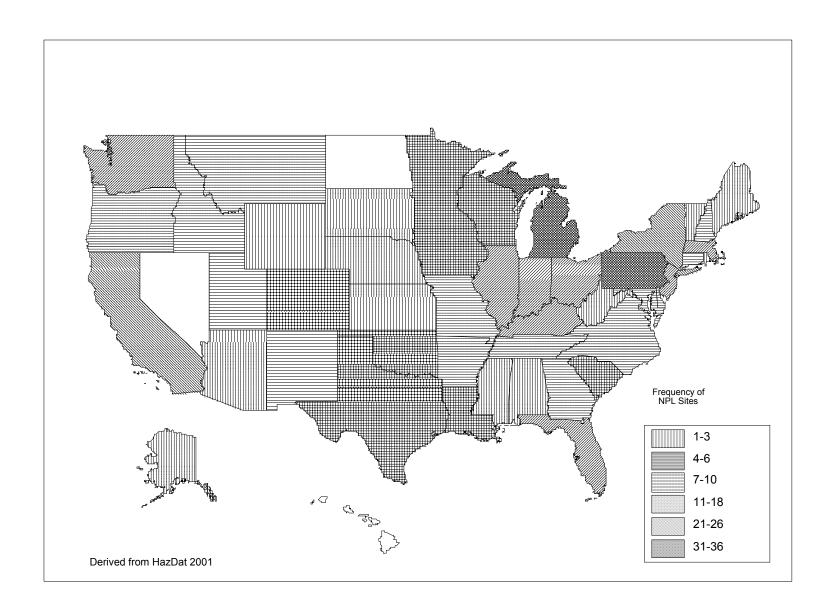
## 6.1 OVERVIEW

Cobalt has been identified in at least 404 of the 1,585 hazardous waste sites that have been proposed for inclusion on the EPA National Priorities List (NPL) (HazDat 2001). <sup>60</sup>Co has been identified in at least 11 of the 1,585 hazardous waste sites that have been proposed for inclusion on the EPA National Priorities List (NPL) (HazDat 2001). However, the number of sites evaluated for cobalt and <sup>60</sup>Co is not known. The frequency of these sites can be seen in Figures 6-1 and 6-2, respectively. Of the cobalt sites, 400 are located within the United States, 1 is located in Guam, and 3 are located in the of Commonwealth of Puerto Rico. All of the sites at which <sup>60</sup>Co has been identified are located within the United States.

Cobalt occurs naturally in the earth's crust, and therefore, in soil. Low levels of cobalt also occur naturally in seawater and in some surface water and groundwater (Smith and Carson 1981). However, elevated levels of cobalt in soil and water may result from anthropogenic activities such as the mining and processing of cobalt-bearing ores, the application of cobalt-containing sludge or phosphate fertilizers to soil, the disposal of cobalt-containing wastes, and atmospheric deposition from activities such as the burning of fossil fuels and smelting and refining of metals (Smith and Carson 1981). Cobalt is released into the atmosphere from both anthropogenic and natural sources. However, emissions from natural sources are estimated to slightly exceed those from manufactured sources. Natural sources include windblown soil, seawater spray, volcanic eruptions, and forest fires. Primary anthropogenic sources include fossil fuel and waste combustion, vehicular and aircraft exhausts, processing of cobalt and cobalt-containing alloys, copper and nickel smelting and refining, and the manufacture and use of cobalt chemicals and fertilizers derived from phosphate rocks (Barceloux 1999; Lantzy and Mackenzie 1979; Nriagu 1989; Smith and Carson 1981). <sup>60</sup>Co and <sup>58</sup>Co may be released to the environment as a result of nuclear research and development, nuclear accidents, operation of nuclear power plants, and radioactive waste dumping in the sea or in radioactive waste landfills.

Cobalt compounds are nonvolatile and cobalt will be emitted to the atmosphere only in particulate form. Their transport in air depends on their form, particle size and density, and meteorological conditions. Cobalt so released will return to land or surface water as wet or dry deposition. Coarse particles, those with aerodynamic diameters  $>2 \mu m$  (such as those obtained during ore processing), may deposit within 10 km from the point of emission; finer particles (such as is obtained from thermal processes) may travel

Figure 6-1. Frequency of NPL Sites with Cobalt Contamination



\*\*\*DRAFT FOR PUBLIC COMMENT\*\*\*

Figure 6-2. Frequency of NPL Sites with Cobalt 60 Contamination



longer distances. It is generally assumed that anthropogenic cobalt originating from combustion sources exists primarily as the oxide; arsenides or sulfides may be released during mining and ore processing (Schroeder et al. 1987). Sediment is the final depository for cobalt released into water. Soluble cobalt released into waterways will sorb to particles and settle into the sediment or be sorbed directly by sediment. It may precipitate out as carbonates and hydroxides or with mineral oxides. It may also sorb to or complex with humic substances in the water column. These processes are sensitive to environmental factors such as pH and the proportion of dissolved cobalt will be higher at low pH. In the case of <sup>60</sup>Co released into an experimental lake in northwestern Ontario, cobalt's half-life in the water column was 11 days; 5% of added <sup>60</sup>Co remained in the water column after 100 days (Bird et al. 1998a). Cobalt can also be transported in dissolved form or as suspended sediment by rivers to lakes and the sea or by ocean currents. The proportion of cobalt transported in each form is highly variable (Smith and Carson 1981). In deep sediment where water is anoxic and hydrogen sulfide is present, some mobilization of cobalt from sediment may occur, probably due to the formation of bisulfides and polysulfides (Bargagli 2000; Brügmann 1988; Finney and Huh 1989; Glooschenko et al. 1981; Knauer et al. 1982; Nriagu and Coker 1980; Shine et al. 1995; Smith and Carson 1981; Szefer et al. 1996; Windom et al. 1989). Cobalt adsorbs rapidly and strongly to soil and sediment in which it is retained by metal oxides, crystalline minerals, and natural organic matter. The mobility of cobalt sediment depends on the nature of the soil or sediment; it increases with decreasing pH and redox potential (Eh) and in the presence of chelating/complexing agents (Brooks et al. 1998; Buchter et al. 1989; King 1988b; McLaren et al. 1986; Schnitzer 1969; Smith and Carson 1981; Swanson 1984; Yashuda et al. 1995).

While cobalt may be taken up from soil by plants, the translocation of cobalt from roots to above-ground parts of plants is not significant in most soils; the transfer coefficient (concentration in plant/concentration in soil) for cobalt is generally 0.01–0.3 (Mascanzoni 1989; Mermut et al. 1996, Smith and Carson 1981). However, in highly acidic soils (pH as low as 3.3) and in some higher plants, significantly higher transfer has been observed (Boikat et al. 1985; Francis et al. 1985; Jenkins 1980; Kloke et al. 1984; Mejstrik and Svacha 1988; Palko and Yli-Hala 1988; Tolle et al.1983; Watabe et al. 1984). The bioaccumulation factors (dry weight basis) for cobalt in marine fish and freshwater fish are ~100–4,000 and <10–1,000, respectively; accumulation is largely in the viscera and on the skin, as opposed to the edible parts of the fish. Cobalt does not biomagnify up the food chain (Barceloux 1999; Evans et al. 1988; Freitas et al. 1988; Smith and Carson 1981).

Atmospheric cobalt is associated with particulate matter. Mean cobalt levels in air at unpolluted sites are generally <1–2 ng/m³. In several open-ocean environments, geometric mean concentrations ranged from 0.0004 to 0.08 ng/m³ (Chester et al. 1991). However in source areas, cobalt levels may exceed 10 ng/m³; the highest average cobalt concentration recorded was 48 ng/m³ at the site of a nickel refinery in Wales (Hamilton 1994; Smith and Carson 1981). By comparison, the Occupational Safety and Health Administration (OSHA) limit for airborne cobalt is 100,000 ng/m³. While <sup>60</sup>Co has been detected in some air samples at the Hanford site and Oak Ridge National Laboratories, levels were not reported (HazDat 2001; PNNL 1996).

The concentrations of cobalt in surface and groundwater in the United States are generally low; <1  $\mu$ g/L in pristine areas and 1–10  $\mu$ g/L in populated areas (Hamilton 1994; Smith and Carson 1981). However, cobalt levels may be considerably higher in mining or agricultural areas. Cobalt levels in most drinking water is <1–2  $\mu$ g/L although levels as high as 107  $\mu$ g/L have been recorded (Greathouse and Craun 1978; Meranger et al. 1981; NAS 1977; Smith and Carson 1981). Little data are available on the levels of  $^{60}$ Co in water. In 1989, subsequent to the largest effluent discharge from the Steam Generating Heavy Water Reactor at Winfrith on the south coast of England,  $^{60}$ Co levels in offshore seawater from 18 sites contained 0.06–2.22 mBq/L (1.6–69 fCi) of particulate  $^{60}$ Co, 0.30–10.3 mBq/L (8–280 fCi) of soluble  $^{60}$ Co(II), and 0.12–1.55 mBq/L (3.2–42 fCi) of soluble  $^{60}$ Co(III) (Leonard et al. 1993a). The U.S. NRC discharge limit is 111,000 mBq/L (NRC 1991).

The average concentrations of cobalt in the earth's crust is 20–25 mg/kg (Abbasi et al. 1989; Merian 1985; Smith and Carson 1981). Most soils contain 1–40 mg cobalt/kg; the average cobalt concentration in U.S. soils is 7.2 mg/kg (Smith and Carson 1981). Soils containing <0.5–3 mg cobalt/kg are considered cobalt-deficient because plants growing on them have insufficient cobalt (<0.08–0.1 mg/kg) to meet the dietary requirements of cattle and sheep. Cobalt-deficient soils are found in some areas of the southeastern and northeastern United States. Soils near ore deposits, phosphate rocks, or ore smelting facilities, and soils contaminated by airport traffic, highway traffic, or other industrial pollution may contain high concentrations of cobalt; concentrations up to 800 mg/kg have been detected in such areas (Kloke et al. 1984; Smith and Carson 1981).

The level of cobalt in most foods is low. However, food is the largest source of exposure to cobalt in the general population. The estimated average daily dietary intake of cobalt in Canada was 11  $\mu$ g/day. Food groups contributing most heavily to this intake were bakery goods and cereals (29.8%) and vegetables

(21.9%). No estimates of the average dietary input of cobalt in the United States was located. People living near mining and smelting facilities or metal shops where cobalt is used in grinding tools may be exposed to higher levels of cobalt in air or soil. Similarly, people living near hazardous waste sites may be exposed to higher levels of cobalt in these media. Contaminated soils pose a hazardous exposure pathway to children because of both hand-to-mouth behavior and intentional ingestion of soil (pica) that contain metals and other contaminants (Hamel et al. 1998). However, much of the cobalt in soil may not be in a form that is available for uptake by the body. People who work in the hard metal industry, metal mining, smelting, and refining or other industries that produce or use cobalt and cobalt compounds may be exposed to substantially higher levels of cobalt, mainly from dusts or aerosols in air. Workers at nuclear facilities, irradiation facilities, or nuclear waste storage sites may be exposed to radioisotopes of cobalt. Exposure would generally be to radiation produced by these isotopes (e.g., gamma radiation from <sup>60</sup>Co).

#### 6.2 RELEASES TO THE ENVIRONMENT

Cobalt has been identified in a variety of environmental media (air, surface water, leachate, groundwater, soil, and sediment) collected at 404 of 1,585 current or former NPL hazardous waste sites (HazDat 2001). <sup>60</sup>Co has been identified in a variety of environmental media (air, surface water, leachate, groundwater, soil, and sediment) collected at 11 of 1,585 current or former NPL hazardous waste sites (HazDat 2001).

According to the Toxic Chemical Release Inventory (TRI), in 1999, total releases of cobalt and cobalt compounds to the environment (including air, water, soil, and underground injection) from 695 reporting facilities that produced, processed, or used cobalt or cobalt compounds were 15,593,293 pounds (TRI99 2001). Table 6-1 lists amounts released from these facilities grouped by state. In addition, 1,296,686 pounds of cobalt and cobalt compounds were transferred off-site by these facilities (TRI99 2001). Starting in 1998, metal mining, coal mining, electric utilities, and Resource Conservation and Recovery Act (RCRA)/solvent recovery industries are required to report to the TRI, industries with potentially large releases of cobalt and cobalt compounds. Industrial sectors producing, processing, or using cobalt that contributed the greatest environmental releases were electric utilities and RCRA/solvent recovery with 32,032 and 51,110 pounds, respectively. Industrial sectors producing, processing or using cobalt compounds that contributed the greatest environmental releases were metal mining and electrical

Table 6-1. Releases to the Environment from Facilities that Produce, Process, or Use Cobalt or Cobalt Compounds

		Reported amounts released in pounds per year <sup>a</sup>								
State <sup>b</sup>	Number of facilities	Air <sup>c</sup>	Water	Under- ground injection	Land	Total on-site released	Total off-site release <sup>e</sup>	Total on and off-site release		
AK	2	25	0	17,000	471,000	488,025	0	488,025		
AL	23	4,510	8,792	0	361,651	374,953	25,268	400,221		
AR	9	726	9	0	20,098	20,833	14,093	34,926		
AZ	12	1,327	5	0	2,437,086	2,438,418	4,912	2,443,330		
CA	28	472	20	0	148,671	149,163	21,603	170,766		
CT	9	1,185	544	0	0	1,729	2,277	4,006		
DE	2	16	101	0	100	217	10,754	10,971		
FL	13	3,277	343	0	78,141	81,761	20,341	102,102		
GA	21	3,387	8	0	331,435	334,830	14,962	349,792		
IA	4	0	0	0	0	0	0	0		
ID	2	52	5	0	47,000	47,057	0	47,057		
IL	35	2,628	13,579	0	77,505	93,712	123,375	217,087		
IN	33	7,730	349	0	360,677	368,756	66,125	434,881		
Ю	20	536	0	0	0	536	2,764	3,300		
KS	5	4,806	0	0	0	4,806	6,162	10,968		
KY	24	3,699	597	0	419,687	423,983	11,853	435,836		
LA	14	311	8,256	8,800	42,299	59,666	42,290	101,956		
MA	9	443	780	0	18	1,241	10,180	11,421		
MD	5	1,007	5	0	0	1,012	1,643	2,655		
ME	1	92	0	0	0	92	2,531	2,623		

Table 6-1. Releases to the Environment from Facilities that Produce, Process, or Use Cobalt or Cobalt Compounds (continued)

			Repo	rted amounts	released in pound	ls per year <sup>a</sup>		
State <sup>b</sup>	Number of facilities	Air <sup>c</sup>	Water	Under- ground injection	Land	Total on-site released	Total off-site release <sup>e</sup>	Total on and off-site release
MI	28	5,448	910	0	130,795	137,153	16,227	153,380
MN	3	20	0	0	0	20	12,859	12,879
МО	7	1,367	7	0	324,027	325,401	250	325,651
MS	11	236	467	16,000	21,945	38,648	3,995	42,643
MT	1	250	0	0	31,000	31,250	1,250	32,500
NC	28	6,999	6,504	0	211,487	224,990	15,378	240,368
ND	2	1,215	250	0	71,400	72,865	52,305	125,170
NE	1	1	28	0	0	29	4,379	4,408
NH	2	0	0	0	0	0	0	0
NJ	12	1,093	49	0	250	1,392	9,509	10,901
NM	10	591	5	0	1,852,990	1,853,586	48,052	1,901,638
٧V	14	1,003	0	1	6,407,931	6,408,935	1,894	6,410,829
NY	13	1,036	59	0	4,200	5,295	14,189	19,484
ЭН	63	6,115	1,107	1,600	323,344	332,166	126,431	458,597
OK	10	1,537	23	0	15	1,575	53,970	55,545
OR	4	1,009	15	0	0	1,024	1,826	2,850
PA	55	13,400	1,309	0	61,176	75,885	201,959	277,844
PR	2	2	0	0	0	2	5,414	5,416
RI	1	1	1	0	0	2	26	28
SC	30	2,306	9,695	0	120,985	132,986	85,353	218,339

Reported amounts released in pounds per year <sup>a</sup>								
State <sup>b</sup>	Number of facilities	Air <sup>c</sup>	Water	Under- ground injection	Land	Total on-site release <sup>d</sup>	Total off-site release <sup>e</sup>	Total on and off-site release
TN	19	5,011	30,726	0	272,945	308,682	25,022	333,704
TX	51	10,102	2,930	4,021	256,594	273,647	120,010	393,657
UT	10	3,014	0	0	48,466	51,480	14,232	65,712
VA	8	1,503	1,079	0	76,000	78,582	7,041	85,623
VI	1	0	0	0	0	0	71	71
WA	2	262	98	0	44,459	44,819	5,664	50,483
WI	21	984	10	0	1,012	2,006	45,715	47,721
WV	13	1,579	709	0	259,020	261,308	42,532	303,840
WY	2	919	0	0	37,856	38,775	0	38,775
Total	695	103,232	89,374	47,422	15,353,265	15,593,293	1,296,686	16,889,979

Source: TRI99 2001

\*\*\*DRAFT FOR PUBLIC COMMENT\*\*\*

<sup>&</sup>lt;sup>a</sup>Data in TRI are maximum amounts released by each facility.

<sup>&</sup>lt;sup>b</sup>Post office state abbreviations are used.

<sup>&</sup>lt;sup>c</sup>The sum of fugitive and stack releases are included in releases to air by a given facility.

<sup>&</sup>lt;sup>d</sup>The sum of all releases of the chemical to air, land, water, and underground injection wells.

eTotal amount of chemical transferred off-site, including to publicly owned treatment works (POTW).

utilities with 11,040,532 and 3,578,014 pounds, respectively. The TRI data should be used with caution because only certain types of facilities are required to report. This is not an exhaustive list.

## 6.2.1 Air

The sources of cobalt in the atmosphere are both natural and anthropogenic (Barceloux 1999). Natural sources include wind-blown continental dust, seawater spray, volcanoes, forest fires, and continental and marine biogenic emissions. The worldwide emission of cobalt from natural sources has been estimated to range from 13 to 15 million pounds/year (Lantzy and Mackenzie 1979; Nriagu 1989). The global atmospheric emission of cobalt from anthropogenic sources is an estimated 9.7 million pounds/year. Therefore, natural sources contribute slightly more to cobalt emission in the atmosphere than anthropogenic sources (Lantzy and Mackenzie 1979). The primary anthropogenic sources of cobalt in the atmosphere are the burning of fossil fuels and sewage sludge, phosphate fertilizers, mining and smelting of cobalt-containing ores, processing of cobalt-containing alloys, and industries that use or process cobalt compounds. Small amounts of cobalt are found in coal, crude oils, and oil shales. Therefore, burning of these fossil fuels for power generation will emit cobalt into the atmosphere. The cobalt contents of the fly ash and flue gases of a coal-burning power plant are approximately 25 mg/kg and 100-700 µg/L, respectively. Gasoline contains <0.1 mg cobalt/kg, but the catalytic converters may contain cobalt; therefore, emissions from vehicular exhaust are also a source of atmospheric cobalt (Abbasi et al. 1989; Holcombe et al. 1985; Ondov et al. 1982; Smith and Carson 1981). Cobalt has been detected in cigarette tobacco and therefore, smoking is a potential source of cobalt emissions that could impact on indoor air quality (Munita and Mazzilli 1986).

Cobalt has been identified in air samples collected at 5 of the 404 current or former NPL hazardous waste sites where it was detected in some environmental media (HazDat 2001). <sup>60</sup>Co has been identified in air samples collected at 2 of the 11 current or former NPL hazardous waste sites where it was detected in some environmental media (HazDat 2001).

Air sampling data were used to estimate <sup>60</sup>Co release from the Savannah River Site (SR) from the plant's start up in 1954 to 1989 (DOE 1991). From this monitoring, it was estimated that 0.092 Ci of <sup>60</sup>Co was released to the atmosphere between 1968 and 1986. SR was a major production facility to the U.S. defense program and included five nuclear reactors, a fuel and target fabrication plant, a naval fuel materials facility, two chemical separation plants, a heavy water production plant, and a laboratory. <sup>60</sup>Co

has also been detected in air samples at the Hanford site and Oak Ridge National Laboratories (HazDat 2001; PNNL 1996).

According to the TRI, in 1999, releases of 103,232 pounds of cobalt and cobalt compounds to air from 695 reporting facilities accounted for 0.7% of the total on-site environmental releases of these substances (TRI99 2001). The industrial sectors contributing the largest release of cobalt and cobalt compounds to air were electrical utilities, primary metals and chemicals. Table 6-1 lists the amounts of cobalt and cobalt compounds released to air from these facilities grouped by state. The TRI data should be used with caution, however, since only certain types of facilities are required to report. This is not an exhaustive list.

## 6.2.2 Water

Compounds of cobalt occur naturally in seawater and in some surface, spring, and groundwater (Smith and Carson 1981). Cobalt is also released into water from anthropogenic sources. While there has been no mine production of cobalt in the United States in recent years, cobalt is a byproduct or coproduct of the refining of other mined metals such as copper and nickel. Waste water from the recovery of cobalt from imported matte or scrap metal, refining of copper and nickel, or during the manufacture of cobalt chemicals are sources of cobalt in water (Smith and Carson 1981). Process water and effluent from coal gasification and residue from solvent-refined coal contain cobalt. The accidental discharge of activated sludge and sewage may be an important sources of cobalamins in waterways, together with bioconcentration by benthic organisms (Smith and Carson 1981). The discharge of waste water by user industries, such as paint and pigment manufacture, also contribute to the release of cobalt into water. In one case, manufacturers of nickel-cadmium batteries operating between 1953 and 1979 discharged cobalt from a battery factory to the Hudson River in Foundry Cove, New York, of which 1.2 MT are estimated to be present in the eastern cove (Knutson et al. 1987). Atmospheric deposition is an additional source of cobalt in water. Lake Huron receives an estimated 76% of its cobalt input from natural sources and 24% from anthropogenic sources. The corresponding estimated values for Lake Superior are 85.4 and 14.6% (Smith and Carson 1981). In these Great Lakes, it therefore appears that natural inputs of cobalt far exceeds anthropogenic ones.

Cobalt has been identified in groundwater and surface water at 242 and 98 sites, respectively, of the 404 NPL hazardous waste sites, where it was detected in some environmental media (HazDat 2001). <sup>60</sup>Co

has been identified in groundwater and surface water at 3 and 2 sites, respectively, of the 11 NPL hazardous waste sites, where it was detected in some environmental media (HazDat 2001).

According to the TRI, in 1999, the reported releases of 89,374 pounds of cobalt and cobalt compounds to water from 695 reporting facilities accounted for 0.6% of the total on-site environmental releases of these substances (TRI99 2001). Table 6-1 lists the amounts of cobalt and cobalt compounds released to water from these facilities grouped by state. As of 1998, TRI no longer separately collects data on substances released indirectly to Publicly-Owned Treatment Works (POTWs), part of which may ultimately be released to surface waters. The TRI data should be used with caution, however, since only certain types of facilities are required to report. This is not an exhaustive list.

<sup>60</sup>Co is present in the low-level aqueous radioactive waste discharges from many nuclear power plants. Cobalt-containing alloys used in piping of nuclear reactors corrode and may be activated, producing <sup>60</sup>Co which accumulates in the reactor and must be periodically decontaminated. A common decontaminating agent includes a reducing metal ion (e.g., vanadium(II)) and a chelating agent (e.g., picolinate) resulting in low-level discharges of uncomplexed <sup>60</sup>Co(II) and complexed <sup>60</sup>Co(III). While soluble ionic and particulate forms predominate, at some sites stable, nonionic trivalent complexes of cobalt are present (Leonard et al. 1993b). For example, in 1987–1989 samples of treated effluent from the Steam Generating Heavy Water Reactor at Winfrith on the south coast of England, the percent of <sup>60</sup>Co as Co(III) picolinate ranged from 6.2 to 75.4%. Between 1978 and 1988, 12 TBq (320 Ci)of <sup>60</sup>Co was released into the Irish Sea by the British Nuclear Fuels reprocessing plant at Sellafield, United Kingdom (McCartney et al. 1994). These discharges are believed to be Co(II) (Leonard et al. 1993a). Both <sup>58</sup>Co and <sup>60</sup>Co are discharged into the Rhone River by the nuclear power plant at Bugey, France. This facility, which consists of a natural Uranium-Graphite-Gas unit and four pressurized water reactor (PWR) units, two of which are cooled by Rhone River water, discharged about 406 and 280 GBq (11.0 and 7.56 Ci) of <sup>58</sup>Co and <sup>60</sup>Co, respectively, in liquid waste during 1986–1990 (Beaugelin-Seiler et al. 1994).

Water sampling data were used to estimate effluent release from the SR Site from the plant's start up in 1954 to 1989 (DOE 1991). From this monitoring, it was estimated that 17.8 Ci (659 GBq) of <sup>60</sup>Co were released into seepage basins and 66.4 Ci (2,460 GBq) were released into streams between 1955 and 1988. In addition, 2.7 Ci (100 GBq) of <sup>58</sup>Co were released into seepage basins between 1971 and 1988; no <sup>58</sup>Co was released into streams. <sup>60</sup>Co has also been reported in surface water and groundwater at the Hanford site and Oak Ridge National Laboratories (HazDat 2001; PNNL 1996). The Columbia River receives

discharges from the unconfined aquifer underlying the Hanford Site via subsurface and surface (riverbank springs) discharges. This aquifer is contaminated by leachate from past waste-disposal practices at the site.

#### 6.2.3 Soil

Cobalt occurs naturally in the earth's crust, and therefore, in soil. However, elevated levels of cobalt in soil may result from anthropogenic activities such as the mining and processing of cobalt-bearing ores, the application of cobalt-containing sludge or phosphate fertilizers to soil, the disposal of cobalt-containing wastes, and atmospheric deposition from activities such as the burning of fossil fuels, smelting, and metal refining (Smith and Carson 1981).

Cobalt has been identified in soil at 203 sites and sediment at 134 sites collected from 404 NPL hazardous waste sites, where it was detected in some environmental media (HazDat 2001). <sup>60</sup>Co has been identified in soil at 6 sites and sediment at 2 sites collected from 11 NPL hazardous waste sites, where it was detected in some environmental media (HazDat 2001).

According to the TRI, in 1999, reported releases of 15,353,265 pounds of cobalt and cobalt compounds to land from 695 reporting facilities accounted for 98.5% of the total on-site environmental releases of these substances (TRI99 2001). An additional 47,422 pounds, accounting for 0.2% of the total on-site environmental releases were injected underground (TRI99 2001). Industrial sectors contributing the largest releases of cobalt and cobalt compounds to land were metal mining and electrical utilities with 11,022,063 and 3,541,815 pounds, respectively. Table 6-1 lists the amounts of cobalt and cobalt compounds released on land from these facilities grouped by state. The TRI data should be used with caution, however, since only certain types of facilities are required to report. This is not an exhaustive list.

#### 6.3 ENVIRONMENTAL FATE

## 6.3.1 Transport and Partitioning

Cobalt compounds are nonvolatile, and thus cobalt is emitted to the atmosphere in particulate form. The transport of cobalt in air depends on its form, particle size and density, and meteorological conditions; it returns to land or surface water as wet or dry deposition. In nonarid areas, wet deposition may exceed dry deposition (Arimoto et al. 1985; Erlandsson et al. 1983). Coarse particles, with aerodynamic diameters  $>2 \mu m$  (such as those obtained during ore processing), may deposit within 10 km from the point of emission; finer particles may travel longer distances. It is the larger particles that may be responsible for elevated local concentrations around emission sources. The mass median diameter for cobalt particles emitted from a power generator with a stack emission controlled by an electrostatic precipitator or scrubber ranged from <2 to  $12 \mu m$ . The mass median diameter of cobalt in the ambient atmosphere is about 2.6  $\mu m$  (Milford and Davidson 1985). Golomb et al. (1997) report average total (wet + dry) deposition rates of cobalt to Massachusetts Bay during the period September 15, 1992 to September 16, 1993. The total deposition rate was  $58 \mu g/m^2$ -year of which  $47 \mu g/m^2$ -year was dry deposition and  $12 \mu g/m^2$ -year was wet deposition. Total cobalt deposition flux at a site in the Rhone delta in southern France in 1988–1989 was  $0.42\pm0.23$  kg/km²-year with 0.15 kg/km²-year in the form of wet deposition (Guieu et al. 1991).

As with most metals, sediment and soil are the final repository for cobalt released into the environment. Cobalt released into waterways will sorb to particles and settle into the sediment or be sorbed directly into the sediment. It can also be transported in dissolved form or as suspended sediment by rivers to lakes and the sea or by ocean currents. Sediment in areas of active sedimentation would receive a large portion of the suspended sediment. In the case of the Peach Bottom Atomic Power Plant where <sup>60</sup>Co is released into the Conowingo Reservoir, an impoundment of the lower Susquehanna River, <20% of the radionuclide is trapped in the reservoir sediment, the rest being transported downstream and into the Chesapeake Bay (McLean and Summers 1990). It is often assumed that the primary mode of transport of heavy metals in aquatic systems is as suspended solids (Beijer and Jernelov 1986). However, in the case of cobalt, the percent that is transported in suspended solids is highly variable. Examples of the percentage of cobalt transported in suspended solids include (water body, percent): Main River (Germany), 33.4–42.2%; Susquehanna River (near its source in New York), 9%; New Hope River (North Carolina), 92%; Yukon River, >98%; Danube Rive (1961–1970), 27.4–85.9%; Columbia River (<sup>60</sup>Co, downstream of the Hanford

site), 95–98%; Strait of Juan de Fuca (Puget Sound, Washington), 11–15%; North Sea, 34%; and Lake Washington (Washington), 0% (Smith and Carson 1981).

In the oxic zones of many surface waters, dissolved cobalt levels decrease with increasing depth. This may be due to cobalt's continuous input into surface water from discharges or to increased adsorption and precipitation of the soluble forms with increasing depth. The fact that cobalt concentration profiles in deep water follow manganese and aluminum profiles strongly suggests that dissolved cobalt is precipitated in the adsorbed state with oxides of iron and manganese and with crystalline sediments such as aluminosilicate and goethite. A part of the cobalt may also precipitates out as carbonate and hydroxide in water. In large unpolluted rivers, cobalt was found in the following forms: dissolved, 1.6–1.7%; adsorbed, 4.7–8.0%; precipitated and coprecipitated with mineral oxides such as iron and manganese, 27.3–29.2%; precipitates such as carbonate and hydroxides, 12.9–19.3%; and in crystalline sediment minerals such as aluminosilicate and goethite, 43.9–51.4%. The corresponding factions in a contaminated river were 12.2, 27.4, 19.2, 14.6, and 26.5%. The higher concentration of organic pollutants in polluted water probably results in the formation of higher concentrations of soluble organic complexes. In a deep sediment where the water was anoxic and contained hydrogen sulfide, some mobilization of cobalt was observed, probably due to the formation of bisulfide and polysulfide complexes (Bargagli 2000; Brügmann 1988; Finney and Huh 1989; Glooschenko et al. 1981; Knauer et al. 1982; Nriagu and Coker 1980; Shine et al. 1995; Smith and Carson 1981; Szefer et al. 1996; Windom et al. 1989).

Cobalt strongly binds to humic substances naturally present in aquatic environments. The lability of the complexes is strongly influenced by pH, the nature of the humic material, and the metal-to-humic substance ratio. The lability of cobalt-humate complexes decreases in time ("aging effect") (Burba et al. 1994). The "aging effect" indicates that after a period of time (~12 hours) complexes that were initially formed are transformed into stronger ones from which the metal ion is less readily dislodged. In the Scheldt Estuary and the Irish Sea, between 45 and 100% of dissolved cobalt was found to occur in these very strong complexes (Zhang et al. 1990). Aquifer material from the contaminated aquifer at a low-level infiltration pit at the Chalk River Nuclear Laboratories in Canada was analyzed to assess the nature of the adsorbed <sup>60</sup>Co using sequential leaching techniques (Killey et al. 1984). Of the sediment-bound <sup>60</sup>Co, <10% was exchangeable, 5–35% was retained by iron oxide, and 55–>90% was fixed. Over 80% of the dissolved <sup>60</sup>Co was present as weakly anionic hydrophilic organic complexes. The average K<sub>d</sub> for <sup>60</sup>Co between particulate matter and Po River (Italy) water was 451 m³/kg over a 2-year monitoring period

(Pettine et al. (1994). The mean  $K_d$  for  $^{60}$ Co in Arctic surface sediment (Kara Sea) where large quantities of radioactive waste by the former Soviet Union was disposed was  $1x10^5$  L/kg (range  $1x10^3$ – $7x10^5$ ), which is comparable to that in temperate coastal regions,  $2x10^5$  L/kg (range,  $2x10^4$ – $1x10^6$ ) (Fisher et al. 1999).

The distribution coefficient of cobalt may vary considerably in the same sediment in response to conditions affecting the pH, redox conditions, ionic strength, and amount of dissolved organic matter (Mahara and Kudo 1981). Uptake of <sup>60</sup>Co from the water column by sediment increased rapidly as the pH was increased from 5 to pH 7–7.5 and then slightly decreases (Benes et al. 1989a, 1989b). Therefore, pH would be an important factor affecting the migration of cobalt in surface water. Uptake was little affected by changes in liquid-to-solids ratio and ionic strength. <sup>60</sup>Co is more mobile in anaerobic marine aquatic environments than in freshwater aerobic ones (Mahara and Kudo 1981). Therefore, <sup>60</sup>Co waste is most suitably stored underground in aerated zones away from possible seawater intrusions. In seawater-sediment systems under anaerobic conditions <sup>60</sup>Co was 250 times more mobile than <sup>60</sup>Co in freshwater-sediment systems under aerobic conditions. Under anaerobic conditions, 30% of the <sup>60</sup>Co added to a sediment-freshwater system was 'exchangeable' and therefore potentially mobile, while under aerobic conditions, 98% of the <sup>60</sup>Co was permanently fixed. Most of the mobile <sup>60</sup>Co produced under anaerobic conditions in seawater consisted of nonionic cobalt associated with low molecular weight organic substances that were stable to changes in pH; the exchangeable <sup>60</sup>Co appeared to be mostly ionic.

Bird et al. (1998b) added <sup>60</sup>Co to the anoxic hypolimnion of a Canadian Shield lake to simulate a nuclear waste scenario where radionuclides entered the bottom waters of a lake, and evaluated its behavior over 5 years. This situation was considered to be a likely pathway by which nuclear fuel waste stored deep underground in the plutonic rock of this region would reach the surface environment via deep groundwater flow into the bottom waters of a lake. It was felt that adding a redox sensitive element such as cobalt to the anoxic hypolimnion might be different from adding it to the epilimnion. Monitoring vertical profiles in the lake established that the cobalt remained confined to the anoxic hypolimnion prior to the fall turnover (first 72 days) when mixing occurred throughout the water column. By day 358, only about 4% of the <sup>60</sup>Co remained in the water column. After the second year, approximately 2% of the <sup>60</sup>Co remained and after 5 years, only 0.4%. These results mirror previous experiments in which the <sup>60</sup>Co was added to the epilimnion, therefore establishing that there is little difference in the overall behavior of cobalt when added to the epilimnion or hypolimnion. The loss rate coefficient of <sup>60</sup>Co was 0.036/day (t<sub>16</sub>=19 days) between days 90 and 131 (lake mixing) during which time, the cobalt sorbed to the

suspended sediment and bottom sediment under anoxic conditions. Loss was to the sediment as there was no hydrological loss from the lake. In the previous experiment in which <sup>60</sup>Co was added to the epilimnion, the initial loss rate coefficient was somewhat higher, 0.056/day (t<sub>1/2</sub>=12 days). Following the initial loss, <sup>60</sup>Co continued to be slowly removed from the water column (loss rate coefficient 0.002/day; t<sub>1/2</sub>=347 days); after 328 days, <sup>60</sup>Co was no longer detectable in the epilimnion. The half life of <sup>60</sup>Co in the water column of an experimental lake in northwestern Ontario was 11 days; 5% of added <sup>60</sup>Co remained in the water column after 100 days (Bird et al. 1998b). The redox potential also affects the behavior of cobalt in sediment. Under moderately reducing conditions, cobalt is released from sediment as Co<sup>2+</sup> and forms CoS in the presence of sulfide. The concentration of cobalt in the bottom water increases as the water becomes more anoxic (Brügmann 1988; Smith and Carson 1981).

The mobility of cobalt in soil is inversely related to how strongly it is adsorbed by soil constituents. Cobalt may be retained by mineral oxides such as iron and manganese oxide, crystalline materials such as aluminosilicate and goethite, and natural organic substances in soil. Sorption of cobalt to soil occurs rapidly (within 1–2 hours). Soil-derived oxide materials absorb were found to adsorb greater amounts of cobalt than other materials examined, although substantial amounts were also adsorbed by organic materials. Clay minerals sorbed relatively smaller amounts of cobalt (McLaren et al. 1986). In addition, little cobalt was desorbed from soil oxides while substantial amounts desorbed from humic acids and montorillonite. In clay soil, adsorption may be due to ion exchange at the cationic sites on clay with either simple ionic cobalt or hydrolyzed ionic species such as CoOH<sup>+</sup>. Adsorption of cobalt onto iron and manganese increases with pH (Brooks et al. 1998). In addition, as pH increases, insoluble hydroxides or carbonates may form, which would also reduce cobalt mobility. Conversely, sorption onto mobile colloids would enhance its mobility. In most soils, cobalt is more mobile than lead, chromium (II), zinc, and nickel, but less mobile than cadmium (Baes and Sharp 1983; King 1988b; Mahara and Kudo 1981; Smith and Carson 1981). In several studies, the  $K_d$  of cobalt in a variety of soils ranges from 0.2 to 3,800. The geometric mean, minimum, median, and maximum K<sub>d</sub>s of <sup>60</sup>Co in 36 Japanese agricultural soils were 1,840, 130, 1,735, and 104,000 L/kg, respectively (Yasuda et al. 1995). The soil properties showing the highest correlation with K<sub>d</sub> were exchangeable calcium, pH, water content, and cation exchange capacity (CEC). In 11 U.S. soils, the mean Freundlich  $K_F$  and n values were 37 L/kg and 0.754, respectively;  $K_F$ values ranged from 2.6 to 363 L/kg and correlated with soil pH and CEC (Buchter et al. 1989). In 13 soils from the southeastern United States whose soil pH ranged from 3.9 to 6.5, cobalt sorption ranged from 15 to 93%; soil pH accounted for 84–95% of the variation in sorption (King 1988b).

Organic complexing agents such as ethylenediaminetetraacetic acid (EDTA), which are used for decontamination operations at nuclear facilities, greatly enhance the mobility of cobalt in soil. Other organic complexing agents, such as those obtained from plant decay, may also increase cobalt mobility in soil. However, both types of complexes decrease cobalt uptake by plants (Killey et al. 1984; McLaren et al. 1986; Toste et al. 1984). Addition of sewage sludge to soil also increases the mobility of cobalt, perhaps due to organic complexation of cobalt (Gerritse et al. 1982; Williams et al. 1985).

Leaching of cobalt has been observed from municipal and low-level radioactive waste sites (Cyr et al. 1987; Czyscinski et al. 1982; Friedman and Kelmers 1988). The mobility of cobalt was assessed in two soils from the Cabriolet and Little Feller event sites at the Nevada Test site as a function of various parameters such as pH, ionic strength, cobalt concentrations, soil solids concentrations, and particle size distribution (DOE 1996). Cobalt was quantitatively sorbed on these soils (at least 90% sorbed) when the pH was above 7 and the solid concentration was at least 20 g/L. The experiments suggest that binding is principally on amphoteric surface-hydroxyl surfaces. Since the pH of these soils is around 8, cobalt would bind strongly under normal environmental conditions. Migration would be severely retarded under all but the most extreme conditions, e.g., pH of 4 or below and high ionic strength soil solutions (approximately 0.1 M). In addition, unrealistically large quantities of water would be need to displace cobalt from the upper layers of the soil profile.

Cobalt may be taken up from soil by plants. Surface deposition of cobalt on leaves of plants from airborne particles may also occur. Elevated levels of cobalt have been found in the roots of sugar beets and potato tubers in soils with high cobalt concentrations (e.g., fly ash-amended soil) due to absorption of cobalt from soil. However, the translocation of cobalt from roots to above-ground parts of plants is not significant in most soils, as indicated by the lack of cobalt in seeds of barley, oats, and wheat grown in high-cobalt soil (Mermut et al. 1996; Smith and Carson 1981). Mermut et al. (1996) found 0.01–0.02 mg/kg in 10 samples of durum wheat grain from different areas of Saskatchewan where surface soil cobalt levels ranged from 3.7 to 16.4 mg/kg. The enrichment ratio, defined as the concentration in a plant grown in amended soil (fly ash) over the concentration in unamended soil, was about 1. Other authors have determined the transfer coefficient (concentration in plant/concentration in soil) for cobalt to be 0.01–0.3. The mean <sup>57</sup>Co soil-plant transfer factors obtained for clover from eight soils over a 4-year period ranged from 0.02 to 0.35, in good agreement with results of other investigators (Mascanzoni 1989). However, in highly acidic soil (pH as low as 3.3), significantly higher than normal concentrations of cobalt were found in rye grass foliage, oats, and barley. For example, cobalt concentrations in rye

grass grown in unlimed soil (pH <5.0) was 19.7 mg/kg compared with 1.1 mg/kg in rye grass grown in limed soil (pH >5.0) (Boikat et al. 1985; Francis et al. 1985; Kloke et al. 1984; Mejstrik and Svacha 1988; Palko and Yli-Hala 1988; Tolle et al. 1983; Watabe et al. 1984). Soil and plant samples taken in the 30-km zone around Chernobyl indicated that <sup>60</sup>Co was not accumulated by plants and mushrooms (Lux et al. 1995). Transfer factors obtained in 1992 ranged from 0.005 to 0.16 and those obtained in 1993 ranged from <0.001 to 0.008.

<sup>60</sup>Co is taken up by phytoplankton and unicellular algae (*Senenastrum capricornutum*) with concentration factors (dry weight) ranging from 15,000 to 40,000 and 2,300 to 18,000, respectively (Corisco and Carreiro 1999). Elimination experiments with the algae indicate a two component biological half-life, 1 hour and 11 days, respectively, and suggests that the cobalt might be absorbed not only on the surface, but also intracellularly. Since these organisms are at the bottom of the food chain, they could play an important role in the trophic transfer of <sup>60</sup>Co released into waterways by nuclear facilities. However, cobalt levels generally diminish with increasing trophic levels in a food chain (Smith and Carson 1981). The low levels of cobalt in fish may also reflect cobalt's strong binding to particles and sediment. The bioaccumulation factors (dry weight basis) for cobalt in marine and freshwater fish are ~100-4,000 and <10–1,000, respectively; accumulation in the muscle of marine fish is 5–500 (Smith and Carson 1981). Cobalt largely accumulates in the viscera and on the skin, as opposed to the edible parts of the fish. In carp, accumulation from water accounted for 75% of 60Co accumulated from both water and food; accumulation from water and food was additive (Baudin and Fritsch 1989). Depuration half-lives were 53 and 87 days for fish contaminated from food and water, respectively. In the case of an accidental release of <sup>60</sup>Co into waterways, the implication is that effects would manifest themselves rapidly since the primary route of exposure is from water rather than food. Uptake of <sup>60</sup>Co by biota in lakes in northwestern Ontario was not affected by the tropic status of the lakes (Bird et al. 1998a). Uptake of <sup>60</sup>Co was very low in whitefish, with concentrations being highest in kidney and undetectable in muscle. Similarly, while accumulation of <sup>60</sup>Co by carp from food was dependent on food type, the transfer factor was very low, approximately 0.01, and no long term bioaccumulation of the radionuclide occurred (Baudin and Fritsch 1987; Baudin et al. 1990). In the experiment described above in which Bird et al. (1998a) added <sup>60</sup>Co to the anoxic hypolimnion of a Canadian Shield lake to simulate a nuclear waste scenario where radionuclides entered the bottom waters of a lake, <sup>60</sup>Co levels in biota were low because of the rapid loss of cobalt to the sediment. Levels in forage fish, minnows, and sculpins were low, <0.3 Bq/g (8 pCi/g) dry weight; an occasional high level, ~4 Bq/g (110 pCi/g) dry weight, in slimy sculpin was thought to reflect the presence of detritus in the gut of the fish. Epilimnion additions of <sup>60</sup>Co

in an earlier study resulted in lower maximum concentrations in fish, 0.07, 0.11, and 0.01 Bq/g (2, 3.0, and 0.3 pCi/g)dry weight in pearl dace, fathead minnows, and slimy sculpins, respectively, when similar quantities of radiocobalt were added to the lake.

Freshwater mollusks have concentration factors of 100–14,000 (~1–300 in soft tissue). Much of the cobalt taken up by mollusks and crustacae from water or sediment is adsorbed to the shell or exoskeleton; very little cobalt is generally accumulated in the edible parts (Amiard and Amiard-Triquet 1979; Smith and Carson 1981). However, the digestive glands of crustaceans, which are sometimes eaten by man, may accumulate high levels of 60Co. Five different species of marine mollusks had whole-body 60Co concentration factors between 6.3 and 84 after 1 month exposure to <sup>60</sup>Co in seawater (Carvalho 1987). The shell accounted for more than half of the body-burden. Among the soft tissue, the gills and viscera had the highest concentrations factors and the muscle had the lowest. Fisher et al. (1996) studied the release of 60Co accumulated in mussels from water and ingested phytoplankton. In both cases, there was a slow and fast component to the release; the rapid release was in the form of fecal pellets if uptake was from food and from desorption from the shell if uptake was from the dissolved phase. Biological halflives obtained in laboratory studies were about 12–21 days from both the shell and soft parts. Higher absorption efficiencies and lower efflux rates were obtained for cobalamine than for inorganic cobalt suggesting that it is a more bioavailable form of cobalt for mussels. Cobalt from fecal pellets is rapidly released into the overlying water and may play a role in its geochemical cycling (Fisher et al. 1996). The concentration of cobalt in clams in the Indian River Lagoon, Florida did not correlate with levels found in either water or sediment (Trocine and Trefry 1996).

# **6.3.2 Transformation and Degradation**

#### 6.3.2.1 Air

There is a paucity of data in the literature regarding the chemical forms of cobalt in air and their transformations in the atmosphere. It is generally assumed that anthropogenic cobalt originating from combustion sources exists primarily as the oxide (Schroeder et al. 1987). In addition, cobalt may be released into the atmosphere as its arsenide or sulfide during ore extraction processes. It is not clear if these species are transformed in the atmosphere. Should a relatively insoluble species such as the oxide be transformed into a more soluble form such as the sulfate, one would expect greater quantities to be washed out of the atmosphere in rain.

### 6.3.2.2 Water

Many factors control the speciation and fate of cobalt in natural waters and sediments. These include the presence of organic ligands (e.g., humic acids, EDTA), the presence and concentration of anions (Cl<sup>-</sup>, OH<sup>-</sup>, CO<sub>3</sub><sup>-2</sup>, HCO<sub>3</sub><sup>-</sup>, SO<sub>4</sub><sup>-2</sup>), pH, and redox potential (Eh). Modeling the chemical speciation of a metal in water depends upon the environmental factors assumed and the stability constants of the various complexes. Mantoura et al. (1978) predicted the equilibrium levels of Co<sup>2+</sup> species in fresh water to follow the order: free  $Co^{+2}$  \$  $CoCO_3$  >  $CoHCO_3^+$  >>  $CoSO_4$  \$ Co•humic acid. However, the mole percent of various cobalt species in a Welsh lake was found to be: free Co<sup>+2</sup>, 76%; CoCO<sub>3</sub>, 9.8%; CoHCO<sub>3</sub><sup>+</sup>, 9.6%; humate complexes, 4.0%; and CoSO<sub>4</sub>, 0.4%. The rank order of species concentration in seawater was estimated to be:  $CoCO_3 > free Co^{+2} > CoSO_4$ \$  $CoHCO_3^+$ . In another model, the speciation of cobalt was completely different with CoCl<sup>+</sup> > free Co<sup>+2</sup> > CoCO<sub>3</sub> > CoSO<sub>4</sub> (Smith and Carson 1981). More recently, Tipping et al. (1998) estimated the equilibrium speciation of cobalt in riverine, estuarine, and marine surface water of the Humber system (England). In all but seawater, cobalt complexes with carbonate (HCO<sub>3</sub><sup>-</sup> and CO<sub>3</sub><sup>2</sup>-) constituted about 70% of dissolved cobalt while the free  $\text{Co}^{2+}$  ion, was a major species, ~25%, which is much lower than the 61% predicted by Mantoura et al. (1978). As the alkalinity of the water increases, the proportion of cobalt complexed with carbonate increases at the expense of free Co<sup>2+</sup>. The proportion of cobalt that exists as the free ion and the carbonate complexes in river water is independent of the level of fulvic acid in the water. In seawater, the carbonate species and the free aquo species assume roughly equal importance. The proportion of dissolved cobalt complexed with fulvic acid decreased with increasing salinity. About 20% of cobalt in seawater was estimated to be present as complexes with sulfate. In a bioconcentration study in which CoCl<sub>2</sub> was initially added to the seawater, at month's end, the cationic form of cobalt was progressively converted into anionic and neutral forms possibly as a result of complexation with organic ligands (Carvalho 1987). Addition of humic acid to natural waters may merely increase the concentration of colloidal dispersed metal rather than form truly soluble humic complexes. In water that contains high organic wastes such as was the case in the Rhone River in France, cobalt was almost completely complexed. Cobalt forms complexes with EDTA that are very stable environmentally. EDTA is often used in agriculture, food and drug processing, photography, and textile and paper manufacturing and therefore, it is a likely constituent of industrial discharges.

The adsorption of cobalt by particulate matter decreases with decreasing pH. This may lead to increased concentrations of dissolved cobalt at low pH. The effect of Eh on the speciation of cobalt has been shown

by the increase in the concentration of dissolved cobalt by orders of magnitude with increasing depth in certain parts of Baltic waters. The increase in the concentration of dissolved cobalt may be due to the formation of soluble bisulfide and polysulfide complexes in the anoxic zones. The residence time of soluble cobalt in seawater has been estimated to range from <1 to 52 years (Brugmann 1988; Knauer et al. 1982; Smith and Carson 1981).

Vitamin  $B_{12}$  is synthesized by 58 species of seven genuses of bacteria as well as blue-green algae and actinomycetes. Consequently, vitamin  $B_{12}$  levels in marine water ranges from very low levels in some open ocean water to much higher levels in some coastal waters. Freshwater environments have comparable levels of vitamin  $B_{12}$ . The high level of cobalamins in coastal water appears to be related to the occurrence of macrophytes in these areas with their high concentrations of vitamin  $B_{12}$ . Cobalamins are released into the water when the organisms die (Smith and Carson 1981).

Alkaline thermal groundwater in granitic areas that have been studied as possible waste disposal sites for radioactive waste (Alaux-Negrel et al. 1993). Water in these areas are characterized by high pH, low  $CO_2$  partial pressure, and generally low redox potential; sulfide concentrations are around a few  $10^{-4}$  to  $10^{-3}$  mol/L. The solubility of cobalt is controlled by the solubility of CoS (log  $K_1$  and log  $K_2$  being 5.7 and 8.7 at 25 EC) and therefore, levels of cobalt are very low,  $10^{-8}$ – $10^{-10}$  mol/L.

The <sup>60</sup>Co (III) picolinate complex that is released into water by some nuclear reactors does not break down immediately on release into seawater, but rather can coexist with the <sup>60</sup>Co (II) forms for lengthy periods in the environment (Leonard et al. 1993a, 1993b). Studies indicate that several processes occur to the Co(III) organic complexes, including reduction to the inorganic form, sorption of both species to particulate matter, and transformations of the uncomplexed species. It is possible that this more soluble and uncharged form of radiocobalt will increase the dispersion of <sup>60</sup>Co from its point of discharge.

#### 6.3.2.3 Sediment and Soil

The speciation of cobalt in soil or sediment depends on the nature of the soil or sediment, concentration of chelating/complexing agents, pH, and redox potential (Eh) of the soil. Dissolved cobalt may be absorbed by ion exchange and other mechanisms, or form complexes with fulvic acids, humic acid, or other organic ligands in soil. The humic and fulvic complexes of cobalt are not very stable compared with those of copper, lead, iron, and nickel. The speciation of cobalt in sediment from nine sites in the Red Sea, a sea

which is unique in that it has no permanent streams flowing into it, was assessed using a sequential extraction technique (Hanna 1992). The mean percentages contained in the various fractions were: exchangeable, 5.5%; carbonate, 5%; Fe/Mn oxides, 24%; organic, 30.4%; sulfides, 13%; and lithogenous, 22%. While the mean concentration of cobalt in the sediment increased from 0.003 to 0.006 ppb between 1934 and 1984, its distribution among the different phases did not change appreciably.

The reduction of soil Eh, which may occur when soil is flooded or in deeper layers of soil that are oxygen-depleted, may change the speciation of cobalt. This may result in the reduction of soil iron and manganese and the subsequent release of adsorbed cobalt from the mineral oxides. Similarly, a decrease in soil pH may result in the solubilization of precipitated cobalt and the desorption of sorbed cobalt resulting in increased cobalt mobility (Smith and Carson 1981). Co<sup>2+</sup> may also be oxidized to Co<sup>3+</sup> by manganese oxides, a common component of soils and aquifer material, with subsequent surface precipitation (Brusseau and Zachara 1993). This process may affect transport of cobalt in the subsurface environment.

EDTA complexes of cobalt are very stable and are likely to form in soils containing EDTA. EDTA is widely used as a decontaminating agent at nuclear facilities. Although cobalt-EDTA complexes are adsorbed by some soils, the mobility of cobalt in soil may increase as a result of complex formation (Schnitzer 1969; Smith and Carson 1981; Swanson 1984). <sup>60</sup>Co that is disposed of in shallow land trenches have sometimes been found to migrate more rapidly than expected from the disposal sites. Organic chelating agents are frequently present at these sites and would possibly increase the solubility and transport of the radionuclide.

Bacterial action can affect the mobility of a substance by mediating reactions or by participating in reactions that lower the pH. Another way of influencing radionuclide mobility is by degrading complexing agents used in cleaning reactors (e.g., citric acid), thereby releasing the radionuclide. However, experiments on the fate and transport of cobalt released upon the biodegradation of the complexing ligand indicate that results are not always predictable; the means of ligand removal and the geochemical environment are important factors that must be considered (Brooks et al. 1998).

### 6.4 LEVELS MONITORED OR ESTIMATED IN THE ENVIRONMENT

Cobalt concentrations in environmental media, including food and human tissue, have been exhaustively tabulated by Smith and Carson (1981) and Young (1979). The International Agency for Research on Cancer (IARC 1991) contains reviews of more recent studies, but is primarily focused on occupational exposures and body burdens of cobalt.

## 6.4.1 Air

Atmospheric cobalt is associated with particulate matter. Mean cobalt levels in air at unpolluted sites are generally <1 to 2 ng/m<sup>3</sup> (Hamilton 1994, Smith and Carson 1981). At the South Pole, cobalt levels of 0.00049±0.00015 ng/m<sup>3</sup> were recorded in 1974–1975 (Maenhaut et al. 1979). Geometric mean cobalt levels in several open-ocean environments ranged from 0.0004 to 0.08 ng/m<sup>3</sup> (Chester et al. 1991). The average annual PM-10 (particles with diameters <10 μm) cobalt concentration at Nahant, Massachusetts (near Boston) in 1992–1993 was 1.7 ng/m<sup>3</sup> (Golomb et al. 1997). Half of the cobalt was contained in fine particles (<2.5 μm) and half in coarse particles (2.5–10 μm). The mean cobalt level in southern Norway in 1985–1986 (n=346) was 0.10 ng/m<sup>3</sup> with 35% of the samples falling below the detection limit of 0.04 ng/m<sup>3</sup> (Amundsen et al. 1992). Atmospheric cobalt levels in industrial settings may exceed 10 ng/m<sup>3</sup>. The highest recorded average cobalt concentration in air was 48 ng/m<sup>3</sup> at the site of a nickel refinery in Clydach, Wales (Smith and Carson 1981). Some ambient atmospheric levels of cobalt are given in Table 6-2. These data show the contribution of anthropogenic sources in increasing the level of cobalt in the ambient air. Typical occupational cobalt levels are 0.01–1.7 mg/m³ (Barceloux 1999; IARC 1991). While <sup>60</sup>Co has been detected in air samples at the Hanford site and Oak Ridge National Laboratories, levels were not reported (HazDat 2001; PNNL 1996). In 1995, the concentration of <sup>60</sup>Co in air at the Hanford site was below the detection limit in over 88% of the air samples.

### 6.4.2 Water

The concentrations of cobalt in surface water and groundwater in the United States are generally low, <1  $\mu$ g/L in pristine areas and 1–10  $\mu$ g/L in populated areas (Hamilton 1994; Smith and Carson 1981). However, cobalt levels may be considerably higher in mining or agricultural areas. Levels as high as 4,500  $\mu$ g/L were reported in Mineral Creek, Arizona, near a copper mine and smelter; levels of 6,500  $\mu$ g/L were reported in the Little St. Francis River, which receives effluent from cobalt mining and

Table 6-2. Concentration of Cobalt in the Atmosphere

Location	Possible	Concentration	Linita	Tuno	Reference
Location  Ambient levels - remote	source/activity	Concentrationa	Units	Туре	Reference
South Pole, 1974–1975	Cruetal meterial	0.00049±0.00015	na/m³	MoonteD	Maenhaut et al. 1979
•	Crustal material	0.00049±0.00013	ng/m³ ng/m³		Chester et al. 1991
Open-ocean		0.0004-0.00	rig/iii	range	Chester et al. 1991
North Atlantic		0.006-0.09	ng/m³	Range	Smith and Carson 1981
Baltic Sea, 1983		0.09, 0.01–0.43	ng/m³	Mean, range	Hasanen etal. 1990
Remote sites		0.001-0.9	ng/m³	Range	Schroeder et al 1987
Ambient levels - rural/suburban/urban					
Rural sites		0.08-10.1	ng/m³	Range	Schroeder et al 1987
Massachusetts, Nahant, 1992–1993		1.7	ng/m³	Annual mean	Golomb et al. 1997
Urban sites					Schroeder et al 1987
United States		0.2–83	ng/m³	Range	
Canada		1–7.9			
Europe		0.4–18.3			
Texas state average (1978–1982)		2.0	ng/m³	Mean	Wiersema et al. 1984
Illinois, urban air (<2.5 Fm; 2.5–10 Fm)					Sweet et al. 1993
Bondville, III (rural)	Background	0.2; 0.1	ng/m³	Mean (fine; coarse)	
Southeast Chicago	Steel mills	0.4; 0.4			
East St. Louis	Smelters	0.5; 0.4			
Washington, DC (1974)	Urban area	1.1	ng/m³	Mean	Smith and Carson 1981
Ambient levels - industrial					
Maryland, Baltimore Harbor Tunnel (1973–1974)					Ondov et al. 1982
Air outside	Vehicular exhaust	0.8–1.9	ng/m³	Range	
Air inside	Vehicular exhaust	2.2–5.3			
Ohio, Cleveland	Be-Cu alloy and other industrial activities	610	ng/m³	Maximum	Smith and Carson 1981

6. POTENTIAL FOR HUMAN EXPOSURE

Texas, El Paso (1978–1982)

Industrial

127

ng/m³ Maximum Wiersema et al. 1984

Table 6-2. Concentration of Cobalt in the Atmosphere (continued)

	Possible				
Location	source/activity	Concentration <sup>a</sup>	Units	Туре	Reference
Texas, Houston (1978–1982)	Urban area	81	ng/m³	Maximum	Wiersema et al. 1984
Arizona, Tucson					Smith and Carson 1981
Urban	Copper smelting	1.9	ng/m³	Mean	
Rural		0.7			
Maryland, Chalk Point Generator	Coal-burning power plant	3.86	ng/m³	Mean	Smith and Carson 1981
Wales, Clydach	Nickel refining	48, 3–300	ng/m³	Mean, range	Smith and Carson 1981
Wales, Llausamlet and Trebanos	Towns near Clydach	3.8		Mean	Smith and Carson 1981
Occupational air levels					
Northern Italy, exposure survey, 1991, area monitoring (n=259)	Diamond abrasive mfg.				Mosconi et al. 1994a
	Mould-filling	220, 47–960	ng/m³	Median, range	
	Sintering	101.5, 32–240			
	Grinding	22, 15–45			
	Mechanical- working	20, 12–44			
	Grinding	5, 2.5–94			
	Tool production	6, 5–47			
	Hard metal alloy filing	2, 0.8–3			
	Other	2.7, 2.3–15			
Northern Italy, exposure survey, 1991, personal sampling (n=259)	Diamond abrasive mfg.				Mosconi et al. 1994a
, ,	Mould-filling	382, 76–2,600	ng/m³	Median, range	
	Sintering Grinding Mechanical- working	309, 238–413 230, 82–690 40, 7.1–65		J	
	Grinding Tool production	9.3, 1.5–178 17, 4–28			

## 6. POTENTIAL FOR HUMAN EXPOSURE

Table 6-2. Concentration of Cobalt in the Atmosphere (continued)

Location	Possible source/activity	Concentration <sup>a</sup>	Units	Туре	Reference
	Hard metal alloy filling	5, 1–107			
	Other	50, 10–290			
Japan, personal sampling, hard metal tool manufacture, 8-hour TWA, 356 workers (n=935)	Powder preparation				Kumagai et al. 1996
	rotation	459, 7–6,390	Fg/m³	Mean, range	
	full-time	147, 26–378			
	Press				
	rubber	339, 48–2,910			
	steel	47, 6–248			
	Shaping	97, 4–1,160			
	Sintering	24, 1–145			
	Blasting	2, 1–4			
	Electron discharging	3, 1–23			
-	Grinding	45, 1–482			

geomean = geometric mean; SD = standard deviation; TWA = time weighted average

milling operations (Smith and Carson 1981). Eckel and Jacob (1988) analyzed U.S. Geological Survey (USGS) data for 6,805 ambient surface water stations and estimated the geometric mean and median dissolved cobalt concentration as 2.9 and 2.0 µg/L, respectively. Mean cobalt levels reported in seawater range from 0.078 µg/L in the Caribbean Sea to 0.39 µg/L in the Indian Ocean (Hamilton 1994). Vitamin B<sub>12</sub> is synthesized by bacteria, macrophytes, blue-green algae, and actinomycetes and cobalt levels in oceans often correlate with biological productivity. In the Baltic Sea, dissolved cobalt levels that are 1.0 ng/L near the surface, rise precipitously to 71.0 ng/L at a depth of 200 m (Brügmann 1988). The rise in dissolved cobalt is coincident with the onset of anoxic conditions and the presence of hydrogen sulfide, indicating that soluble bisulfide and polysulfide complexes may be present. Some cobalt levels reported in water are given in Table 6-3.

In a 1962–1967 survey, cobalt was detected in 2.8% of 1,577 U.S. raw surface waters from which drinking water is derived; the detection limit was 1  $\mu$ g/L and the maximum concentration was 48  $\mu$ g/L (NAS 1977). Of 380 U.S. finished drinking waters, only 0.5% contained cobalt levels exceeding 1  $\mu$ g/L; the maximum concentration found was 29  $\mu$ g/L (NAS 1977). These values are higher than the respective median and maximum levels of <2.0 and 6.0  $\mu$ g/L found in Canadian finished drinking water (Meranger et al. 1981). Meranger et al. (1981) tested source water and drinking water in 71 municipalities across Canada and concluded that, in general, both surface water and groundwater used for drinking water supplies contain negligible amounts of cobalt. Greathouse and Craun (1978) analyzed 3,834 grab samples of household tap water from 35 geographical areas in the United States for 28 trace elements. Cobalt was found in 9.8% of the samples at concentrations ranging from 2.6 to 107  $\mu$ g/L. It is not clear whether these higher levels could indicate that cobalt was picked up in the distribution system. In the earlier National Community Water Supply Study (2,500 samples), 62% of the samples contained <1  $\mu$ g Co/L; the average and maximum cobalt concentrations were 2.2 and 19  $\mu$ g/L, respectively (Smith and Carson 1981). Cobalt was not detected (detection limit 8  $\mu$ g/L) in a 1982–1983 survey of drinking water in Norway that covered 384 waterworks serving 70.9% of the Norwegian population (Flaten 1991).

The mean concentrations of cobalt in rain is around  $0.03-1.7~\mu g/L$  with levels generally ranging from  $0.002~\mu g/L$  at Enewetak Atoll to about  $2.9~\mu g/L$  in the Swansea Valley, Wales (Arimoto et al. 1985; Dasch and Wolff 1989; Hansson et al. 1988; Heaton et al. 1990; Helmers and Schrems 1995; Nimmo and Chester 1993; Nimmo and Fones 1997; Smith and Carson 1981). The highest recorded level of cobalt in precipitation was  $68.9~\mu g/L$  in the vicinity of a nickel smelter in Monchegorsk in the Russian Arctic (Reimann et al. 1997). An analysis of rain in the Mediterranean and urban and coastal sites in northwest

Table 6-3. Cobalt Levels in Water

Nature/location of water	Level	Units	Туре	Reference
Sea water				
Florida (Indian River Lagoon) (43 sites)	0.031, 0.006–0.050	μg/L	Mean, range	Trocine and Trefry 1996
California (Baja) 2–45 km offshore (n=11)	0.022–0.17	nM	Range	Sañudo-Wilhelmy and Flegal 1996
<100 m off shore (n=11)	0.11–0.59			
Agean Sea, 1994; 8 sites (dissolved)	0.168–0.632, 1.917	nM	Range of means, maximum	Voutsinou-Taliadouri 1997
Baltic Sea (Gotland Deep site)				Brügmann 1988
10 m	1.0	ng/L	Mean (dissolved Co)	
50 m	1.0			
100 m	3.5			
150 m	4.2			
200 m (anoxic)	71.0			
235 m (anoxic)	49.2			
Seawater background	0.04	μg/L		Bargagli 2000
Seawater	0.27	μg/L	Mean	Abbasi et al. 1989
Fresh surface water				
Freshwater background	0.05	μg/L		Bargagli 2000
U.S. ambient surface water (6,805 stations)	<2.9, 2.0	μg/L	Mean, median	Eckel and Jacob 1988
Five Great Lakes waters	ND-0.09	μg/L	Range	Rossmann and Barres 1988
Japan, unpolluted lake	<0.004	μg/L		Nojiri et al. 1985
Norway, 11 rivers	0.94	μg/L	Maximum	Flaten 1991
Streams near populated areas	1–10	μg/L	Range	Smith and Carson 1981
Streams in agricultural and mining areas	11–50	μg/L	Range	Smith and Carson 1981
Suspended solids in rivers	7–94	mg/kg	Range	Smith and Carson 1981
Groundwater				
Canada (Chalk River nuclear waste site)	0.0001-0.002	μg/L		Cassidy et al. 1982
Colorado (Denver) - shallow groundwater, (n=30)	<1 (<1–9)	μg/L	Median, range	Bruce and McMahon 1996

<sup>\*\*\*</sup>DRAFT FOR PUBLIC COMMENT\*\*\*

## 6. POTENTIAL FOR HUMAN EXPOSURE

Table 6-3. Cobalt Levels in Water (continued)

Nature/Location of water	Level	Units	Туре	Reference
Drinking water				_
Canadian drinking water (71 municipalities)				Meranger et al. 1981
Raw:	<2.0	μg/L	Median	
Treated:	<2.0			
Distributed:	#2.0			
Precipitation				
Massachusetts, 1984 (12 events)	0.045 (0.008), 0.02–0.12	μg/L	Mean (SD), range	Dasch and Wolff 1989
Rhode Island (rain/snow), 1985 (n=269)	0.038 (0.067)	ppb	Median (mean)	Heaton et al. 1990
	0.001-0.80		Range	
Western Mediterranean, 1988- 1989				Nimmo and Chester 1993
Total cobalt	0.029–0.134, 0.043	μg/L	Range, mean	
Labile cobalt	0.009–0.104, 0.025			
Organic cobalt	ND-0.613, 0.019			
Arctic (7 sites in Finland, Norway, Russia)	<0.02–1.07, 3.32	μg/L	Median range, maximum	Reimann et al. 1997
Russia (Monchegorsk), nickel smelter	11.8, 68.9		Median, maximum	

ND = not detected; SD = standard deviation

England showed that about 33–44% of the cobalt occurred as very stable dissolved organic complexes (Nimmo and Chester 1993; Nimmo and Fones 1997).

As it was pointed out in Section 6.3.2.2, <sup>60</sup>Co discharged from the Steam Generating Heavy Water Reactor at Winfrith on the south coast of England was shown to be largely in the form of the nonionic trivalent complex, <sup>60</sup>Co(III) picolinate. The <sup>60</sup>Co(III) species is not immediately reduced to the more particle-reactive divalent form and both oxidation states may coexist for long periods of time in the environment. The proportion of the more soluble and mobile <sup>60</sup>Co(III) would be expected to increase with time and distance from the point of discharge. Shoreline water samples (n=22) taken in 1987–1988 at two locations in the vicinity of the discharge from the Steam Generating Heavy Water Reactor at Winfrith contained 0.3–16.2 mBq/L (8–437 fCi/L) of particulate <sup>60</sup>Co, 2.8–44.4 mBq/L (76–1,200 fCi/L) of soluble <sup>60</sup>Co(II), and 0.2–4.8 mBq/L (5–130 fCi/L) of soluble <sup>60</sup>Co(III) (Leonard et al. 1993). The percent of the soluble <sup>60</sup>Co present as Co(III) ranged from 4.3 to 18.6%. In 1989, in conjunction with the largest discharge of effluent from the plant, offshore seawater samples from 18 sites contained 0.06–2.22 mBq/L (fCi/L) of particulate <sup>60</sup>Co, 0.30–10.3 mBq/L (8.1–278 fCi/L) of soluble <sup>60</sup>Co(II), and 0.12–1.55 mBq/L (3.2–41.9 fCi/L) of soluble <sup>60</sup>Co(III). The percent of the soluble <sup>60</sup>Co present as Co(III) ranged from 6.0 to 28.6%.

## 6.4.3 Sediment and Soil

Cobalt is the 33<sup>rd</sup> most abundant element in the earth's crust. Its average concentrations in the earth's crust and in igneous rocks are 20–25 and 18 mg/kg, respectively (Abbasi et al. 1989; Merian 1985; Smith and Carson 1981). Trace metals in soils may originate from parent rock or from anthropogenic sources, primarily fertilizers, pesticides, and herbicides. Most soils contain 1–40 mg cobalt/kg. The average cobalt concentration in U.S. soils is 7.2 mg/kg (Smith and Carson 1981). Soils containing <0.5–3 mg cobalt/kg are considered cobalt-deficient because plants growing on them have insufficient cobalt (<0.08–0.1 mg/kg) to meet the dietary requirements of cattle and sheep. Cobalt-deficient soils include the humus podzols of the southeastern United States, and the podzols, brown podzolic soils, and humus groundwater podzols in the northeastern parts of the United States. The cobalt content of surface soils from 13 sites in the brown and dark brown soil zones of southwestern Saskatchewan ranged from 3.7 to16.0 mg/kg and only in one case was the soil appreciably elevated above the corresponding parent material (Mermut et al. 1996). Fertilizers used in this agricultural area contained 0.12–102 mg Co/kg, median 5.7 mg/kg.

Mean cobalt concentrations in surface soil from nine sites on two active volcanic islands off of Sicily ranged from 5.1 to 59.0 mg/kg (Bargagli et al. 1991). Soils near ore deposits, phosphate rocks, or ore smelting facilities, and soils contaminated by airport traffic, highway traffic, or other industrial pollution may contain much higher concentrations of cobalt; concentrations up to 800 mg/kg have been detected in such areas (Kloke et al. 1984; Smith and Carson 1981). Soils around the large copper-nickel smelters in Sudbury, Ontario have been shown to contain high levels of cobalt. Fifty kilometers from the smelters, cobalt levels in surface soil were 19 mg/kg. These levels increased to 48 mg/kg at 19 km, 33 mg/kg at 10 km and 42–154 mg/kg between 0.8 and 1.3 km from the smelter (Smith and Carson 1981). Soils around a cemented tungsten carbide tool grinding factory contained cobalt levels as high as 12,700 mg/kg, almost 2,000 times the average in U.S. soils (Abraham and Hunt 1995). However, neighborhood soils between 30 and 160 meters from the factory only contained 12–18 mg Co/kg.

Unpolluted freshwater sediment contains about the same levels of cobalt as does cobalt-sufficient soil, generally <20 mg/kg (Smith and Carson 1981). In the Hudson River Estuary, cobalt levels in suspended sediment was an order of magnitude higher than in bottom sediment (Gibbs 1994). This can be attributed to the finer grain size of suspended sediment or local sources. Cobalt levels in core samples (surface to 42 cm) from the Upper St. Lawrence Estuary were independent of depth indicating the lack of any recent significant anthropogenic releases (Coakley et al. 1993). Cobalt levels in sediment are shown in Table 6-4.

No broad-based monitoring studies of <sup>60</sup>Co or other radioactive cobalt isotopes in soil or sediment were found in the literature. Soil samples from the O-horizon taken from three sites in the 30-km zone around Chernobyl in 1992 and again in 1993, contained 14–290 and 4.5–245 Bq/kg (380–7,800 and 120–6,620 pCi/kg) dry weight of <sup>60</sup>Co, respectively (Lux et al. 1995). The Columbia River receives radiological contaminants along the Hanford Reach primarily through seepage of contaminated groundwater (PNNL 1996). The regional median concentration of <sup>60</sup>Co was highest along this reach. <sup>60</sup>Co activity in a sediment cores in water off of Southampton in southern England contained up to 28 Bq/kg (760 pCi/kg) in the upper 3 cm; no activity was found below 12.5 cm (Croudace and Cundy 1995). Discharges of treated effluent occurred on closing a steam generating heavy water reactor west of where the sampling was done. The maximum discharge occurred in 1980–1981.

# 6. POTENTIAL FOR HUMAN EXPOSURE

Table 6-4. Cobalt Levels in Sediment

Nature/location of sediment	Level	Units	Туре	Reference
Freshwater				_
Polluted lakes and rivers	0.16-133	mg/kg	Range	Smith and Carson 1981
Lake Ontario near Miesissaqua, Canada	4.1–19.8	mg/kg	Range	Glooschenko et al. 1981
Hudson River, Foundry Cove, 1983, Ni-Cd battery plant, 1953–1979, Surficial (0–5 cm) sediment, 16 sites	18–700	mg/kg	Range	Knutson et al. 1987
Estuaries and Marine				
Hudson River Estuary (0–80 km from ocean), 1991				Gibbs 1994
Bottom sediment	1–13	mg/kg	Range	
Suspended sediment (near surface)	30–140			
Upper St. Lawrence Estuary, 1989–1990				Coakley et al. 1993
Core C168	3.1 (0.6)	mg/kg	Mean (SD)	
Cores LE and LO	2.7 (0.5)			
Massachusetts, New Bedford Harborcore (0–25 cm)				Shine et al. 1995
Outer Harbor	7.03, 3.64–9.79	mg/kg	Mean, range	
Inner Harbor	6.38, 2.62–10.52			
Buzzards Bay (control site)	4.76, 1.64–8.19			
Indian River Lagoon, Florida (43 sites)	2.3, 0.4–6.3	mg/kg	Mean, range	Trocine and Trefry 1996
Gulf of Mexico				Villanueva and Botello 1998
Coastal areas (11 sites)	12.30–36.26	mg/kg	Range of means	
Continental shelf (3 sites)	6.39-21.00			
Antarctica (Ross Sea) continental shelf (n=12)	19, 0.10–13	mg/kg	Mean, range	Bargagli 2000
Northern Arctic Alaska, continental shelf (n=136)	9, 3.3–18	mg/kg	Mean, range	Bargagli 2000
Chukchi Sea, northeast Alaska (31 stations, surficial sediment)	32.7, 19–74	mg/kg	Mean, range	Naidu et al. 1997
Baltic Sea, southern, off Poland (surficial sediment)	0.69–18.10	mg/kg	Range	Szefer et al 1996
Baltic Sea (Gotland Deep site)	19, 11–33	mg/kg	Mean, range	Brügmann 1988

SD = standard deviation

#### 6.4.4 Other Environmental Media

The cobalt content of plants depends on the plant, the cobalt content of the soil and numerous environmental factors. The mean cobalt concentration reported for terrestrial plants was  $0.48~\mu g/g$ , while the mean and median levels for freshwater vascular plants were  $0.48~and~0.32~\mu g/g$ , respectively (Outridge and Noller 1991). The median cobalt level in freshwater vascular plants from polluted waters was about the same as in unpolluted waters,  $0.37~\mu g/g$ , although extremely high levels of cobalt, up to  $860~\mu g/g$ , was reported in one species, *Myriophyllum verticillatum*, from central Ontario lakes. Grasses normally contain  $0.2-0.35~\mu g/g$  of cobalt, but grasses from cobalt deficient regions contain  $0.02-0.06~\mu g/g$  of cobalt (Hamilton 1994). Durum wheat grown in southeastern Saskatchewan contained 0.01-0.02~m g/kg dry weight (Mermut et al. 1996). In view of the cobalt content of the soil and the fact that almost half of the cobalt in fertilizers used in the area was in a readily available form, the uptake of cobalt by wheat was negligible.

<sup>60</sup>Co levels in plants and mushrooms in the 30-km zone around Chernobyl were mostly below the detection limit in samples obtained in 1992 and 1993; the highest activity recorded was 3.9 Bq/kg (110 pCi/kg) dry weight in *Athyrium filix femina* (Lux et al. 1995).

Eel and a freshwater fish from three Dutch polder lakes contained 2.5–25.0 and 2.50–5.63 mg cobalt/kg wet weight, respectively, (Badsha and Goldspink 1988). Ocean fish and rock crabs caught near dump sites off New York City, New Haven, Connecticut, and Delaware Bay contained 10–40 and 16.0 μg/kg, respectively in muscle tissue (Greig and Jones 1976). In a study of the levels and distribution of 14 elements in oceanic seabirds, the concentration of cobalt, an essential element, appeared to be highly regulated with over 80% of the body burden residing in the skeleton. The mean cobalt concentration in the livers of 11 seabird species ranged from 0.048 to 0.078 μg/g dry weight, and cobalt had the lowest coefficient of variation in the different species of the elements studied (Kim et al. 1998a). In another study in Antarctica, mean cobalt levels in fish and amphipods were 0.11–0.14 and 1.01 μg/g dry weight, respectively, while those in the tissue of penguin and other sea birds ranged from 0.09 to 0.11 μg/g (Szefer et al. 1993). The concentration of cobalt in the tissue of 14 bluefin tuna caught by various commercial fishing vessels off Newfoundland was essentially the same, 0.01±0.004 μg/g (Hellou et al. 1992a). Similarly, in a broad survey of contaminant levels in nine species of fish and fiddler crabs from 11 sites in the lower Savannah River, Georgia and the Savannah National Wildlife Refuge, mean cobalt levels among different species and sites were statistically indistinguishable (Winger et al. 1990). These

and other studies indicate that cobalt does not biomagnify up the food chain (Smith and Carson 1981). While cobalt was found in high levels in sediment from the Tigris River in Turkey and in low levels in the water, it was not detected in two species of fish (Gümgüm et al. 1994).

Some female birds sequester metals into their eggs under certain conditions, a phenomenon that may jeopardize the developing embryos. The geometric mean concentrations of cobalt in tern eggs collected from coastal New Jersey in 1971 and 1982 were 0.48 and 0.50 mg/kg, respectively. Unlike the levels of seven other common metals (e.g., mercury, cadmium, copper, lead, manganese, nickel, and zinc), the level of cobalt in tern eggs (and in the environment) showed no decline over the 11-year period (Burger and Gochfeld 1988).

Table 6-5 shows the levels of cobalt in foods items and food categories from different countries. The level of cobalt in most Canadian foods was low; items with the highest concentrations were waffles (76 ng/g), corn cereal (74 ng/g), and potato chips (70 ng/g) (Barceloux 1999; Dabeka and McKenzie 1995). Green leafy vegetables and fresh cereals are the richest sources of cobalt (0.2–0.6 μg/g dry weight), while dairy products, refined cereals, and sugar contain the least cobalt (0.1–0.3 μg/g dry weight). The levels of cobalt were determined in 50 different food items, mainly meat, fish, fruit, vegetables, pulses, and cereals on the Swedish market during the years 1983–1990 (Jorhem and Sundström 1993). Beef liver and seeds were fairly high in cobalt and fish, fruit, and root and leafy vegetables were under 0.01 mg cobalt/kg fresh weight. The cobalt levels in mg/kg fresh weight were highest in alfalfa seeds, 0.86; linseed, 0.56; milk chocolate, 0.34; dark chocolate, 0.24; white poppy seeds, 0.30; blue poppy seeds, 0.15; soya beans, 0.084; green lentils, 0.054; and beef liver, 0.043. The cobalt content of 20 brands of alcoholic and nonalcoholic beer widely consumed in Spain ranged from 0.16 to 0.56 μg/L with a median of 0.39 μg/L (Cameán et al. 1998). Cobalt, which was at one time added to beer to increase the foam head, has been associated with cardiomyopathies in heavy beer drinkers.

Cobalt is present in various consumer products including cleaners, detergents, and soaps, which have resulted in dermatitis in sensitive individuals (Kokelj et al. 1994; Vilaplana et al. 1987). Tobacco contains about <0.3–2.3 µg Co/g dry weight and approximately 0.5% of the cobalt appears in mainstream smoke (Barceloux 1999; Munita and Mazzilli 1986; Ostapczuk et al. 1987; Stebbens et al. 1992).

Table 6-5. Cobalt Levels in Food

Food item	Level	Units <sup>a</sup>	Туре	Reference
Infant formulas/milk				
Evaporated milk (n=21)	0.74, 0.52–2.6	µg/kg⁵	Median, range	Dabeka 1989
Ready-to-use formula (n=49)	0.53, 0.21–5.2	µg/kg⁵	Median, range	Dabeka 1989
Milk-based (n=33)	0.40, 0.21–0.99			
No added iron (n=6)	0.36, 0.21–0.61			
Added iron (n=27)	0.87, 0.41–0.99			
Soy-based (n=16)	2.27, 1.71–5.2			
Concentrated liquid formula (n=50)	2.27,0.25-11.8	µg/kg⁵	Median, range	Dabeka 1989
Milk-based (n=34)	1.57, 0.25–3.11			
No added iron (n=20)	1.06, 0.25–1.77			
Added iron (n=14)	2.59, 2.03–3.11			
Soy-based (n=16)	4.33, 2.7–11.8			
Powdered formula (n=64)	9.54, 2.6–53	µg/kg⁵	Median, range	Dabeka 1989
Milk-based (n=36)	4.96, 2.6–10.6			
No added iron (n=23)	4.24, 2.6–9.6			
Added iron (n=13)	8.26, 5.1–10.6			
Soy-based (n=28)	20.0, 10.6–53			
Agricultural crops				
Cabbage, United States	0.2	mg/kg <sup>c</sup>	Typical level	NAS 1977
Corn seed, United States	0.01	mg/kg <sup>c</sup>	Typical level	NAS 1977
Fruits, 12 types, Poland	0.01-0.02	mg/kg	Range	Bulinski et al. 1986
Lettuce, Sweden 1983–1990 (n=7)	0.002, 0.006	mg/kg	Mean, maximum	Jorhem and Sundström 1993
Lettuce, United States	0.2	mg/kg <sup>c</sup>	Typical level	NAS 1977
Onions, 11 Danish sites (n=110)	1.51, 0.119–5.1	μg/kg	Median, range	Bibak et al. 1998a
Peas, 10 Danish sites (n=93)	4.6, 0.57–17	μg/kg	Median, range	Bibak et al. 1998b
Potatoes, Sweden (n=8)	0.008, 0.017	mg/kg	Mean, maximum	Jorhem and Sundström 1993
Spinach, United States	0.4–0.6	mg/kg <sup>c</sup>	Typical range	NAS 1977
Strawberries, Sweden (n=10)	0.004, 0.010	mg/kg	Mean, maximum	Jorhem and Sundström 1993
Vegetables, 30 types, Poland	0.008-0.032	mg/kg	Range	Bulinski et al. 1986
White flour, United States	0.003	mg/kg <sup>c</sup>	Typical level	NAS 1977

<sup>\*\*\*</sup>DRAFT FOR PUBLIC COMMENT\*\*\*

# Table 6-5. Cobalt Levels in Food (continued)

Food item	Level	Units <sup>a</sup>	Туре	Reference
Meat, fish, beverages			-	
Beef, Sweden (n=3)	0.001, 0.001	mg/kg	Range, maximum	Jorhem and Sundström 1993
Beef liver, Sweden (n=3)	0.043, 0.074	mg/kg	Range, maximum	Jorhem and Sundström 1993
Beef kidney, Sweden (n=3)	0.008, 0.010	mg/kg	Range, maximum	Jorhem and Sundström 1993
Beer, Spain, 20 brands	0.39, 0.16–0.56	Fg/L	Median, range	Cameán et al. 1998
Cocoa, Germany	1.31	mg/kg <sup>c</sup>		Ostapczuk et al. 1987
Coffee (whole), South Africa	0.93	mg/kg <sup>c</sup>		Horwitz and Van der Linden 1974
Coffee (whole), Germany (61% water extractable)	0.11–0.31	mg/kg <sup>c</sup>	Range	Ostapczuk et al. 1987
Fish, Sweden, 10 varieties (n=40)	<0.001–.008, 0.020	mg/kg	Range of mean, maximum	Jorhem and Sundström 1993
Pork, Sweden (n=36)	0.001, 0.012	mg/kg	Range, maximum	Jorhem and Sundström 1993
Pork liver, Sweden (n=36)	0.010, 0.023	mg/kg	Range, maximum	Jorhem and Sundström 1993
Pork kidney, Sweden (n=36)	0.004, 0.011	mg/kg	Range, maximum	Jorhem and Sundström 1993
Tea (whole), South Africa	0.2	mg/kg <sup>c</sup>		Horwitz and Van der Linden 1974
Tea (whole), Germany (40% water extractable)	0.18–6.7	mg/kg <sup>c</sup>	Range	Ostapczuk et al. 1987
Food categories				
Bakery good/ cereals, Canada (n=24)	10.9, 75.7	μg/kg	Median, maximum	Dakeba and McKenzie 1995
Beverages, Canada (n=7)	5.9, 9.1	µg/kg	Median, maximum	Dakeba and McKenzie 1995
Fats and oils, Canada (n=3)	<2.6, 37.6	μg/kg	Median, maximum	Dakeba and McKenzie 1995
Fish, Canada (n=6)	18.6, 14.3–29.4	μg/kg	Median, range	Dakeba and McKenzie 1995
Fruits and fruit juices, Canada (n=25)	<6.6, 35.7	μg/kg	Median, maximum	Dakeba and McKenzie 1995
Meat and poultry, Canada (n=18)	<5.5, 38.2	μg/kg	Median, maximum	Dakeba and McKenzie 1995

# 6. POTENTIAL FOR HUMAN EXPOSURE

Table 6-5. Cobalt Levels in Food (continued)

Food item	Level	Units <sup>a</sup>	Туре	Reference
Milk and milk products, Canada (n=13)	<1.4, 18.9	μg/kg	Median, maximum	Dakeba and McKenzie 1995
Soups, Canada (n=4)	5.6, 8.5	µg/kg	Median, maximum	Dakeba and McKenzie 1995
Sugar and candy, Canada (n=7)	<0.4, 3.5	μg/kg	Median, maximum	Dakeba and McKenzie 1995
Vegetables, Canada (n=38)	2.4, 18.1	μg/kg	Median, maximum	Dakeba and McKenzie 1995

<sup>&</sup>lt;sup>a</sup>produce on a fresh weight basis, unless otherwise specified. <sup>b</sup>as sold <sup>c</sup>dry weight basis

**Table 6-6. Cobalt Content of Miscellaneous Substances** 

Substance/source	Level	Units	Туре	Reference
Bituminous coal used for power generation	6.4	mg/kg	Median	Rubin 1999
Coal, United States	~5	mg/kg	Mean	Smith and Carson 1981
Fly ash	~25	mg/kg	Mean	Smith and Carson 1981
MSW Incinerator ash, Mississippi				Buchholz and Landsberger 1995
Fly ash (n=30)	11.3–13.5	Fg/g	Range	
Bottom ash (n=30)	65.2-90.3			
Combined ash (n=30)	24.8-30.5			
MSW Incinerator ash, United States 1987	,			Mumma et al. 1990
Fly ash (n=5)	18.2-54.0	Fg/g	Range	
Bottom ash (n=7)	13.5–35.1			
Combined ash (n=8)	11.2-43.4			
Compost, Toronto				Evans and Tan 1998
Residential compost	8.1, 3.2–12	mg/kg	Median, range	
Greenhouse finished compost	6.1±1.03		Mean ± SD	
Sewage sludge				
16 large U.S. cities	11.3, 6.08–29.1	mg/kg	Median, range	Gutenmann et al 1994
32 U.S. cities	7.2, 2.4–30.1	mg/kg	Median, range	Mumma et al. 1984
Cow manure (comparison)	6.1	mg/kg		Mumma et al. 1984
Miscellaneous soil amendments <sup>a</sup>				Raven and Loeppert 1997
Compost	3.55, 3.57	mg/kg	Individual means	
Diammonium phosphate	3.24, 0.68			
Dolomite	0.33			
Manure	2.23			
Monoammonium phosphate	0.78, 3.38			
Rock phosphate, Tilemsi	19.6			
Rock phosphate, North Carolina	<0.08			
Sewage sludge, Austinite	4.10			
Sewage sludge, Milorganite	4.07			
Triple superphosphate	6.61, 2.24			
Street dust, New York City	8.7–12.9	μg/g	Range	Fergusson and Ryan 1984

<sup>&</sup>lt;sup>a</sup>The rest of the 24 fertilizers and soil amendments tested were below the detection limit (typically <0.07 ppm).

MSW = municipal solid waste; SD = standard deviation

The cobalt content of sewage sludge, incinerator ash, fertilizers, soil amendments, and other substances appears in Table 6-6. The concentration of cobalt in U.S. coal averages about 5 mg/kg, levels in crude oil and fuel oil ranges from 0.001 to 10 and 0.03 to 0.3 mg/kg and those in gasoline are <0.1 mg/kg (Smith and Carson 1981). Cobalt levels were below the detection limit of 0.05 ppm dry weight in all but 1 of 26 samples of composted yard waste, sewage sludge, and municipal solid waste samples nationwide in 1991. The one positive sample of composted yard waste contained 1.53 ppm of cobalt (Lisk et al. 1992).

### 6.5 GENERAL POPULATION AND OCCUPATIONAL EXPOSURE

Exposure of the general population to cobalt occurs through inhalation of ambient air and ingestion of food and drinking water. In general, exposure from food is much greater than from drinking water, which in turn, is much greater than from air. From the limited monitoring data available, the average concentration of cobalt in ambient air in the United States is approximately  $0.4 \text{ ng/m}^3$ . However, levels may be orders of magnitude higher in source areas. Therefore, exposure to cobalt in air will vary substantially from nonsource areas to areas with cobalt-related industries. Similarly, the median cobalt concentration in U.S. drinking water is <2.0 µg/L; however, values as high as 107 µg/L have been reported in surveys of water supplies (Smith and Carson 1981). Therefore, exposure from drinking water may vary considerably from one location to another. In Canada, the daily cobalt intake of the average adult from drinking water is #2.6 µg; this could increase to 10 µg for those living in areas with the highest cobalt levels (Meranger et al. 1981).

General population exposure to cobalt from food is highly variable and normally higher than intake from drinking water. Most of the cobalt ingested is inorganic; vitamin B<sub>12</sub>, which occurs almost entirely in food of animal origin, constitutes only a very small fraction of cobalt intake. The cobalt intake in food has been estimated to be 5.0–40.0 μg/day (Jenkins 1980). The daily cobalt intake, including food, water, and beverages of two men that were followed for 50 weeks was much higher, 310 and 470 μg (Smith and Carson 1981). The estimated average daily cobalt intake from diet in Canada was 11 μg/day; the intake varied between 4 and 15 μg/day between the various age/sex groups (see Table 6-7) (Barceloux 1999; Dabeka and McKenzie 1995). The contributions of various food groups to cobalt intake in this study were (category, contribution of dietary intake): bakery goods and cereals, 29.8%; vegetables, 21.9%; beverages, 9.8%; milk and milk products, 9.4%; meat and poultry, 9.1%; soups, 6.4%; fruit and fruit juices, 5.0%; sugar and candies, 2.8%; fish, 2.7%; fats and oils, 2.2%; and miscellaneous, 1.1%. The

## 6. POTENTIAL FOR HUMAN EXPOSURE

Table 6-7. Mean Daily Dietary Intake of Cobalt for Selected Population Groups in Canada

Group	Mean daily intake (μg/day)	
1–4 years	7	
5–11 years	10	
12–19 years; male	14	
12–19 years; female	10	
20–39 years; male	15	
20-39 years; female	9	
40–65 years; male	12	
40-65 years; female	9	
65+; male	10	
65+; male	8	

Source: Dabeka and McKenzie 1995

average daily intakes of cobalt in France was estimated to be 29 µg/day (Biego et al. 1998). In this study, foods were divided into nine categories. The foods accounting for the greatest contributions of cobalt intake were milk and dairy products, fish-crustaceans, and condiments-sugar oil, respectively contributing 32, 20, and 16% to the daily intake. The U.S. Department of Agriculture (USDA) conducted a special exploratory study in 1985–1986 to determine the concentration of trace metals in tissue of health livestock and poultry randomly selected from those slaughtered. Between 0.6 and 5.9% of samples in the 11 production classes had levels of cobalt that exceeded the lowest reliable quantitation level of 0.15 ppm and the mean of positive samples ranged from 0.20 to 0.23 ppm in all classes but heifer/steer, which had a level of 1.92 ppm (Coleman et al. 1992). Cobalt, which has been added to beer to increase the foam head, has been associated with cardiomyopathies in heavy beer drinkers. However, according to a recent Spanish study, the low levels of cobalt presently found in beer do not make a significant contribution to the total cobalt intake in heavy beer drinkers (Cameán et al. 1998). Smokers may be exposed to cobalt in mainstream smoke, but the level of exposure has not been assessed (Barceloux 1999).

Since cobalt and other heavy metals have been used on hand painted china, a study was conducted to see whether these metals are released into food under acidic conditions. Forty-six samples of porcelain dinnerware from Europe or Asia that was manufactured before the mid-1970s and had hand-painted designs over the glaze was filled with 4% acetic acid to within 7 mm of the rim and analyzed after 24 hours (Sheets 1998). Of these, 36 samples released <0.02  $\mu$ g/mL of cobalt and 10 released 0.020–2.9  $\mu$ g/mL. The Food and Drug Administration (FDA) has not established dinnerware extraction limits for cobalt.

Data are lacking on the levels of cobalt in tissues and fluids of the general populations in the United States; values from various countries are given in Table 6-8. As can be seen from this table, cobalt concentrations are greatest in nail, hair, and bone. The differences in cobalt levels in similar human tissues (e.g., hair, nail) in different countries may be due to differences in dietary and living habits and levels of cobalt in food (Takagi et al. 1988). The total amount of cobalt in the body of an adult as vitamin  $B_{12}$  is about 0.25 mg, of which 50–90% in contained in the liver (IARC 1991).

Surgical implants for knee and hip replacements often use cobalt-containing alloys which may lead to elevated cobalt levels in body fluids. Indeed, cobalt levels in serum and urine have been used as an index

Table 6-8. Cobalt Levels in Human Tissues and Fluids

Tissue or fluid	Level	Unitsa	Туре	Reference
Urine, U.S., NHANES, representative	0.36, 0.11–0.89		Geomean,	CDC 2001
population (n=1007)	0.30, 0.11-0.09	µg/L	10 <sup>th</sup> –90 <sup>th</sup>	CDC 2001
,			percentile	
Urine, The Netherlands	<0.2–1.2	μg/L	Range	Bouman et al. 1986
Urine, Sweden	0.5, 0.1–2.2	μg/L	Mean, range	Alexandersson 1988
Urine, Denmark (3 reference groups)				Poulsen et al 1994
Unexposed control females (n=46)	1.5, LOD-20.5	$nmol^{\mathtt{b}}$	Mean, range	
Unexposed males (n=12)	0.9, LOD-2.31			
Unexposed females (n=11)	5.9, LOD-25.02			
Urine, hip arthroplasty patients, observed 7–15 years (n=17)	0.9–1.05	μg/L	Range	IARC 1991
Urine, hip arthroplasty patients, observed 5–15.5 years (n=10)	3.8	μg/L	Mean	IARC 1991
Urine, 48 metal sharpening workers in 12 Italian factories	0–40.3, 86	μg/L	Range of means, maximum	Imbrogno et al. 1994
Urine, 12 female cobalt powder sintering workers, Italy				Ferdenzi et al. 1994
Monday, before shift	25, 1–51	μg/L	Mean, range	
Friday, before shift	29, 3–159			
Friday, end-of shift	85, 6–505			
After 3-week holiday	11, 4–34			
Urine, Italian workers wet grinding of hard metal tools (end of shift)				Sesana et al. 1994
Factory A no local exhausts (n=3)	138.3 (108), 123.7 (74)	μg/L	Mean (SD) Monday, Friday	
Factory B local exhausts (n=5)	15.3 (7.7), 24.4 (14.1)			
Factory C local exhausts (n=3)	48.2 (7.3), 74.7 (13)			
Urine, Northern Italy, 1991, occupational exposure survey, 314 exposed people				Mosconi et al. 1994
Diamond abrasive production				
Mould-filling	320, 587, 39–2,100	µg/L	Median, mean, range	
Sintering	168, 193, 102–390			
Grinding	61, 151, 34–520			
Mechanical-working	50, 67, 143–165			
Grinding	15, 32. 0.8–730			
Tool production	12, 19, 0.8–100			

# 6. POTENTIAL FOR HUMAN EXPOSURE

240

Hard metal alloy filling 5, 5, 0.8–18
Other 1, 2.9, 0.8–72

Table 6-8. Cobalt Levels in Human Tissues and Fluids (continued)

Tissue or fluid	Level	Units <sup>a</sup>	Туре	Reference
Blood, Denmark, porcelain factory		_		Raffn et al. 1988
Plate painters, off work for 6 weeks (n=46)	8.05, 1.70–22.1	nmol/L	Mean, range	
Plate painters, working 4 weeks (n=46)	36.7, 3.40–407			
Top glaze painters (unexposed) (n=51)	4.04, <1.70–10.2			
Urine, Denmark, porcelain factory				Raffn et al. 1988
Plate painters, off work for 6 weeks (n=46)	81.8, <1.70–445	nmol/L	Mean, range	
Plate painters, working 4 weeks (n=46)	1,308, 37.4–14,397			
Top glaze painters (unexposed) (n=51)	16.0, <1.70–234			
Plasma, Sweden	0.1–1.2	μg/L	Range	Alexandersson 1988
Whole Blood, Denmark (3 Reference groups)				Poulsen et al 1994
Unexposed control females (n=46)	4.1, <1.7–10.2	nmol/L	Mean, range	
Unexposed males (n=12)	3.1, <1.7–6.8			
Unexposed females (n=11)	7.6, <1.7–30.5			
Lung, Sweden				Gerhardsson et al. 1988
rural	0.007	mg/kg	Mean	
urban	0.011			
Liver, New Zealand (n=96)	0.120	mg/kg	Mean	IARC 1991
Tissue, Japan				Yamagata et al. 1962
Pectoral muscle	0.016	mg/kg	Mean	
Rib bone	0.036			
Stomach	0.021			
Liver	0.017			
Brain	0.0055			
Urinary bladder	0.0055			
Kidney	0.012			
Aorta	0.021			
Nails				Takagi et al. 1988
Canada (n=40)	0.09	mg/kg	Mean	
India (n=100)	0.06			
Japan (n=252)	0.17			

## 6. POTENTIAL FOR HUMAN EXPOSURE

Table 6-8. Cobalt Levels in Human Tissues and Fluids (continued)

Tissue or fluid	Level	Units <sup>a</sup>	Туре	Reference
Poland (n=49)	0.04			
U.S. (n=71)	0.06			
Adipose tissue	0.035-0.078	mg/kg	Range	EPA 1986
Hair				Takagi et al. 1986
Canada (n=92)	0.043	mg/kg	Mean	
India (n=255)	0.051			
Japan (n=457)	0.18			
Poland (n=46)	0.022			
United States (n=55)	0.047			
Hair, Italy				Vienna et al. 1995
Male biology students (n=20)	0.007, 0.001–0.07	mg/kg	Geomean, range	
Female biology students (n=20)	0.017, 0.001–0.28			
Hair, Pakistan				Ashraf et al 1995
rural (n=28)	2.05, 0.10-4.80	mg/kg	Mean, range	
urban (n=39)	3.86, 1.10–5.90			

<sup>&</sup>lt;sup>a</sup>fresh weight, unless otherwise specified

geomean = geometric mean; NHANES = Nation Health and Nutrition Examination Survey; SD = standard deviation

bcreatinine basis

of prosthesis wear. In some cases, significant increases in cobalt levels have been observed, while in other cases, elevations were much lower or only sporadic (IARC 1991). These differences have been ascribed to greater release rates from metal to metal than metal to polyethylene articular surfaces as well to differences in the cobalt-containing alloys.

There are several reports of cobalt exposure among occupational groups. The concentrations of cobalt in the air of hard metal manufacturing, welding, and grinding factories may range from 1 to 300  $\mu$ g/m³, compared to normal atmospheric levels of 0.4–2.0 ng/m³ (Burr and Sinks 1989; Haddad and Zikovsky 1985; Koponen et al. 1982; Lichtenstein et al. 1975). The maximum OSHA permissible level is  $100~\mu$ g/m³. The concentration of cobalt in the dust of an electric welding factory was 4.2  $\mu$ g/g compared to its normal dust level of 0.1–1.0  $\mu$ g/g (Baumgardt et al. 1986). The higher rate of exposure to cobalt for occupational groups is also reflected in the higher cobalt content in tissues and body fluids of living and deceased workers in this group. The levels of cobalt in the urine of workers in the hard metal industry varied with the levels of cobalt concentration in the working atmosphere. At a concentration of 0.09 mg/m³, the urinary excretion of cobalt exceeded normal values by orders of magnitude. When the cobalt concentration in the working atmosphere was 0.01 mg/m³ or lower, urinary cobalt excretion was 4–10 times higher than normal level (Alexandersson 1988; Scansetti et al. 1985). At high exposure levels, the cobalt concentration in blood was 20 times higher than normal; in the low exposure group, it was only slightly higher than in the control group (Alexandersson 1988).

An extensive survey of workers potentially exposed to cobalt in the Bergamo Province in northern Italy in 1991 identified 403 exposed workers in different production areas (Mosconi et al. 1994a). Significant cobalt exposure occurred especially for operators working in diamond abrasive production, and in particular, in mould filling and sintering units where environmental limits are regularly exceeded. Exposure in tool production, tool sharpening, and hard metal alloy filling is much more restrained. Occupational cobalt air levels and urinary excretion levels recorded in the survey appear in Tables 6-2 and 6-8.

In the hard metal industry in Japan, Kumagai et al. (1996) found that mean 8-hour time weighted averages (TWAs) were  $>50 \mu g/m^3$  for workers involved in powder preparation (rotation), powder preparation (full-time), rubber press, and shaping operations; mean exposure levels were 459, 147, 339, and 97  $\mu g/m^3$ , respectively. Workers involved in the manufacture and maintenance of hard metal and stellite blades in Finland were exposed to breathing zone cobalt concentrations ranging from 0.002 to 0.240 mg/m<sup>3</sup>, with a

geometric mean of 0.017 mg/m³ (Linnainmaa et al. 1996). The average proportion of water soluble cobalt in airborne cobalt was 68% (range 14–100%). Wet grinding was not sufficient to adequately control cobalt levels and coolant cobalt levels were high. In a group of 12 factories in Italy in which 48 workers were tested who were exposed to cobalt in operations such as sharpening with diamond grinding stones, the mean concentration of cobalt in air was 21.2 and 137.7  $\mu$ g/m³ (PEL-TWA 100  $\mu$ g/m³) in work places with and without dust ventilation, respectively (Imbrogno et al. 1994).

Ferdenzi et al. (1994) obtained a correlation between Friday TWA air cobalt levels and Friday end-ofshift urine levels among women in the powder sintering industry. The mean urine cobalt level in the factories ranged from 0 to 40.3 µg/L and the maximum was 86 µg/L. The average urinary cobalt level among workers using wet/mixed sharpening methods was 4 times higher than those using dry sharpening methods. Cobalt urine levels increased rapidly during a shift and dropped significantly overnight and after a weekend. Gallorini et al. (1994) found that the ratio of inorganic to organic cobalt in the urine of hard metal workers was 2.3 compared to 1.01 in controls; the ratio was constant over the range of urinary cobalt levels analyzed (180–1,254 µg/L). Exposure to cobalt during the wet grinding of hard metal tools (Widia tools) used in the wood industry produced exposure to cobalt above the PEL-TWA of 100 µg/m<sup>3</sup> (Sesana et al. 1994). However, exhausts near the grinding wheels were shown to substantially reduce exposure levels (see Table 6-8). In the processing department of a small company producing carbide tip saw blades for the woodworking industry, area air sampling showed that exposure levels were low in all departments except tip grinding where wet and dry tip grinding areas contained 55 and 21 µg/m<sup>3</sup> of cobalt, respectively (Stebbins et al. 1992). Respirable cobalt levels ranged from 2 to 28 µg/m<sup>3</sup>. Wet grinding is a traditional method for controlling dust during grinding. However, some coolants may contain significant concentrations of cobalt (in this case, 61–538 mg/mL) that can contribute to exposure during grinding. Among cobalt blue dye plate painters in a porcelain factory in Denmark, the blood and urine cobalt levels were, respectively, 2–4 and 5–15 times higher than in control groups (Raffn et al. 1988). Similarly, lungs taken from deceased, occupationally exposed workers also had higher levels of cobalt than lungs from control groups. Lungs of deceased hard metal industry workers in Sweden contained 2.5–4 times higher levels of cobalt than control lungs (Gerhardsson et al. 1988). Similarly, the lungs of coal miners from England contained 6 times higher cobalt levels than control lungs (Hewitt 1988).

Workers at nuclear facilities, irradiation facilities, or nuclear waste storage sites may be accidentally exposed to radioisotopes of cobalt. Also, workers using cobalt isotopes in tracer studies, in calibration or

other devises, or  $^{57}$ Co in Mössbauer spectroscopy, may be exposed to radiocobalt. Exposure would generally be to radiation produced by these isotopes (e.g., gamma radiation from  $^{60}$ Co). Patients receiving  $^{60}$ Co radiotherapy will obviously be exposed to its radiation. According to the NRC (1999), the collective intake of  $^{60}$ Co by ingestion and inhalation at power reactors in 1998 was 352  $\mu$ Ci (13 MBq) for 25 intake records and 27,000  $\mu$ Ci (1,000 MBq) for 281 intake records (NRC 1999). The collective intake at fuel fabrication facilities was 0.486  $\mu$ Ci (0.180 MBq) for 502 intake records.

#### 6.6 EXPOSURES OF CHILDREN

This section focuses on exposures from conception to maturity at 18 years in humans. Differences from adults in susceptibility to hazardous substances are discussed in Section 3.7 Children's Susceptibility.

Children are not small adults. A child's exposure may differ from an adult's exposure in many ways. Children drink more fluids, eat more food, breathe more air per kilogram of body weight, and have a larger skin surface in proportion to their body volume. A child's diet often differs from that of adults. The developing human's source of nutrition changes with age: from placental nourishment to breast milk or formula to the diet of older children who eat more of certain types of foods than adults. A child's behavior and lifestyle also influence exposure. Children crawl on the floor, put things in their mouths, sometimes eat inappropriate things (such as dirt or paint chips), and spend more time outdoors. Children also are closer to the ground, and they do not use the judgment of adults to avoid hazards (NRC 1993).

As with adults, most children are exposed to cobalt largely through their diet. Dabeka and McKenzie (1995) estimated that the dietary cobalt intake by Canadian children ages 1–19 ranged from 7 to 14 mg/day (see Table 6-7). Milk constitutes a larger part of children's diets than that of adults, and infants may consume infant formula. Cow's milk contains 0.3–0.8 ng/g cobalt (Dakeba 1989). The levels of cobalt in human milk from Nigeria, Zaire, Guatemala, Hungary, Philippines, and Sweden ranged from 0.15 (Hungary) to 1.4 µg/g (Philippines), median 0.32 µg/g (Nriagru 1992). Garg et al. (1993) reported much lower cobalt levels in three samples of human milk in India, 2.42 ng/g, and 5.07 ng/g in cow's milk. Dakeba (1989) determined cobalt levels in various infant formulas (see Table 6-5). Milk-based infant formulas and evaporated milk contained <1 ng/g of cobalt on a "ready-to-use" basis. Milk-based formulas with added iron contained about twice those with no added iron and soy-based formulas contained about 5 times more. The influence of added iron suggests that the cobalt in formula is not primarily from vitamin B<sub>12</sub>. Using literature values of cobalt in food, Dakeba also estimated that infants

0–12 months old ingest an average of 0.52  $\mu$ g Co/kg-day (3.93  $\mu$ g/day) from food and water and that for an infant, 0–12 months old, the total dietary cobalt intake would range from 0.42  $\mu$ g/kg-day (3.39  $\mu$ g/day) for a breast or milk-based formula fed infant to 1.0  $\mu$ g/kg-day (7.33  $\mu$ g/day) for an infant fed soy-based formula powder. The recommended dietary allowance for Canadian infants is 0.012  $\mu$ g/day cobalt as vitamin B<sub>12</sub>. In a 1967 study of the total dietary intake of some trace elements, excluding drinking water, of institutionalized children aged 9–12 in 28 U.S. cities, cobalt intake ranged from 0.297 to 1.767 mg/day with a mean value of 1.024 mg/day (Murthy et al. 1971).

Cobalt exposure in communities near mining and smelting facilities or metal shops where cobalt is used in grinding tools are a public health concern, especially for infants and children. Since cobalt remains in the surface soil indefinitely and long past land uses may be forgotten, people may not realize that they are living in areas where high levels of cobalt may occur in soil. Contaminated soils pose a particular hazard to children because of both hand-to-mouth behavior and intentional ingestion of soil (pica) that contain metals and other contaminants (Hamel et al. 1998). In these communities, cobalt may have been tracked in from outdoors and contaminate carpeting. Cobalt-containing dust may be brought home in the clothing of parents working in industries where they are exposed to cobalt. Children may be exposed to this cobalt while crawling around or playing on contaminated carpeting. Exposure may also result from dermal contact with soil, or by inhaling dust and then swallowing it after mucociliary transport up out of the lungs. Because there is little absorption of cobalt through the skin following dermal exposure, and because much of the cobalt in soil is embedded in or adsorbed to soil particles or insoluble, it may not be in a form accessible for uptake by the body, and therefore may not pose a serious health hazard.

# 6.7 POPULATIONS WITH POTENTIALLY HIGH EXPOSURES

In addition to workers in the hard metal industry (tool production, grinding, etc.) and industries such as coal mining, metal mining, smelting and refining, cobalt dye painters, and the cobalt chemical production, the general population living near these industries may be exposed to high levels of cobalt in air and in soil. Exposure to cobalt during the wet grinding of hard metal tools is especially high when local exhausts are not in use (Sesana et al. 1994). People living near hazardous waste sites may be exposed to cobalt by inhaling dust from contaminated sites or through dermal contact with cobalt-contaminated soil. In the case of children playing in and around unrestricted landfill sites, exposure via dermal and ingestion routes is possible. The general populations in agricultural areas that use sewage sludge or cobalt-containing fertilizers or other soil amendments may be exposed to higher levels of cobalt via inhalation of

dust or dermal contact with the soil. However, no experimental evidence of higher than normal exposures for these population groups was found in the literature. People who live in areas that naturally contain higher levels of cobalt minerals may also be exposed to higher levels of cobalt from both the inhalation and dermal contact routes.

The higher exposure of cobalt in patients with cobalt-chromium knee implants has been demonstrated by the slightly higher levels of cobalt in whole blood, serum, and urine, and by very high levels of cobalt in bone of these patients (IARC 1991; Ostapczuk et al. 1985; Sunderman et al. 1989). Prosthetic devices that contain polyethylene components to avoid metal-to-metal contact do not appear to cause elevated levels of cobalt in tissues and body fluids (IARC 1991; Ostapczuk et al. 1985; Sunderman et al. 1989). People who use cobalt supplements as a treatment for anemia and those who take large amounts of vitamin B-12 as a dietary supplement would have higher intakes of cobalt than the general population.

Workers at nuclear facilities and nuclear waste storage sites may be exposed to potentially high levels of radiation exposure from <sup>60</sup>Co and <sup>58</sup>Co. Workers at irradiation facilities using <sup>60</sup>Co may be exposed to potentially high levels of gamma radiation exposure from this isotope. Patients receiving <sup>60</sup>Co radiotherapy will intentionally be exposed to high levels of gamma radiation.

## 6.8 ADEQUACY OF THE DATABASE

Section 104(i)(5) of CERCLA, as amended, directs the Administrator of ATSDR (in consultation with the Administrator of EPA and agencies and programs of the Public Health Service) to assess whether adequate information on the health effects of cobalt is available. Where adequate information is not available, ATSDR, in conjunction with the National Toxicology Program (NTP), is required to assure the initiation of a program of research designed to determine the health effects (and techniques for developing methods to determine such health effects) of cobalt.

The following categories of possible data needs have been identified by a joint team of scientists from ATSDR, NTP, and EPA. They are defined as substance-specific informational needs that if met would reduce the uncertainties of human health assessment. This definition should not be interpreted to mean that all data needs discussed in this section must be filled. In the future, the identified data needs will be evaluated and prioritized, and a substance-specific research agenda will be proposed.

### 6.8.1 Identification of Data Needs

Physical, Chemical and Radiological Properties. As can be seen from Table 4-2 and Section 4.2, the relevant physical and chemical properties of cobalt and its compounds are sufficiently known to enable prediction of environmental fate and transport of cobalt compounds (Budavari 1996; Lide 1994; Stokinger 1981; Weast 1985). Information on the radiological properties of important cobalt isotopes are also well known (see Table 4-3) (ICRP 1983, Lide 1994).

Production, Import/Export, Use, Release, and Disposal. Information on the production, import/export, use, release, and disposal of a chemical is important because it is an indicator of possible environmental contamination and human exposure. Large releases and consumer use would indicate higher general population exposure from environmental sources (e.g., air, drinking water, and food) and use of consumer products. Occupational exposure may also increase with increased production and use. U.S. production of cobalt is derived primarily from scrap (secondary production). Information is available on cobalt consumption derived from secondary production, import/export, and release of cobalt from the National Defense Stockpile (USGS 1998, 1999). However, production volumes of individual cobalt compounds are not available. Information on the production of individual compounds would be useful in assessing exposure to specific cobalt compounds. Cobalt isotopes, primarily <sup>60</sup>Co and <sup>57</sup>Co, are not commercially produced in the United States, but rather are imported from Canada and the United Kingdom; consumption amounts are not available. Information of the uses of cobalt are available (Cobalt Development Institute 2000; Donaldson 1986; Hodge 1993; IARC 1991; Richardson 1993; USGS 1998). Cobalt-containing products are mostly used in the workplace, although some consumer products contain cobalt.

According to the Emergency Planning and Community Right-to-Know Act of 1986, 42 U.S.C. Section 11023, industries are required to submit chemical release and off-site transfer information to the EPA. The TRI for 1999 became available in May of 2001 (TRI99 2001). Starting in 1998, metal mining, coal mining, electric utilities, and RCRA/solvent recovery industries were required to report to the TRI. These sectors include those contributing greatest environmental releases of cobalt and cobalt compounds, giving us a much more complete picture of cobalt releases to the environment. The TRI also contains information on the on-site and off-site disposal and management of wastes (e.g., recycling, treatment, transfer to POTWs). EPA guidelines address the disposal of hazardous cobalt wastes. The TRI database will be updated yearly and provides a list of industrial production facilities and emissions. The TRI data

should be used with caution since the 1987 data represent first-time reporting by these facilities. Only certain types of facilities were required to report. This is not an exhaustive list.

**Environmental Fate.** There are data that permit assessment of the environmental fate and transport of cobalt in water and soil (Section 6.3). Sediment and soil are the ultimate sinks for cobalt. There is a paucity of data in the literature regarding the chemical forms of cobalt released to the atmosphere and their transformations in air. This information would facilitate the determination of the transport and persistence of cobalt in the atmosphere. Additional data elucidating the mode of speciation of cobalt in water and soil would also be desirable. For example, under what circumstances might Co(III) compounds be formed in the environment and how long would they persist?

Bioavailability from Environmental Media. No information was located regarding absorption of cobalt in humans following dermal exposure. Absorption by the inhalation and oral routes in humans has been studied, but the results vary considerably (see Section 3.3.1.1.) (Foster et al. 1989; Harp and Scoular 1952; Sedlet et al. 1958; Sorbie et al. 1971; Valberg et al. 1969). These variations were attributed to differences in the types and doses of cobalt compounds given, to the nutritional status of the subjects following oral exposure, and to particle size differences following inhalation exposure. Additional data assessing the absorption of cobalt following soil ingestion by children may be helpful. Data in animals are plentiful for both inhalation and oral routes and correlate well with the human data (Andre et al. 1989; Bailey et al. 1989; Collier et al. 1989; Kreyling et al. 1986; Patrick et al. 1989; Talbot and Morgan 1989). Data in animals following dermal exposure suggested that cobalt is not absorbed well through intact skin, but is rapidly taken up through damaged skin. Data regarding the bioavailability of cobalt following dermal exposure is important because dermal exposure to cobalt in the workplace is probable.

**Food Chain Bioaccumulation.** Bioaccumulation in the food chain is important in assessing the human exposure to cobalt from the consumption of food. Data are available that indicate that cobalt is not taken up appreciably by plants and does not biomagnify up the food chain (Baudin and Fritsch 1987; Baudin et al. 1990; Boikat et al. 1985; Francis et al. 1985; Kloke et al. 1984; Lux et al. 1995; Mascanzoni 1989; Mejstrik and Svacha 1988; Mermut et al. 1996; Palko and Yli-Hala 1988; Smith and Carson 1981; Tolle et al. 1983; Watabe et al. 1984).

**Exposure Levels in Environmental Media.** Monitoring data on levels of cobalt in air, water, and food permits the estimation of exposure from these sources. Data are available on the cobalt levels in ambient air (Golomb et al. 1997; Hasanen et al. 1990; Schroeder et al. 1987; Smith and Carson 1981; Sweet et al. 1993; Wiersema et al. 1984). However, the data are not sufficiently recent or broad-based for estimating the current levels of exposure to cobalt in the general U.S. population and particularly those living near cobalt-containing hazardous waste sites. In addition, in only isolated studies was there an assessment of the concentration of cobalt associated with coarse and fine particles (Sweet et al. 1993) or an average annual level obtained at a site (Golomb et al. 1997). Similarly, levels of cobalt in ambient water, while generally low, are also not sufficiently broad-based or recent to be satisfactory (Bargagli 2000; Bruce and McMahon 1996; Cassidy et al. 1982; Eckel and Jacob 1988; Flaten 1991; Nojiri et al. 1985; Rossmann and Barres 1988; Smith and Carson 1981). This deficiency may be satisfied when the EPA's improved and updated STORET database comes on line. Cobalt levels in Canadian drinking water are #2.0 mg/L (Meranger et al. 1981). However, U.S. drinking water levels haven't been reported. The levels of cobalt in sediment are available (Bargagli 2000; Coakley et al. 1993; Gibbs 1994; Glooschenko et al. 1981; Knutson et al. 1987; Naidu et al. 1997; Shine et al. 1995; Smith and Carson 1981; Trocine and Trefry 1996; Villanueva and Botello 1998), but more data on levels in soil and in the vicinity of industrial and hazardous waste sites would be useful. Few data on the levels of cobalt in U.S. foods are available, although studies from Canada and Sweden are available that indicate that cobalt levels in food items are generally low (Barceloux 1999; Dabeka and McKenzie 1995; Jorhem and Sundström 1993). In particular, total diet studies of cobalt in U.S. food is lacking. A Canadian total diet study estimated average daily cobalt intake to range from 7 to 15 µg/day for different age-sex groups (Dabeka and McKenzie 1995).

Few data are available on levels of <sup>60</sup>Co and other cobalt isotopes in environmental media.

Exposure Levels in Humans. The levels of cobalt in hair, nail, and adipose tissues of the general U.S. population are known (EPA 1986; Takagi et al. 1986, 1988). No reliable data on the levels of this substance in blood (or plasma) and urine of the general U.S. population were found, although such data are available for certain European populations including occupationally-exposed groups (Table 6-8). These data may be important for establishing the background exposure level of cobalt. No data on the levels of cobalt in any body tissue or fluid for populations living near hazardous waste sites are available. Such data would be important in assessing the exposure levels of this group of people.

**Exposures of Children.** Dabeka (1989) reported the levels of cobalt in various formulas and milk products consumed by children in Canada, and Dabeka and McKenzie (1995) determined the mean dietary intake of Canadian children as young as 1–4 years of age. Nriagru (1992) reported levels of cobalt in human milk from several countries. No analogous U.S. studies were found. Cobalt levels is the tissue and body fluids of children have not been found.

Child health data needs relating to susceptibility are discussed in Section 3.12.2 Identification of Data Needs: Children's Susceptibility.

**Exposure Registries.** No exposure registries for cobalt were located. This compound is not currently one of the compounds for which a subregistry has been established in the National Exposure Registry. The compound will be considered in the future when chemical selection is made for subregistries to be established. The information that is amassed in the National Exposure Registry facilitates the epidemiological research needed to assess adverse health outcomes that may be related to the exposure to cobalt and its compounds.

# 6.8.2 Ongoing Studies

The Federal Research in Progress (FEDRIP 2000) database provides additional information obtainable from a few ongoing studies that may fill in some of the data needs identified in Section 6.8.1. These studies are summarized in Table 6-9.

Remedial investigations and feasibility studies conducted at the NPL sites known to be contaminated with cobalt will add to the available database on exposure levels in environmental media, exposure levels in humans, and exposure registries, and will increase the current knowledge regarding the transport and transformation of cobalt in the environment. No other long-term research studies pertaining to the environmental fate of cobalt or to occupational or general population exposures to cobalt were identified.

Table 6-9. Ongoing Studies on Cobalt

Investigator	Affiliation	Research description	Sponsor
Bates, GT	Alcorn State University, Agriculture, Lorman, Michigan	To develop profiles of selected trace minerals in the Memphis soil and the forage crops grown on this soil and to compare these to those reported in the literature relative to toxicities or deficiencies. To monitor levels of trace minerals in the hair, tissues, and body fluids of beef cattle fed forages produced on the Memphis soil and to compare these levels to those in the soil and forages.	USDA Cooperative State Research Service
Longnecker, Matthew	NIEHS, NIH	Evaluate the use of toenail levels as a measure of exposure by analyzing toenail and whole-diet homogenates by neutron activation analysis. Toenails reflect exposure over a longer period of time than do blood or urine measures, and are less likely to be influenced by contamination than hair.	NIEHS
Moffett, James F	Woods Hole Oceanographic Institution, Woods Hole, Massachusetts	It is known that trace metals in seawater affect ocean phytoplankton and bacteria differently depending upon their concentrations and chemical forms. In addition, such organisms are capable of secreting complexating agents that change trace metal speciation, making them more or less assimilable and more or less toxic. This phenomena will be studied as a two-way street: water column cobalt affecting oceanic microbial communities and oceanic microbial communities affecting the water column cobalt chemistry. The studies will involve field (Sargasso Sea) and laboratory studies with two species of cyanobacteria and their biogeochemical relationship.	NSF, Division of Ocean Sciences
Odom, JW	Auburn University, Agronomy and Soils Deptartment	The occurrence, measurement, and mapping of plant micronutrients and trace elements, including cobalt, in Alabama soils will be determined including both total and extractable forms of these elements in selected soil profiles.	USDA Cooperative State Research Service
Perry, DL	Lawrence Berkeley Laboratory, University of California	One of the most important mechanisms that affects toxic metal subsurface transport is that of organic complexation. The research will involve first the synthesis of organic-metal ion complexes involving cobalt, uranium, and chromium with the multidentate chelating carboxylic acids diethylenetriaminepentaacetic acid (DTPA), ethylenediaminetetraacetic acid (EDTA), and cyclohexanediaminetetraacetic acid (CDTA). Auger, X-ray photoelectron, FT- IR, and Raman spectroscopies will then be used to study the chemistry and spectroscopy of the mixed contaminants and devise new analytical approaches for the detection of the mixed contaminants in soil and groundwater. The data derived from this research will provide an extremely critical experimental chemical base to model hydrologic transport of cocontaminant organic-metal ion species in subsoils and groundwater.	USDOE Energy Research

Table 6-9. Ongoing Studies on Cobalt (continued)

Investigator	Affiliation	Research description	Sponsor
Stone, AT	Johns Hopkins University, Baltimore, Maryland	This project is concerned with the mobilization of MnO <sub>2</sub> and FeOOH-bound toxic metals in subsurface arising from an influx of naturalorganic matter or organics-containing wastes. Work is reported that examines how reductant, complexant, and adsorptive characteristics of constituent organic chemicals affect the oxidation state and solid-solution partitioning of cobalt, nickel, copper, and lead. In this work, representative low-molecular weight organic compounds and natural organic matter (NOM)-containing subsurface samples are added to hydrous oxide suspensions containing toxic metals such as cobalt, lead, mercury, and uranium. Changes in toxic metal oxidation state, solubility, and speciation are explored in relation to the complexant, reductant, and adsorptive characteristics of the added organic constituents. The goal of this work is to improve the ability to predict toxic metal speciation and the potential for migration at contaminated subsurface sites.	USDOE Energy Research
Tyler, J	University of Missouri, Columbia, Missouri	To survey the prevalence and geographic distribution of selenium, copper, zinc and cobalt deficiency in Missouri feeder calves. Blood will be analyzed.	USDA Cooperative State Research Service
Zachara, JM	Pacific Northwest Laboratory, Richland, Washington	To investigate interfacial geochemical reactions of subsurface materials to chemical mixtures in order to improve reaction-based models of sorption and abiotic degradation used to predict contaminant concentrations in subsurface environments. The scientific focus is on multispecies interactions between organic substances and inorganic contaminants (e.g., metals, radionuclides) such as competitive sorption, cosorption, and cosolvation. Organic substances (humic materials, organic acid complexing agents, and organic solvents) and metals-radionuclides (U, Co, Pb, Cs, Sr, and Tc) representative of chemical constituents on DOE lands are used in carefully selected mixtures to test surface chemical hypotheses about multispecies interactions. Laboratory experimentation involves the contact of these mixtures with (1) subsurface materials having diverse mineral and chemical properties, (2) natural clays, oxides, and organic matter isolated from subsurface sediments, and (3) reference mineral-organic surfaces. Various instrumental techniques employed to investigate the nature of the multispecies surface reactions include laser microelectrophoresis, luminescence spectroscopy, and calorimetry. Multispecies, multisite surface complexation models, are employed to evaluate hypotheses regarding dominant surface reaction mechanisms, reactive contributions of different sorbents, and the effects of different cocontaminant interactions.	USDOE Energy Research

NIEHS = National Institute of Environmental Health Sciences; NIH = National Institute of Health; NSF = National Science Foundation USDA = U.S. Department of Agriculture; USDOE = U.S. Department of Energy

COBALT 253

# 7. ANALYTICAL METHODS

The purpose of this chapter is to describe the analytical methods that are available for detecting, measuring, and/or monitoring cobalt, its metabolites, and other biomarkers of exposure and effect to cobalt. The intent is not to provide an exhaustive list of analytical methods. Rather, the intention is to identify well-established methods that are used as the standard methods of analysis. Many of the analytical methods used for environmental samples are the methods approved by federal agencies and organizations such as EPA and the National Institute for Occupational Safety and Health (NIOSH). Other methods presented in this chapter are those that are approved by groups such as the Association of Official Analytical Chemists (AOAC) and the American Public Health Association (APHA). Additionally, analytical methods are included that modify previously used methods to obtain lower detection limits and/or to improve accuracy and precision.

## 7.1 BIOLOGICAL SAMPLES

Entry of cobalt and its radioisotopes into the human body can be gained through ingestion, inhalation, or penetration through skin. The quantities of cobalt within the body can be assessed through the use of bioassays that are comprised of either *in vivo* and/or *in vitro* measurements. *In vivo* measurements can be obtained through techniques that directly quantitate internally deposited cobalt using, for example, whole body counters. These *in vivo* measurement techniques are commonly used to measure body burdens of cobalt radioisotopes (i.e., <sup>60</sup>Co), but cannot be used to assess the stable isotope of cobalt (<sup>59</sup>Co). Instead, *in vitro* measurements provide an estimate of internally deposited cobalt (both the stable and radioactive isotopes), utilizing techniques that measure cobalt in body fluids, feces, or other human samples.

Examples of these analytical techniques are given in NRCP Report No. 87 (1987) and are also listed in Tables 7-1 and 7-2.

## 7.1.1 Internal Cobalt Measurements

*In vivo* measurement techniques are the most direct and widely used approach for assessing the burden of cobalt radioisotopes within the body. The *in vivo* measurement of these radioisotopes within the body is performed with various radiation detectors and associated electronic devices that are collectively known as whole body counters. These radiation detectors commonly utilize sodium iodide (NaI), hyperpure germanium, and organic liquid scintillation detectors to measure the 1,172 and 1,332 keV gamma rays

Table 7-1. Analytical Methods for Determining Stable Cobalt in Biological Materials

Sample matrix	Preparation method	Analytical method	Sample detection limit	Percent recovery	Reference
Urine	Direct injection	GF-AAS with Zeeman background correction	0.3µg/L	101% at 40μg/L	Bouman et al. 1986
	Addition of magnesium nitrate and nitric acid matrix modifiers and equal volume dilution of sample with water	GF-AAS with Zeeman background correction	2.4 μg/L	107.6% at 50 μg/L	Kimberly et al. 1987
	Sample chelated with dithiocarbamic acid derivative, solvent extracted	GF-AAS with Zeeman background correction	0.1 µg/L	No data	Alexandersson 1988; Ichikawa et al. 1985
	Sample wet digested with acid and chelated with 2,3-butanedion dioxide and complex preconcentrated at hanging mercury drop electrode	DPCSV	0.2 μg/L	No data	Heinrick and Angerer 1984
	Direct injection	GF-AAS with Zeeman background correction	0.1 μg/L	No data	Sunderman et al. 1989
Whole blood	Sample diluted with a homogenizer	GF-AAS with D <sub>2</sub> background correction	2 μg/L	No data	Heinrick and Angerer 1984
	Sample wet digested with acid and chelated with 2,3-butanedion dioxine and complex preconcentrated at hanging mercury drop electrode	DPCSV	0.8 µg/L	No data	Heinrich and Angerer 1984
	Sample acid digested, complexed with thiocyanate and N-phenylcinnamohydroxamic acid and extracted into ethyl acetate	Colorimetric	0.15 mg/L	No data	Afeworki and Chandravanshi 1987

Table 7-1. Analytical Methods for Determining Cobalt in Biological Materials (continued)

Sample matrix	Preparation method	Analytical method	Sample detection limit	Percent recovery	Reference
Serum	Direct injection	GF-AA with Zeeman background correction	0.02 μg/L	No data	Sunderman et al. 1989
Blood or tissue	Acid digestion	ICP-AES (NIOSH method 8005)	10 μg/g (blood); 0.2 μg/g (tissue)	81% at 110 μg/L (blood)	NIOSH 1984

 $D_2$  = deuterium; DPCSV = differential pulse cathodic stripping voltammetry; GF-AAS = graphite furnace atomic absorption spectrometry; ICP-AES = inductively coupled plasma-atomic emission spectrometry; NIOSH = National Institute for Occupational Safety and Health

Table 7-2. Analytical Methods for Determining Radioactive Cobalt in Biological Samples

Sample matrix	Preparation method	Analytical method	Sample detection limit	Percent recovery	Reference
Urine	Direct count of sample	γ-spectrometry with Nal detector	No data ( <mdl)< td=""><td>No data</td><td>Miltenberger et al. 1981</td></mdl)<>	No data	Miltenberger et al. 1981
Soft tissue	Sample wet-ashed	γ-spectrometry (NaI)	No data	No data	Baratta et al. 1969
	Sample directly counted in detector	γ-spectrometry	5 pCi/g	No data	Rabon and Johnson 1973
	Sample digested in acid, oxidized with HClO <sub>4</sub> , concentrated by precipitation with AMP, purified by resin column, precipitated with hexachloroplatinic acid	β-counter	0.1 pCi/g	40-85%	Nevissi 1992
Feces	Direct count of sample	γ-spectrometry	No data	No data	Smith et al. 1972
Blood	Red cells separated from plasma and washed	γ-spectrometry with NaI detector	No data	No data	Smith et al. 1972

AMP = ammonium molybdophosphate; MDL = minimum detectable level; NaI = sodium iodide

from the decay of <sup>60</sup>Co. Because of the relatively low attenuation of the high energy gamma rays emitted from <sup>60</sup>Co by most tissues, cobalt radioisotopes can easily be detected and quantified using whole body counting techniques (Lessard et al. 1984; NCRP 1987; Raghavendran et al. 1978; Smith et al. 1972; Sun et al. 1997). Many configurations of the whole body counter and scanning methods have been utilized, ranging from unshielded single-crystal field detectors to shielded, multi-detector scanning detectors (IAEA 1962, 1970, 1972, 1976, 1985; NCRP 1987). Where appropriate, shielding of the room that houses the whole body counter and/or the detector is often used to increase the detection sensitivity of the equipment by minimizing background radiation. Additionally, care must be exercised to insure that external contamination with radioactive cobalt or other gamma-emitting radioisotopes on the clothing or skin of the individual to be scanned has been removed. Also, *in vitro* measurements of cobalt (see Section 7.1.2) are often used in conjunction with whole body counting when monitoring individuals working with cobalt, especially in conjunction with the assessment of individuals who have experienced accidental exposures to cobalt (Bhat et al. 1973).

Calibration of whole body counters is achieved through the use of tissue-equivalent phantoms. These phantoms are constructed to mimic the shape and density of the anatomical structure using tissue equivalent materials such as water-filled canisters or masonite (Barnaby and Smith 1971; Bhat et al. 1973; Sun et al. 1997). For example, the bottle mannequin absorber (BOMAB) consists of a series of water-filled polyethylene canisters constructed into seated or reclined human forms (Sun et al. 1997). <sup>60</sup>Co standards are measured either as point sources along the phantom or dissolved within the water-filled canisters. Comparisons of the actual counts obtained from the phantom to the known activity of the cobalt standards are used to determine the efficiency of the counting technique and, thus, provide the basis for calibrating the technique. Even so, differences in whole body measurement techniques, calibration methods, and background radiation count calculations between different laboratories can complicate the direct comparisons of body burden measurements and clearance rates for cobalt radioisotopes and should be taken into consideration when comparing data obtained from independent laboratories.

# 7.1.2 External Radiation Measurements

*In vitro* analyses of cobalt are routinely performed in situations where *in vivo* analyses can not be obtained or in support of an *in vivo* monitoring program. Urine and feces are the preferred samples for *in vitro* analyses of cobalt, although other sample types, such as tissue, bone, or blood, can also be used on a

more limited basis. Urine provides for an analysis of soluble (inorganic) cobalt, fecal analysis can be used to assess the cobalt (organic) that is eliminated into the gut or the fraction of ingested cobalt not absorbed by the gut, and tissue/blood/bone are used to assess whole or regional body burdens of cobalt (NCRP 1987; Smith et al. 1972).

The analytical methods for determining the stable cobalt isotope, <sup>59</sup>Co, in biological matrices are given in Table 7-1. For accurate determination of cobalt, contamination of samples during sample collection, storage, and treatment must be avoided, particularly for biological samples containing low levels of cobalt. Cobalt contamination in blood samples has been reported from disposable syringes and technical-grade anticoagulants. Menghini needles, often used for liver biopsy, and mortar, pestles, and grinding devices used for homogeneous mixing may contaminate samples. Other sources of contamination may be collection and storage containers and chemical reagents used for preparing samples. In fact, sample contamination was responsible for erroneous reports in the earlier literature of grossly high levels of cobalt in biological specimens of unexposed persons. Therefore, blanks should always be run with the samples.

The commonly used classical methods for determining cobalt in biological samples are polarographic and colorimetric methods. Details about these methods are given by Saltzman and Keenan (1957). Since these older methods have interference problems and are unsuitable for determining low levels of cobalt in many biological samples, the samples are pretreated before quantification. Precipitation, chelation, chromatography, and ion-exchange are some of the methods used for this purpose. In recent years, the two single-element instrumental techniques most frequently used methods for determining cobalt are graphite furnace atomic absorption spectrometry (GF-AAS) (also called electrothermal atomic absorption spectrometry) and differential pulse anodic stripping voltammetry (DPAVS). Multi-element techniques commonly used for cobalt determination are neutron activation analysis and inductively coupled plasma atomic emission spectrometry. Several other methods are available for determining cobalt in biological samples; these include x-ray fluorescence and Spark source mass spectrometry (Adeloju et al. 1985; Smith and Carson 1981).

For the *in vitro* analysis of cobalt radioisotopes in human samples, the majority of the analytical methods measure the cobalt radioisotopes directly in the samples without the requirement for an extensive sample preparation procedure using gamma spectrometry or scintillation techniques. Of the cobalt radioisotopes that have been detected in the environment (e.g., <sup>57</sup>Co, <sup>58</sup>Co, and <sup>60</sup>Co), <sup>60</sup>Co is the most common.

Consequently, most of the analytical methods that will be described in this chapter are those developed for the detection and quantitation of <sup>60</sup>Co in biological (see Table 7-2) and environmental samples (see Table 7-4).

The radiochemical analysis of <sup>60</sup>Co in urine has been used in conjunction with whole body scanning methods to assess acute and long-term body burdens of this isotope. The analysis of <sup>60</sup>Co in urine is the same as that described for a standardized method of analysis of cesium radioisotopes in urine (Gautier 1983). A urine sample of approximately 2 liters is collected (either over 24 hours or before and after bedtime) and a 1 liter aliquot transferred to a Marinelli beaker for counting in a gamma ray spectrometer (Gautier 1983). This simple procedure offers high recoveries of cobalt (98%) and the minimum detection sensitivity (100 pCi/L) that is required to evaluate individuals for exposures to radioactive cobalt (Gautier 1983). Direct counting methods are also used for the analysis of cobalt radioisotopes in tissues, feces, and blood (Smith et al. 1972, Table 7-2). However, some of these methods may require sample preparation to reduce volume or increase concentration.

Accuracy of *in vivo* and *in vitro* measurements of cobalt is determined through the use of standard, certified solutions or radioactive sources with known concentrations or activities of cobalt. Certified standards for <sup>59</sup>Co can be obtained through a number of commercial sources. The primary source of certified cobalt radioisotope standards is the National Institute of Standards and Technology (NIST). Gamma ray point sources for <sup>60</sup>Co (SRM 4200, 60,000 Bq [1.6 μCi] and SRM 4207, 300,000 Bq [56 μCi]) and standard solutions of <sup>60</sup>Co (SRM 4233, 600,000 Bq/g [16 μCi/g]) are available from NIST. Also, the determination of accuracy of a method often requires standard reference materials (SRMs). Unfortunately, very few biological SRMs are available. An SRM for cobalt in animal muscle is available from the International Atomic Energy Agency (IAEA), Vienna; an SRM for bovine liver (SRM-1577) is available from NIST (formerly the National Bureau of Standards) (Adeloju et al. 1985; Smith and Carson 1981).

## 7.2 ENVIRONMENTAL SAMPLES

There are two common approaches for measuring cobalt in the environment. Cobalt radioisotopes can either be measured directly in the field (*in situ*) using portable survey instruments or samples can be procured from the field and returned to the laboratory for quantitation. However, quantitation of the stable cobalt isotope <sup>59</sup>Co in environmental samples is generally conducted in the laboratory.

#### 7.2.1 Field Measurements of Cobalt

In situ measurement techniques are extremely useful for the rapid characterization of radionuclide contamination in the environment, such as soils, sediments, and vegetation, or when monitoring personnel for exposure to radionuclides. The measurement of gamma-ray-emitting radionuclides, like cobalt, in the environment is conducted with portable survey instruments such as Gieger-Mueller detectors, sodium iodide scintillation detectors, and gamma-ray spectrometers. However, the use of gamma-spectrometers in field survey equipment is preferred for measuring cobalt in the field because of its selectivity and sensitivity. The relatively high energy and penetrability of the gamma-ray that is emitted during the decay of <sup>60</sup>Co provides an advantage for assessing the level of cobalt both on and below the surface using portable field survey instruments such as the gamma-ray spectrometer. These gamma-ray spectrometers are equipped with a high purity germanium detector that is able to selectively and sensitively differentiate the 1,173 and 1,332 keV gamma-rays emitted from <sup>60</sup>Co from the gamma-rays emitted from other radionuclides, for example <sup>40</sup>K or <sup>137</sup>Cs (NRC 1997). Minimum detectable activities (MDAs) of 0.005 Bq/g (0.05 pCi/g) for <sup>60</sup>Co are routinely achieved using p-type germanium gamma spectrometers with 10 minute counting times (NRC 1997). However, counting errors can occur where the simultaneous detection of the 1,173 and 1,332 keV gamma-rays produces a sum peak at 2,505 keV or a count in the continuum between the individual peaks and the sum peak (APHA 1998; NRC 1997). These errors can be minimized by changing the geometry of the detector or the distance of the detector from the source of radioactivity. Computational methods have been derived to aid in determining the concentrations and distributions of 60Co in different soil types and depths (NRC 1997). The concentrations and distributions of <sup>60</sup>Co that have been derived from the computational analysis of the survey data is often verified by laboratory-based analyses of soil samples procured from the survey area.

# 7.2.2 Laboratory Analysis of Environmental Samples

Analytical methods for quantifying cobalt and cobalt radioisotopes in environmental samples (e.g. air, water, soil, and biota) are summarized in Tables 7-3 (<sup>59</sup>Co) and 7-4 (<sup>60</sup>Co). The methods that are commonly used in the analysis of <sup>59</sup>Co are based on instrumental analytical techniques, such as atomic absorption spectrometry (AAS), instrumental neutron activation analysis (INAA), and mass spectrometry (MS). The analysis of <sup>60</sup>Co can be determined either as total mass or total activity, depending on the

 Table 7-3. Analytical Methods for Determining Stable Cobalt in Environmental Samples

Sample matrix	Preparation method	Analytical method	Sample detection limit	Percent recovery	Reference
Air (workplace)	Weighed filter irradiated in a reactor	INAA	0.17 μg/m³	No data	Haddad and Zikovsky 1985
	Sample filter digested by wet acid ashing	Flame-AAS with background correction (NIOSH method 7027)	0.4 μg/m³	98% with 12–96 μg spiked filter	NIOSH 1984
	Sample filter digested by wet acid ashing	ICP-AES (NIOSH method 7300)	0.5 μg/m³	95–100% with 2.5–1,000 spiked filter	NIOSH 1984
Water (low ionic strength)	Direct injection	GF-AAS with Zeeman or deuterium back-ground correction	<0.5 μg/L	93–115% at 8.5–30 μg/L	Fishman et al. 1986
Lake water	Sample complexed with 8-hydroxyquinoline absorbed on a column, desorbed and digested with acid	ICP-AES	<0.004 μg/L	No data	Nojiri et al. 1985
Rainwater	Sample preconcentrated onto polystyrene films by spray-drying	PIXE	0.08 μg/L	No data	Hansson et al. 1988
Seawater	Sample complexed with 8-hydroxyquinoline absorbed on a column, desorbed and digested with acid	GF-AAS with Zeeman background correction	0.0002 μg/L	90%	Nakashima et al. 1988
Water and waste water	Direct aspiration of sample	Flame-AAS (EPA method 219.1)	0.05 mg/L	97–98% at 0.2–5.0 mg/L	EPA 1983
	Direct injection	GF-AAS with back- ground correction (EPA method 219.2)	1 μg/L	No data	EPA 1983

7. ANALYTICAL METHODS

Table 7-3. Analytical Methods for Determining Stable Cobalt in Environmental Samples (continued)

Sample matrix	Preparation method	Analytical method	Sample detection limit	Percent recovery	Reference
Groundwater or leachate	Direct aspiration	Flame-AAS with background correction (EPA method 7200)	0.05 mg/L	97–98% at 0.2–5.0 mg/L	EPA 1986b
Groundwater or leachate	Direct injection	GF-AAS with background correction (EPA method 7201)	1 μg/L	No data	EPA 1986b
Food	Sample digested with acid	GF-AAS with background correction	1.88 µg/L in dissolved extract	100–107% at 0.2–0.6 mg/kg (leaves, liver)	Barbera and Farre 1988

AAS = atomic absorption spectrometry; EPA = Environmental Protection Agency; GF-AAS= graphite furnace atomic absorption spectrometry; ICP-AES = inductively coupled plasma-atomic emission spectrometry; INAA= instrumental neutron activation analysis; NIOSH = National Institute for Occupational Safety and Health; PIXE = photon induced x-ray emission

Table 7-4. Analytical Methods for Determining Radioactive Cobalt in Environmental Samples

Sample matrix	Preparation method	Analytical method	Sample detection limit	Percent recovery	Reference
Air	Direct count of sample collected on paper filter	γ-spectrometry with Ge/Li detector	0.001 pCi/m <sup>3</sup>	No data	USAEC 1974a
Air	Sample filter ashed	Scintillation counter with Nal detector	No data	No data	De Franceschi et al. 1974
Drinking water	Direct count of sample	γ-spectrometry with Ge detector	<2 pCi/L	99%	APHA 1998
Drinking water	Direct count of sample	γ-spectrometry	2 pCi/L	No data	USAEC 1974b
Water	Direct count of sample	γ-spectrometry with Ge/Li detector	2 pCi/L	No data	ASTM 1999
Water	Direct count of sample	γ-spectrometry	10 pCi/L	No data	Cahill et al. 1972
Seawater	Sample concentrated using continuous-flow coprecipitation-flotation separation technique	Scintillation detector	50 fCi/L	92–95%	Hiraide et al. 1984
Sediments	Sample dried and ground	γ-spectrometry	0.04 pCi/g	No data	Cahill et al. 1972
Fish	Samples dried and ashed	γ-spectrometry	0.001 pCi/g (DW)	No data	Cushing et al. 1981
Mollusc	Samples dried and ashed	γ-spectrometry	<0.01 pCi/g	No data	De Franceschi et al. 1976

DW = dry weight; Ge/Li = lithium drifted geranium; NaI = sodium iodide

analytical technique that is used. Typically, radiochemical methods of analysis employing gamma spectrometry techniques are used to quantitate <sup>60</sup>Co in environmental samples.

Analytical methods for determining cobalt in environmental samples are given in Table 7-3. Since cobalt exists in the particulate form in the atmosphere, it is sampled by pumping air through a metal-free filter (usually cellulose ester membrane), and the metal is quantified in the collected particles. Sample treatment prior to quantification is important for environmental samples. For example, the use of sodium carbonate for dry ashing plant materials results in poor cobalt recovery. Low-temperature ashing may be inadequate for some samples, and losses may occur during rigorous dry ashing. Wet ashing is the preferred method when sample treatment is necessary. Wet extraction with dilute nitric acid is most suitable for analyzing cobalt in dust samples. In some samples, the determination of soluble and insoluble cobalt is important, and analytical methods used to determine cobalt in filtered and unfiltered samples are available for this purpose.

As in the case of biological samples, contamination of environmental samples during sample collection, storage, and treatment should be avoided. Loss of cobalt from aqueous samples due to adsorption on storage containers should be avoided by using polyethylene or similar containers and acidifying the solution to the proper pH (Smith and Carson 1981). Because of its rapidity, accuracy, and low detection limit, graphite furnace atomic absorption spectrometry with Zeeman background correction is the most commonly used method for quantifying cobalt in environmental samples. To meet the detection limits of the available analytical methods, preconcentration prior to quantification may be necessary for some samples (e.g., seawater). A few commonly used methods for determining cobalt in environmental samples are given in Table 7-3. Other less frequently used methods are inductively coupled plasma-mass spectrometry (ICP-MS) (Henshaw et al. 1989; McLaren et al. 1985), gas, liquid, and ion chromatography with colorimetric, electron capture, and electrochemical detection (Bond and Wallace 1984; Carvajal and Zienius 1986; Cheam and Li 1988; King and Fritz 1987; Schaller and Neeb 1987), photoacoustic spectroscopy with colorimetry (Kitamori et al. 1986), electrothermal vaporization with ICP-AES (Malinski et al. 1988); and chemiluminescence with spectrofluorimetry (Jones et al. 1989).

Analytical methods for determining cobalt radioisotopes in the environment are shown in Table 7-4. The analysis of cobalt in air is based on quantifying cobalt within aerosols or particles that become trapped on cellulose (paper) or glass fiber filters after a calibrated amount of air is passed through the filters. Since the cobalt radioisotopes do not occur naturally but are formed as a result of nuclear weapons testing

(which has been discontinued for several years), neutron-activation of specific materials, or reactor core damage, the amount of these isotopes within the ambient environment are near or below the minimum detectable levels for these isotopes (DOE 1995). However, trace amounts of <sup>60</sup>Co can be detected in air, water, and sediments within or near nuclear weapons or fuel production facilities, nuclear reactors, and nuclear waste storage sites (DOE 1995; Boccolini et al. 1976; USAEC 1973). Analysis of cobalt radioisotopes in air filters, water, sediments, vegetation, and biota can be performed directly using gamma spectormetry, or following some sample preparation (e.g., drying, ashing, or extraction) (Boccolini et al. 1976; Cahill et al. 1972; Cushing 1981; Hiraid et al. 1984; Windham and Phillips 1973).

The detection limits, accuracy, and precision of any analytical methodology are important parameters in determining the appropriateness of a method for quantifying a specific analyte at the desired level of sensitivity within a particular matrix. The Lower Limit of Detection (LLD) has been adopted to refer to the intrinsic detection capability of a measurement procedure (sampling through data reduction and reporting) to aid in determining which method is best suited for the required sample quantitation (NRC 1984). Several factors influence the LLD, including background, size or concentration of sample, detector sensitivity, recovery of desired analyte during sample isolation and purification, level of interfering contaminants, and, particularly, counting time. Because of these variables, the LLDs between laboratories, utilizing the same or similar measurement procedures, will vary.

The accuracy of a measurement technique in determining the quantity of a particular analyte in environmental samples is greatly dependent on the availability of standard reference materials. Several SRMs for cobalt in environmental samples are also available. Some of these are coal, fly ash, diet, and orchard leaf SRMs available from NIST. The Community Bureau of Reference, European Communities offers SRMs for cobalt in sludges, and an SRM for cobalt in thin polymer films is available from NIST for x-ray fluorescence analysis in aerosol particle samples (Dzubay et al. 1988; Miller-Ihli and Wolf 1986; Schramel 1989; Smith and Carson 1981; Tinsley et al. 1983). Gamma ray point sources for  $^{60}$ Co (SRM 4200, 60,000 Bq [1.6  $\mu$ Ci] and SRM 4207, 300,000 Bq [56  $\mu$ Ci]) and standard solutions of  $^{60}$ Co (SRM 4233, 600,000 Bq/g [16  $\mu$ Ci/g]) are available from NIST.

## 7.3 ADEQUACY OF THE DATABASE

Section 104(i)(5) of CERCLA, as amended, directs the Administrator of ATSDR (in consultation with the Administrator of EPA and agencies and programs of the Public Health Service) to assess whether adequate information on the health effects of cobalt is available. Where adequate information is not available, ATSDR, in conjunction with the National Toxicology Program (NTP), is required to assure the initiation of a program of research designed to determine the health effects (and techniques for developing methods to determine such health effects) of cobalt.

The following categories of possible data needs have been identified by a joint team of scientists from ATSDR, NTP, and EPA. They are defined as substance-specific informational needs that if met would reduce the uncertainties of human health assessment. This definition should not be interpreted to mean that all data needs discussed in this section must be filled. In the future, the identified data needs will be evaluated and prioritized, and a substance-specific research agenda will be proposed.

## 7.3.1 Identification of Data Needs

Methods for Determining Biomarkers of Exposure and Effect. Cobalt concentrations in blood or urine can serve as exposure indicator (Alexandersson 1988; Ichikawa et al. 1985; Scansetti et al. 1985). The available analytical methods are capable of determining the levels of cobalt in both the blood and urine of normal and occupationally exposed persons (Table 7-1). For the quantitation of cobalt radioisotopes, whole body counters can be used to assess radiocobalt body burdens that have occurred both from acute and chronic exposures to cobalt radioisotopes (Bhat et al. 1973; NCRP 1987). *In vitro* analytical methods are available for analyzing cobalt radioisotopes in urine, feces, and tissues obtained from normal and occupational exposed persons (Table 7-2).

Sensitive serum protein responses were found in animals exposed to cobalt at levels below those that produce hematopoietic effects. This unique serum protein response to cobalt exposure includes an increase in alpha globulin fractions of serum proteins and associated serum neuraminic acid. Details of this effect are given in Chapter 2. If similar changes occur in humans, this measurement may provide the earliest indications of effects of cobalt exposure. The available analytical methods are capable of determining these effects of cobalt exposure.

#### Methods for Determining Parent Compounds and Degradation Products in Environmental

**Media.** Analytical methods with good sensitivity and specificity are available for determining cobalt in air, water, soil, and other environmental media (Table 7-3). Analytical methods for cobalt, like most metals, measure total metal content rather than the particular compound. Therefore, analytical methods do not generally differentiate between the parent compound and a transformation product as would be the case, for example, were cobalt oxide to be converted to cobalt sulfate. (An exception to this would be the case of radioactive decay in which the parent could be readily distinguished from the decay product.) Analytical methods with the capability of distinguishing between different cobalt species would be important an important tool for assessing the fate of cobalt compounds in the environment. However, methods for quantifying specific cobalt compounds were not found in the literature.

The levels of the parent compound or its reaction products in different environmental media can be used to assess the exposure to cobalt by humans through the inhalation of air and ingestion of food and drinking water. In the case of cobalt, a correlation between its levels in environmental media (e.g., occupational air) and biological tissues and body fluids has been found (Alexandersson 1988; Ichikawa et al. 1985; Scansetti et al. 1985). Therefore, it is possible to estimate the total body burden of cobalt in workers exposed to airborne cobalt vapor and fumes from its concentration in workplace air.

For cobalt radioisotopes, analytical methods also exist that have good sensitivity and specificity for determining radiocobalt in air, water, soil, and other environmental media are available (Table 7-4). Because <sup>60</sup>Co decays to the stable element <sup>60</sup>Ni, there is no need to develop methods to detect and quantify the decay products.

#### 7.3.2 Ongoing Studies

Ongoing studies concerning analytical methods for the determination of total cobalt, cobalt compounds, and radiocobalt are contained in Table 7-5. The information in this table was found as a result of a search of Federal Research in Progress (FEDRIP 2000).

#### 7. ANALYTICAL METHODS

Table 7-5. Ongoing Analytical Studies on Cobalt

Investigator	Affiliation	Research description	Sponsor
Hyatt, DE	ADA Technologies, Inc., Environmental Sciences	This project describes the development of a novel continuous emissions monitor using plasma emission spectroscopy for direct simultaneous monitoring of part per billion levels of several toxic metals in flue gas.	USDOE Energy Research
Miller-Ihli NJ	Agricultural Research Service, Beltsville, Maryland	Develop single and multielement methods for the determination of trace elements of nutritional and health concern. Develop new/improved methods permitting direct analysis of solids by graphite furnace atomic absorption spectrometry and electrothermal vaporization inductively coupled plasma mass spectrometry (ICP-MS). Develop methods for determination of different chemical forms of these elements by couplinecapillary zone electrophoresis with ICP-MS.	USDA Agricultural Research Service
Wright, AE	Radiological Physics And Engineering Co., Nassau Bay, Texas	The objective of this effort is to develop and market a mailable thermoluminescent dosimeter (TLD) device and readout service suitable for frequent, periodic monitoring of radiation calibrations at medical institutions treating cancer patients with cobalt-60 units and electrical linear accelerators, and having inadequate physics support. The specific aims are (1) to utilize current scientific knowledge and the 10 year experience of the NCI-funded centers for radiological physics in the use of mailed TLDs to develop design criteria for a prototype system; i.e., a positioning jig/miniphantom with self-contained TLD inserts, and (2) to remedy those aspects of currently used TLDs which hinder commercial application; specifically, labor-intensive individual assembly and readout, and difficulty in proper placement when used by personnel other than trained medical physicists.	DHHS

DHHS = Department of Health and Human Services; ICP-MS = inductively coupled plasma-mass spectrometry; NCI = National Cancer Institute; TLD = thermoluminescent dosimeter; USDA = U.S. Department of Agriculture; USDOE = U.S. Department of Energy

COBALT 269

#### 8. REGULATIONS AND ADVISORIES

International and national guidelines and state regulations regarding exposure to stable cobalt and its compounds are summarized in Table 8-1. The regulations regarding radioactive cobalt are summarized in Table 8-2.

Stable Cobalt. An MRL of 1x10<sup>-4</sup> mg cobalt/m³ has been derived for chronic-duration inhalation exposure. The MRL is based on a NOAEL of 0.0053 mg cobalt/m³ for decreased respiratory function in exposed workers (Nemery et al. 1992). An MRL of 1x10<sup>-2</sup> mg/kg-day has been derived for intermediate-duration oral exposure, based on a LOAEL of 1 mg/kg-day for polycythemia in human volunteers (Davis and Fields 1958). No other inhalation or oral MRLs were derived.

The EPA has not derived an RfC or RfD for cobalt and compounds. Similarly, no cancer classification has been performed by the EPA (IRIS 2000). The American Conference of Governmental Industrial Hygienists (ACGIH) has given cobalt a classification of A3, Confirmed Animal Carcinogen with Unknown Relevance to Humans, and established an 8-hour time-weighted average (TWA) of 0.02 mg/m³ for occupational exposure (ACGIH 1999). The Occupational Safety and Health Administration (OSHA) has promulgated an 8-hour Permissible Exposure Limit (PEL) of 0.1 mg/m³ (OSHA 1993), and the National Institute for Occupational Safety and Health (NIOSH) recommends an 8-hour TWA of 0.05 mg/m³ (NIOSH 2000). IARC (1991) reports that cobalt and cobalt compounds are possibly carcinogenic to humans (group 2B), based on sufficient evidence for cobalt metal and cobalt oxides and limited evidence for cobalt chloride and cobalt sulfate.

Cobalt and its compounds are regulated by the Clean Water Effluent Guidelines for the following industrial point sources: nonferrous metal manufacturing, asbestos, timber products processing, paving and roofing, paint formulating, ink formulating, gum and wood, carbon black, and battery manufacturing (EPA 1988).

*Radioactive Cobalt.* No MRLs were derived for inhalation or oral exposure to radioactive cobalt. MRLs for acute and chronic exposure to ionizing radiation exist (ATSDR 1999) and are applicable to cobalt. The EPA has not derived an RfC or RfD for radioactive cobalt (IRIS 2000). Slope factors have been derived for exposure to cobalt radioisotopes (EPA 1997). The slope factors for <sup>60</sup>Co are 1.89x10<sup>-11</sup>/pCi for ingestion, 6.88x10<sup>-11</sup>/pCi for inhalation exposure, and 9.76x10<sup>-6</sup>/year/pCi/g soil for external exposure.

# COBALT 270 8. REGULATIONS AND ADVISORIES

The slope factors for  $^{58}$ Co are  $2.82 \times 10^{-12}$ /pCi for ingestion,  $5.17 \times 10^{-12}$ /pCi for inhalation exposure, and  $3.73 \times 10^{-6}$ /year/pCi/g soil for external exposure, and the slope factors for  $^{58}$ mCo are  $9.46 \times 10^{-14}$ /pCi for ingestion,  $8.90 \times 10^{-14}$ /pCi for inhalation exposure, and  $3.21 \times 10^{-11}$ /year/pCi/g soil for external exposure. For  $^{57}$ Co, the slope factors are  $9.71 \times 10^{-13}$ /pCi for ingestion,  $2.88 \times 10^{-12}$ /pCi for inhalation exposure, and  $2.07 \times 10^{-7}$ /year/pCi/g soil for external exposure.

Table 8-1. Regulations and Guidelines Applicable to Stable Cobalt

Agency	Description	Information	Reference
INTERNATIONAL Guidelines:			
IARC	Carcinogenicity classification Cobalt and cobalt compounds <sup>a</sup>	Group 2B <sup>b</sup>	IARC 2001b
NATIONAL Regulations and Guidelines:			
a. Air			
ACGIH	TLV-TWA Cobalt, elemental, and inorganic compounds (as Co)	0.02 mg/m <sup>3</sup>	ACGIH 2000
NIOSH	REL (TWA) Cobalt metal, dust, and fumes (as Co)	0.05 mg/m <sup>3</sup>	NIOSH 2001
	IDLH Cobalt metal, dust, and fumes (as Co)	20 mg/m <sup>3</sup>	
OSHA	PEL (8-hour TWA) General industry Cobalt metal, dust, and fumes (as Co)	0.1 mg/m³	OSHA 2001e 29CFR1910.1000 Table Z
	PEL (8-hour TWA) Construction industry Cobalt metal, dust, and fumes (as Co)	0.1 mg/m³	OSHA 2001d 29CFR1926.55
	PEL (8-hour TWA) Shipyard industry Cobalt metal, dust, and fumes (as Co)	0.1 mg/m³	OSHA 2001c 29CFR1915.1000
USC	HAP Cobalt compounds		USC 2001a 42USC7412
b. Water			
EPA	NPDES permit application testing requirements —conventional and nonconventional pollutants required to be tested by existing dischargers if expected to be present		EPA 2001g 40CFR122 Appendix D Table IV

Table 8-1. Regulations and Guidelines Applicable to Stable Cobalt (continued)

Agency	Description	Information	Reference
NATIONAL (cont.)	Восоприон	mormaton	11010101100
EPA	BPT effluent limitations Maximum for 1 day Average of daily values for 30 consecutive days	3x10 <sup>-4</sup> kg/kkg 1.2x10 <sup>-4</sup> kg/kkg	EPA 2001b 40CFR415.652
	Groundwater monitoring	Suggested         method       PQL         6010       70 μg/L         7200       500 μg/L         7201       10 μg/L	EPA 2001d 40CFR264 Appendix IX
c. Food			
FDA	Drug products withdrawn or removed from the market for reasons of safety or effectiveness	All drug products containing cobalt salts (except radioactive forms of cobalt and its salts and cobalamin and its derivatives)	FDA 2000a 21CFR216.24
	New drug status accorded through rulemaking procedures	Cobalt preparations intended for use by man	FDA 2000b 21CFR310.502 (a)(7)
	Over-the-counter drugs; recommended warning and caution statement Cobalt as a cobalt salt	Required on articles containing \$0.5 µg per dose and \$2 µg per 24-hour period	FDA 2000e 21CFR369.20
	Substances generally recognized as safe—trace minerals added to animal feeds	Cobalt acetate Cobalt carbonate Cobalt chloride Cobalt oxide Cobalt sulfate	FDA 2000f 21CFR582.20
	Substances prohibited from use	Cobaltous salts and its	FDA 2000g
d. Other	in human food	derivatives	21CFR189.120
ACGIH	Carcinogenicity classification		ACGIH 2000
7.00	Cobalt, elemental, and inorganic compounds (as Co)	A3°	7.00.11.2000
	BEI Cobalt in urine—end of shift	15 μg/L	
	at end of workweek Cobalt in blood—end of shift at end of workweek	1 μg/L	

Table 8-1. Regulations and Guidelines Applicable to Stable Cobalt (continued)

Agency	Description	Information		Reference
NATIONAL (cont.)				
EPA	RfC RfD Carcinogenicity classification	No data		IRIS 2000
	Toxic chemical release reporting; Community Right-to-Know—effective date	01/01/87		EPA 2001c 40CFR372.65(a)
	Hazardous waste—identification and listing	Contain #1 pp synthesis gas generated fro hazardous wa	fuel m	EPA 2001e 40CFR261.38 (b)(5)
	TSCA—health and safety data reporting			EPA 2001j 40CFR716.120
	Municipal solid waste landfills —hazardous constituent for detection monitoring	Suggested method 6010 7200 7201	<u>PQL</u> 70 μg/L 500 μg/L 10 μg/L	EPA 2001f 40CFR258 Appendix I and II
	Reportable quantity Cobalt compounds	1 pound		EPA 2001h 40CFR302.4
USC	Superfund imposition of tax on cobalt	\$4.45 per ton		USC 2001c 26USC4661
	Exemption of tax imposed on recycled cobalt			USC 2001b 26USC4662
<u>STATE</u>				
a. Air				
Alabama	HAP Cobalt compounds			BNA 2001
Alaska	Air contaminant standard TWA Cobalt metal, dust, and fumes	0.05 mg/m <sup>3</sup>		BNA 2001
California	Airborne contaminant Cobalt metal, dust, and fumes			BNA 2001
	HAP Cobalt compounds			BNA 2001
	Toxic air contaminant Cobalt compounds			CA Air Resources Board 2000

Table 8-1. Regulations and Guidelines Applicable to Stable Cobalt (continued)

Agency	Description	Information	Reference
STATE (cont.)			
Colorado	HAP Cobalt metal, dust, and fumes		BNA 2001
	"High-concern" pollutant Cobalt (and compounds)		BNA 2001
	Reportable pollutants Cobalt metal, dust, and fumes		CO Dept. of Public Health and Environment 2000
Connecticut	HAP—hazard limiting value Cobalt metal, dust, and fumes		BNA 2001
	8 hours 30 minutes	2 μg/m³ 10 μg/m³	
Delaware	Reportable quantities Cobalt carbonyl Cobaltous sulfamate Cobalt, ((2,2'-(ethane diylbis(nitrilomethylidyne)	1 pound 1,000 pounds 1 pound	DE Air Quality Management 2000
Hawaii	Air contaminant limit PEL-TWA Cobalt metal, dust, and fumes	0.05 mg/m <sup>3</sup>	BNA 2001
	HAP Cobalt compounds		BNA 2001
Idaho	TAP non-carcinogenic increments Cobalt carbonyl and cobalt hydrocarbonyl (as Co) OEL EL AAC (24-hour average)	1x10 <sup>-1</sup> mg/m <sup>3</sup> 7x10 <sup>-3</sup> pounds/hour 5x10 <sup>-3</sup> mg/m <sup>3</sup>	ID Dept. of Environmental Quality 2000
	Cobalt metal, dust, and fumes OEL EL AAC (24-hour average)	5x10 <sup>-2</sup> mg/m <sup>3</sup> 3.3x10 <sup>-3</sup> pounds/hour 2.5x10 <sup>-3</sup> mg/m <sup>3</sup>	
Illinois	Toxic air contaminant Cobalt		IL EPA 2000a
Kansas	HAP Cobalt compounds		KS Dept. of Health and Environment 2000
Kentucky	HAP Cobalt compounds		BNA 2001

Table 8-1. Regulations and Guidelines Applicable to Stable Cobalt (continued)

Agency	Description	Information	Reference
STATE (cont.)			
Louisiana	Toxic air pollutant Cobalt compounds		BNA 2001
Maine	Emissions standards	2,000 pounds	BNA 2001
Maryland	Toxic air pollutant Cobalt compounds		BNA 2001
Michigan	High concern toxic air pollutants Cobalt compounds		BNA 2001
Minnesota	HAP threshold Cobalt metal and cobalt carbonyl	0.1 tons/year	BNA 2001
Missouri	HAP Cobalt compounds		BNA 2001
Montana	Occupational air contaminant Cobalt metal, dust, and fumes	0.1 mg/m <sup>3</sup>	BNA 2001
Nebraska	HAP Cobalt compounds and cobalt		BNA 2001
New Mexico	Toxic air pollutant Cobalt metal, dust, and fumes (as Co) OEL Emissions	1x10 <sup>-1</sup> mg/m <sup>3</sup> 6.67x10 <sup>-3</sup> pounds/hour	BNA 2001
New York	Annual guideline concentrations	5x10 <sup>-3</sup> μg/m <sup>3</sup>	NYS Dept. of Environmental Conservation 2000
	Dangerous air contaminants TLV Cobolt motel, dust, and	0.1 mg/m <sup>3</sup>	BNA 2001
	Cobalt metal, dust, and fumes	o. i ilig/ili	
	HAP Cobalt compounds		BNA 2001
	Transition limits PEL		BNA 2001
	Cobalt metal, dust, and fumes Final rule limits	0.1 mg/m <sup>3</sup>	
	TWA Cobalt metal, dust, and fumes	0.05 mg/m <sup>3</sup>	

Table 8-1. Regulations and Guidelines Applicable to Stable Cobalt (continued)

Agency	Description	Information		Reference
STATE (cont.)				
North Carolina	PEL-TWA Cobalt metal, dust, and fumes	0.05 mg/m <sup>3</sup>		BNA 2001
Ohio	TRI			Ohio EPA 2000
Oregon	Air contaminant Cobalt metal, dust, and fumes	0.1 mg/m <sup>3</sup>		BNA 2001
Rhode Island	HAP Cobalt compounds			BNA 2001
South Carolina	Toxic air emissions—MAC Cobalt compounds	0.25 μg/m³		BNA 2001
Texas	HAP Cobalt metal, dust, and fumes	0.1 mg/m <sup>3</sup>		BNA 2001
Vermont	HAP Cobalt compounds			BNA 2001
	Hazardous ambient air standards Cobalt compounds Annual average Averaging time Action level	0.12 µg/m³ 24 hours 6.2x10 <sup>-3</sup> poun	ds/8 hours	BNA 2001
Washington	Class B TAP and ASIL (24-hour average) Cobalt metal, dust and fumes Cobalt carbonyl and cobalt hydrocarbonyl	0.17 μg/m³ 0.33 μg/m³		WA Dept. of Ecology 2000
	Thresholds for HAPs Cobalt carbonyl Cobalt metal, dust, and fumes	0.1 tons/year 0.1 tons/year		BNA 2001
Wisconsin	HAP—existing sources AAC <25 feet AAC \$25 feet	4.08x10 <sup>-3</sup> pounds/hour 1.704x10 <sup>-2</sup> pounds/hour		WI Dept. of Natural Resources 1999
b. Water				
Alabama	Groundwater monitoring Cobalt	Suggested methods 6010 7200 7201	<u>PQL</u> 70 μg/L 500 μg/L 10 μg/L	BNA 2001
Arizona	Drinking water guideline	0.70 μg/L		FSTRAC 1999

Table 8-1. Regulations and Guidelines Applicable to Stable Cobalt (continued)

Agency	Description	Information		Reference
STATE (cont.)				
Arkansas	Groundwater monitoring Cobalt	Suggested methods 6010 7200 7201	<u>PQL</u> 70 μg/L 500 μg/L 10 μg/L	BNA 2001
California	Chemicals known to cause cancer or reproductive toxicity—date of initial appearance on the list  Cobalt metal powder  Cobalt[II] oxide  Cobalt sulfate hepta-hydrate	07/01/92 07/01/92 06/02/00		CA EPA 2000
Colorado	Groundwater standard Cobalt	0.05 mg/L		BNA 2001
Delaware	Groundwater monitoring Cobalt	Suggested methods 6010 7200 7201	<u>PQL</u> 70 μg/L 500 μg/L 10 μg/L	BNA 2001
Illinois	Groundwater quality standards for Class II	1 mg/L		IL EPA 2000b
Kentucky	Hazardous waste constituent for groundwater monitoring Cobalt			BNA 2001
Louisiana	Groundwater monitoring Cobalt	Suggested methods 6010 7200 7201	<u>PQL</u> 70 μg/L 500 μg/L 10 μg/L	BNA 2001
Massachusetts	Groundwater monitoring Cobalt	Suggested methods 6010 7200 7201	<u>PQL</u> 70 μg/L 500 μg/L 10 μg/L	BNA 2001
Minnesota	Drinking water guideline	2 μg/L		FSTRAC 1995
	Groundwater protection hazardous constituent Cobalt (total)			BNA 2001
Missouri	Water quality standards Livestock, wildlife watering Groundwater	1x10³ μg/L 1x10³ μg/L		BNA 2001

Agency Description Information Reference STATE (cont.) BNA 2001 Standards for groundwater of New Mexico 10,000 mg/L TDS concentration or less Cobalt 0.05 mg/L New York Groundwater monitoring Suggested BNA 2001 Cobalt methods PQL 6010 70 μg/L 7200 500 µg/L 10 μg/L 7201 Tennessee Effluent limitations—daily BNA 2001 maximum concentration 10 mg/L Cobalt Wisconsin Drinking water guideline 40 µg/L **FSTRAC 1999** Groundwater standards BNA 2001 Cobalt 40 µg/L Enforcement standard Preventive action limit 8 µg/L c. Food No data d. Other Alabama Detection limit values for BNA 2001 comparable fuel specification Cobalt Concentration limit 4.6 mg/kg at 10,000 BTU/pound Arizona Soil remediation levels BNA 2001 Cobalt 4.6x10<sup>3</sup> mg/kg Residential 9.7x104 mg/kg Non-residential Arkansas Detection limit values for BNA 2001 comparable fuel specification Cobalt Concentration limit 4.6 mg/kg at 10,000 BTU/pound BNA 2001 Solid waste management Suggested Cobalt methods PQL 6010 70 μg/L 500 µg/L 7200 7201 10 μg/L

	Description	lufo was ations	Deference
Agency	Description	Information	Reference
STATE (cont.)			
California	Characteristics of toxicity Cobalt and cobalt compounds STLC TTLC	80 mg/L 8,000 mg/kg (wet- weight)	BNA 2001
	Chemicals known to cause cancer or reproductive toxicity Cobalt metal powder Initial appearance on the list	07/01/92	BNA 2001
	Hazardous substance Cobalt, cobalt carbonyl, and cobalt hydrocarbonyl		BNA 2001
Delaware	Detection limit values for comparable fuel specification Cobalt Concentration limit	4.6 mg/kg at 10,000 BTU/pound	BNA 2001
Florida	Toxic substance in the workplace Cobalt metal, dust, and fumes		BNA 2001
Georgia	Soil concentration Cobalt	20 mg/kg	BNA 2001
Illinois	Analytical parameters and required quantitation limits Cobalt Water Soil Method	50 μg/L 10 mg/kg 6010A	BNA 2001
Indiana	Constituent subject to assessment monitoring Cobalt (total and dissolved)		BNA 2001
Maine	Screening standards for beneficial use Cobalt	5,875 mg/kg (dry weight)	BNA 2001
Michigan	Identification and listing of hazardous waste Cobalt	When in the form of 100 microns or less	BNA 2001

Table 8-1. Regulations and Guidelines Applicable to Stable Cobalt (continued)

Agency	Description	Information	Reference
STATE (cont.)			
Minnesota	Hazardous substance Cobalt metal, dust, and fumes (as Co) Cobalt carbonyl (as Co) Cobalt, elemental and inorganic compounds (as Co) Cobalt hydrocarbonyl (as Co)		BNA 2001
Missouri	Hazardous constituent Cobalt (total)		BNA 2001
New Jersey	Hazardous substance Cobalt Cobalt carbonyl Cobalt compounds		BNA 2001
New York	Occupational lung disease Hard metal disease Cobalt		BNA 2001
Ohio	Toxic release inventory		BNA 2001
Oklahoma	Fertilizer labels and labeling; minimum percentage accepted for registration Cobalt	5x10 <sup>-4</sup> percent	BNA 2001
Oregon	Toxic substance Cobalt		BNA 2001
Pennsylvania	Hazardous substance Cobalt and cobalt fumes		BNA 2001

<sup>&</sup>lt;sup>a</sup>Cobalt compounds: includes cobalt(II) carbonate, cobalt(II) chloride, cobalt(II) nitrate, cobalt(II) oxide, cobalt(III) oxide, cobalt(III) oxide, and cobalt(II) sulfate

AAC = acceptable ambient concentrations; ACGIH = American Conference of Governmental Industrial Hygienists; ASIL = acceptable source impact level; BEI = biological exposure indices; BNA = Bureau of National Affairs; BPT = best practicable control technology; BTU = British thermal unit; CFR = Code of Federal Regulations; EL = emissions levels; EPA = Environmental Protection Agency; FDA = Food and Drug Administration; FSTRAC = Federal-State Toxicology and Risk Analysis Committee; HAP = hazardous air pollutant; IARC = International Agency for Research on Cancer; IDLH = immediately dangerous to life and health; IRIS = Integrated Risk Information System; MAC = maximum allowable concentration; NIOSH = National Institute for Occupational Safety and Health; NPDES = National Pollutant Discharge Elimination System; OEL = occupational exposure limit; OSHA = Occupational Safety and Health Administration; PEL = permissible exposure limit; PQL = practical quantitation limit; REL = recommended exposure limit; RfC = reference concentration; RfD = reference dose; STLC = soluble threshold limit concentrations; TAP = toxic air pollutant; TDS = total dissolved solids; TLV = threshold limit value; TRI = Toxic Release Inventory; TSCA = Toxic Substances Control Act; TTLC = total threshold limit concentrations; TWA = time-weighted averages; USC = United States Code

<sup>&</sup>lt;sup>b</sup>Group 2B: possibly carcinogenic to humans

<sup>&</sup>lt;sup>c</sup>A3: confirmed animal carcinogen with unknown relevance to humans

Table 8-2. Regulations and Guidelines Applicable to Radioactive Cobalt

Agency	Description	Information	Reference
INTERNATIONAL Guidelines:			
IARC	Carcinogenicity classification	Group 1 (carcinogenic to humans)	IARC 2001b
ICRP	Occupational dose limits Effective dose	20 mSv per year, averaged over defined periods of 5 years	ICRP 1991
	Annual equivalent dose Lens of the eye Skin Hands and feet	150 mSv 500 mSv 500 mSv	
	General population dose limits Effective dose	1 mSv in a year	ICRP 1991
	Annual equivalent dose Lens of eye Skin	15 mSv 50 mSv	
WHO	Drinking water quality	No data	
NATIONAL Regulations and Guidelines:			
a. Air			
ACGIH	All radiation exposures must be kept as low as reasonably achievable		ACGIH 2000
	Effective dose Any single year Averaged over 5 years	50 mSv 20 mSv per year	ACGIH 2000
	Annual equivalent dose Lens of the eye Skin Hands and feet	150 mSv 500 mSv 500 mSv	
	Embryo-fetus exposures once the pregnancy is known Monthly equivalent dose Dose to the surface of women's abdomen (lower trunk) Intake of radionuclide	0.5 mSv 2 mSv for the remainder of the pregnancy 1/20 of the ALI	

Table 8-2. Regulations and Guidelines Applicable to Radioactive Cobalt *(continued)* 

Agency	Description	Information		Reference
NATIONAL (cont.)	r r r			
DOE	Radiation standards Inhalation DAC (µCi/mL)  55Co 56Co 57Co 58mCo 58Co 60mCo 60Co 61Co 62mCo	Class W <sup>a</sup> 1x10 <sup>-6</sup> 1x10 <sup>-7</sup> 1x10 <sup>-6</sup> 4x10 <sup>-5</sup> 5x10 <sup>-7</sup> 2x10 <sup>-3</sup> 7x10 <sup>-8</sup> 3x10 <sup>-5</sup> 7x10 <sup>-5</sup>	Class Y <sup>b</sup> 1x10 <sup>-6</sup> 8x10 <sup>-8</sup> 3x10 <sup>-7</sup> 3x10 <sup>-5</sup> 3x10 <sup>-7</sup> 1x10 <sup>-3</sup> 1x10 <sup>-8</sup> 2x10 <sup>-5</sup> 7x10 <sup>-5</sup>	DOE 2000 10CFR835 Appendix A
	Radiation standards Air immersion DAC <sup>c</sup> (µCi/mL) <sup>60m</sup> Co	1x10 <sup>-3</sup>		DOE 2000 10CFR835 Appendix C
EPA	Slope factors Inhalation (pCi) <sup>57</sup> Co <sup>58m</sup> Co <sup>58</sup> Co <sup>60</sup> Co	2.88x10 <sup>-12</sup> 8.90x10 <sup>-14</sup> 5.17x10 <sup>-12</sup> 6.88x10 <sup>-11</sup>		EPA 1997b
NIOSH	REL	No data		
NRC	Effluent concentrations—air <sup>55</sup> Co  Class W <sup>d</sup> Class Y <sup>e</sup> <sup>56</sup> Co  Class W <sup>d</sup>	ALI (μCi/mL)  4x10 <sup>-9</sup> 4x10 <sup>-9</sup> 4x10 <sup>-10</sup>		NRC 2001k 10CFR20 Appendix B Table 2
	Class W Class Y <sup>e</sup> 57Co Class W <sup>d</sup> Class Y <sup>e</sup>	3x10 <sup>-10</sup> 4x10 <sup>-9</sup> 9x10 <sup>-10</sup>		
	58Co Class W <sup>d</sup> Class Y <sup>e</sup>	2x10 <sup>-9</sup> 1x10 <sup>-9</sup>		
	Class W <sup>d</sup> Class Y <sup>e</sup> <sup>60</sup> Co	1x10 <sup>-7</sup> 9x10 <sup>-8</sup>		
	Class W <sup>d</sup> Class Y <sup>e</sup> 60mCo	2x10 <sup>-10</sup> 5x10 <sup>-11</sup>		
	Class W <sup>d</sup> Class Y <sup>e</sup>	6x10 <sup>-6</sup> 4x10 <sup>-6</sup>		

Table 8-2. Regulations and Guidelines Applicable to Radioactive Cobalt *(continued)* 

Agency	Description	Informati	on	Reference
NATIONAL (cont.)				
NRC (cont.)	Effluent concentrations—air  61Co Class Wd	<u>ALI (μCi/r</u> 9x10 <sup>-8</sup>	nL)	NRC 2001k 10CFR20 Appendix B
	Class Y <sup>e</sup> <sup>62m</sup> Co	8x10 <sup>-8</sup>		Table 2
	Class W <sup>d</sup> Class Y <sup>e</sup>	2x10 <sup>-7</sup> 2x10 <sup>-7</sup>		
	Occupational values			NRC 2001k
	—inhalation <sup>55</sup> Co	<u>ALI (μCi)</u>	DAC (µCi/mL)	10CFR20 Appendix B
	Class W <sup>d</sup>	$3x10^3$	1x10 <sup>-6</sup>	Table 1
	Class Y <sup>e</sup> <sup>56</sup> Co	3x10 <sup>3</sup>	1x10 <sup>-6</sup>	
	Class W <sup>d</sup>	$3x10^{2}$	1x10 <sup>-7</sup>	
	Class Y <sup>e</sup> <sup>57</sup> Co	2x10 <sup>2</sup>	8x10 <sup>-8</sup>	
	Class W <sup>d</sup>	$3x10^{3}$	1x10 <sup>-6</sup>	
	Class Y <sup>e</sup> <sup>58</sup> Co	7x10 <sup>2</sup>	3x10 <sup>-7</sup>	
	Class W <sup>d</sup>	$1x10^{3}$	5x10 <sup>-7</sup>	
	Class Y <sup>e</sup> <sup>58m</sup> Co	7x10 <sup>2</sup>	3x10 <sup>-7</sup>	
	Class W <sup>d</sup>	9x10 <sup>4</sup>	4x10 <sup>-5</sup>	
	Class Y <sup>e</sup> <sup>60</sup> Co	6x10⁴	3x10 <sup>-5</sup>	
	Class W <sup>d</sup>	2x10 <sup>2</sup>	7x10 <sup>-8</sup>	
	Class Y <sup>e</sup> <sup>60m</sup> Co	3x10 <sup>1</sup>	1x10 <sup>-8</sup>	
	Class W <sup>d</sup>	4x10 <sup>6</sup>	2x10 <sup>-3</sup>	
	Class Y <sup>e</sup> <sup>61</sup> Co	3x10 <sup>6</sup>	1x10 <sup>-3</sup>	
	Class W <sup>d</sup>	6x10 <sup>4</sup>	3x10 <sup>-5</sup>	
	Class Y <sup>e</sup> <sup>62m</sup> Co	6x10⁴	2x10 <sup>-5</sup>	
	Class W <sup>d</sup> Class Y <sup>e</sup>	2x10⁵ 2x10⁵	7x10⁻⁵ 6x10⁻⁵	
OSHA	Safety and health regulations for construction—ionizing radiation			OSHA 2001e 29CFR1926.53
	Toxic and hazardous substances—ionizing radiation			OSHA 2001d 29CFR1910.1096

Table 8-2. Regulations and Guidelines Applicable to Radioactive Cobalt *(continued)* 

Agency	Description	Information	Reference
NATIONAL (cont.)			
b. Water			
EPA	Drinking water standards		EPA 2000
	Beta particle and photon activity (formerly man-made radionuclides)  MCL  Caner risk at 10 <sup>-4</sup>	4 mrem 4 mrem/year	
	Gross alpha particle activity MCL Caner risk at 10 <sup>-4</sup>	15 pCi/L 15 pCi/L	
	Carcinogenic classification	Group A (human carcinogen)	
NRC	Effluent concentrations—water	ALI (μCi/mL)	NRC 2001k
	<sup>55</sup> Co Class W <sup>d</sup>	2x10 <sup>-5</sup>	10CFR20 Appendix B
	<sup>56</sup> Co Class W <sup>d</sup> <sup>57</sup> Co	6x10 <sup>-6</sup>	Table 2
	Class W <sup>d</sup>	6x10 <sup>-5</sup>	
	<sup>58</sup> Co Class W <sup>d</sup> <sup>58m</sup> Co	2x10 <sup>-5</sup>	
	Class W <sup>d</sup>	8x10 <sup>-4</sup>	
	<sup>60</sup> Co Class W <sup>d</sup> <sup>60m</sup> Co	3x10 <sup>-6</sup>	
	Class W <sup>d</sup>	2x10 <sup>-2</sup>	
	<sup>61</sup> Co Class W <sup>d</sup> <sup>62m</sup> Co	3x10 <sup>-4</sup>	
	Class W <sup>d</sup>	7x10 <sup>-4</sup>	
	Releases to sewers—monthly average concentration <sup>55</sup> Co	ALI (µCi/mL)	NRC 2001k 10CFR20
	Class W <sup>d</sup>	2x10 <sup>-4</sup>	Appendix B Table 3
	<sup>56</sup> Co Class W <sup>d</sup> <sup>57</sup> Co	6x10 <sup>-5</sup>	
	Class W <sup>d</sup> ⁵8Co	6x10 <sup>-4</sup>	
	Class W <sup>d</sup> 58mCo	2x10 <sup>-4</sup>	
	Class W <sup>d</sup>	8x10 <sup>-3</sup>	
NATIONAL (cont.)			

NATIONAL (cont.)

Table 8-2. Regulations and Guidelines Applicable to Radioactive Cobalt *(continued)* 

Agency	Description	Information		Reference
NRC (cont.)	Releases to sewers—monthly average concentration <sup>60</sup> Co	ALI (μCi/mL)		NRC 2001k 10CFR20 Appendix B
	Class W <sup>d</sup>	3x10 <sup>-5</sup>		Table 3
	Class W <sup>d</sup>	2x10 <sup>-1</sup>		
	Class W <sup>d</sup> <sup>62m</sup> Co	3x10 <sup>-3</sup>		
	Class W <sup>d</sup>	7x10 <sup>-3</sup>		
c. Food and Drug				
FDA	Ionizing radiation for the treatment of poultry feed and poultry feed ingredients (energy sources)	loninzing radiati- limited to gamm from sealed unit	a rays	FDA 1999 21CFR579.40
	Requirements regarding certain radioactive drugs— <sup>58</sup> Co or <sup>60</sup> Co	Labeled cyanoc for use in intesti absorption studi	nal	FDA 2000d 21CFR310.503(c)
	Sources of radiation used for inspection of food, for inspection of packaged food, and for controlling food processing			FDA 2000c 21CFR179.21 (a)(2)
d. Other				
DOE	Values for establishing sealed radioactive source accountability and radioactive material posting and labeling requirements  56Co 57Co 58Co 60Co	Activity (μCi) 4.0x10 <sup>1</sup> 2.3x10 <sup>2</sup> 1.4x10 <sup>2</sup> 1.8x10 <sup>1</sup>		DOE 2000 10CFR835 Appendix E
DOT	Activity values (Ci) <sup>55</sup> Co <sup>56</sup> Co <sup>57</sup> Co <sup>58m</sup> Co <sup>58</sup> Co <sup>60</sup> Co	<u>A</u> <sub>1</sub> 13.5 8.11 216 1080 27.0 10.8	A <sub>2</sub> 13.5 8.11 216 1080 27.0 10.8	DOT 2001a 49CFR173.435 Table

Table 8-2. Regulations and Guidelines Applicable to Radioactive Cobalt *(continued)* 

Agency	Description	Information	Reference
NATIONAL (cont.)			
DOT	Superfund, reportable quantity (Ci) <sup>55</sup> Co <sup>56</sup> Co <sup>57</sup> Co <sup>58</sup> Co <sup>58m</sup> Co <sup>60</sup> Co <sup>60</sup> Co <sup>60m</sup> Co <sup>61</sup> Co <sup>62m</sup> Co	10 10 100 10 1,000 10 1,000 1,000 1,000	DOT 2001b 49CFR172.101 Appendix A Table 2
EPA	RfC RfD Carcinogenicity classification	No data	IRIS 2000
	Annual possession quantities for environmental compliance (Ci/year) <sup>56</sup> Co <sup>57</sup> Co <sup>58</sup> Co <sup>58m</sup> Co <sup>60</sup> Co <sup>60m</sup> Co <sup>61</sup> Co	Liquid/ Gas powder Solid 2.3x10-6 2.3x10-3 2.3 1.8x10-2 1.8x10-1 1.8x10-4 2.5x10-6 2.5x10-3 2.5 2.3x10-6 2.3x10-3 2.3 4.6x10-2 4.6x10-1 4.6x10-4 7.0 7.0x10-3 7.0x10-6 9.8x10-1 9.8x10-2 9.8x10-5	EPA 2001a 40CFR61 Appendix E Table 1
	Concentration levels for environmental compliance (Ci/m³) <sup>56</sup> Co <sup>57</sup> Co <sup>58</sup> Co <sup>58m</sup> Co <sup>60</sup> Co <sup>60</sup> Co <sup>60</sup> Co	1.8x10 <sup>-13</sup> 1.3x10 <sup>-12</sup> 6.7x10 <sup>-13</sup> 1.2x10 <sup>-10</sup> 1.7x10 <sup>-14</sup> 4.3x10 <sup>-9</sup> 4.5x10 <sup>-9</sup>	EPA 2001a 40CFR61 Appendix E Table 2
	Slope factors—ingestion (pCi) <sup>57</sup> Co <sup>58m</sup> Co <sup>58</sup> Co <sup>60</sup> Co	9.71x10 <sup>-13</sup> 9.46x10 <sup>-14</sup> 2.82x10 <sup>-12</sup> 1.89x10 <sup>-11</sup>	EPA 1997b
	Slope factors—soil for external exposure (year/pCi/g) <sup>57</sup> Co <sup>58m</sup> Co <sup>58</sup> Co <sup>60</sup> Co	2.07x10 <sup>-7</sup> 3.21x10 <sup>-11</sup> 3.73x10 <sup>-6</sup> 9.76x10 <sup>-6</sup>	EPA 1997b
NATIONAL (cont.)			

Table 8-2. Regulations and Guidelines Applicable to Radioactive Cobalt *(continued)* 

Agency	Description	Information	Reference
EPA	Superfund, reportable quantities (Ci)  55C0 56C0 57C0 58mC0 58C0 60mC0 60C0 61C0 62mC0	10 10 100 1,000 10 1,000 10 1,000 1,000	EPA 2001i 40CFR302.4 Appendix B
NCRP	Occupational exposures Effective dose limits Annual Cummulative	50 mSv 10 mSv x age	NCRP1993
	Equivalent dose annual limits Lens of eye Skin, hands, and feet	150 mSv 500 mSv	
	Public exposures (annual) Effective dose limits, continuous or frequent exposure	1 mSv	
	Effective dose limits, infrequent exposures	5 mSv	
	Equivalent dose limits Lens of eye Skin, hands, and feet	15 mSv 50 mSv	
	Embryo and fetus exposures (monthly) Effective dose limit	0.5 mSv	
NRC	Activity values for radionuclides (Ci) <sup>55</sup> Co <sup>56</sup> Co <sup>57</sup> Co <sup>58m</sup> Co <sup>58</sup> Co <sup>60</sup> Co	8.11 8. 216 216 1080 10 27.0 27	NRC 2001a 10CFR71 3.5 11 080 7.0 0.8

Table 8-2. Regulations and Guidelines Applicable to Radioactive Cobalt *(continued)* 

Agency	Description	Information		Reference
NATIONAL (cont.)				
NRC	Byproduct material listing  —exempt concentrations Liquid and solid concentration (µCi/mL²)  57 C 58 C 60 C	5x10 <sup>-3</sup> 1x10 <sup>-3</sup> 5x10 <sup>-4</sup>		NRC 2001e 10CFR30.70 Schedule A
	Byproduct material listing (μCi) <sup>58m</sup> Co <sup>58</sup> Co <sup>60</sup> Co	10 10 1		NRC 2001b 10CFR30.71 Schedule B
	Byproduct material listing (Ci) <sup>58m</sup> Co <sup>58</sup> Co <sup>60</sup> Co	Column I <sup>f</sup> 100 1.0 0.1	Column II <sup>9</sup> 1.0 0.01 1x10 <sup>-4</sup>	NRC 2001c 10CFR33.100 Schedule A
	Items containing byproduct material listing— <sup>60</sup> Co (µCi) Electron tubes Spark gap irradiators	1.0 1.0		NRC 2001d 10CFR30.15(a)(8)
	Medical use— <sup>60</sup> Co as a source for brachytherapy	As a sealed someodles and a cells for topical interstitial, and intracavitary to cancer	applicator al, d	NRC 2001h 10CFR35.400
	Occupational values—oral ingestion  55Co Class W <sup>d</sup> 56Co	<u>ALI (μCi)</u> 1x10 <sup>3</sup>		NRC 2001k 10CFR20 Appendix B Table 1
	Class W <sup>d</sup> Class Y <sup>e</sup> 57Co	5x10 <sup>2</sup> 4x10 <sup>2</sup>		
	Class W <sup>d</sup> Class Y <sup>e</sup> 58Co	8x10 <sup>3</sup> 4x10 <sup>3</sup>		
	Class W <sup>d</sup> Class Y <sup>e</sup> <sup>58m</sup> Co	2x10 <sup>3</sup> 1x10 <sup>3</sup>		
	Class W <sup>d</sup> <sup>60</sup> Co	6x10 <sup>4</sup>		
	Class W <sup>d</sup> Class Y <sup>e</sup>	5x10 <sup>2</sup> 2x10 <sup>2</sup>		

Table 8-2. Regulations and Guidelines Applicable to Radioactive Cobalt *(continued)* 

Agency	Description	Information	Reference
NATIONAL (cont.)			
NRC (cont.)	Occupational values—oral ingestion  60mCo Class Wd St. wall 61Co	<u>ALI (μCi)</u> 1x10 <sup>6</sup> 1x10 <sup>6</sup>	NRC 2001k 10CFR20 Appendix B Table 1
	Class W <sup>d</sup> Class Y <sup>e</sup> <sup>62m</sup> Co Class W <sup>d</sup> St. wall	2x10 <sup>4</sup> 2x10 <sup>4</sup> 5x10 <sup>4</sup> 4x10 <sup>4</sup>	
	Quantities of radioactive material requiring labeling (µCi) 58mCo 58Co 60Co	10 10 1	NRC 2001g 10CFR30 Appendix B
	Quantities of licensed material requiring labeling (μCi) <sup>55</sup> Co <sup>56</sup> Co <sup>57</sup> Co <sup>58m</sup> Co <sup>58</sup> Co <sup>60m</sup> Co <sup>60</sup> Co <sup>61</sup> Co <sup>62m</sup> Co	100 10 100 1,000 100 1,000 1 1,000 1,000	NRC 2001i 10CFR20 Appendix C
	Quantities of radioactive materials requiring need for an emergency plan Release fraction Quantity (Ci)	0.001% 5,000	NRC 2001j 10CFR30.72 Schedule C
	Radioactive waste classification Class A (Ci/m³)	#700	NRC 2001I 10CFR61.55
	Reports of individual monitoring—processing or manufacturing for distribution, byproduct material in quantities exceeding <sup>60</sup> Co (Ci)	1.0	NRC 2001f 10CFR20.2206 (a)(7)

Table 8-2. Regulations and Guidelines Applicable to Radioactive Cobalt (continued)

Agency	Description	Information	Reference
<u>STATE</u>			
Regulations and Guidelines:			
a. Air			
Alabama	HAP—radionuclides		BNA 2001
California	HAP—radionuclides		BNA 2001
Hawaii	HAP—radionuclides		BNA 2001
Illinois	Toxic air contaminant—radionuclides		BNA 2001
Kansas	HAP—radionuclides		BNA 2001
Kentucky	HAP—radionuclides		BNA 2001
Minnesota	HAP—radionuclides		BNA 2001
Missouri	HAP—radionuclides		BNA 2001
Nebraska	HAP—radionuclides		BNA 2001
New York	HAP—radionuclides		BNA 2001
Rhode Island	HAP—radionuclides		BNA 2001
Wyoming	HAP—radionuclides		BNA 2001

<sup>&</sup>lt;sup>a</sup>Class W: refers to the approximate length of retention in the pulmonary region which is 10–100 days for this class <sup>b</sup>Class Y: refers to the approximate length of retention in the pulmonary region which is greater than 100 days for this class

ACGIH = American Conference of Governmental Industrial Hygienists; ALI = annual limits on intake; BNA = Bureau of National Affairs; CFR = Code of Federal Regulations; DAC = derived air concentrations; DOE = Department of Energy; DOT = Department of Transportation; EPA = Environmental Protection Agency; FDA = Food and Drug Administration; IARC = International Agency for Research on Cancer; ICRP = International Commission on Radiological Protection; IRIS = Integrated Risk Information System; mSv = millisievert; NCRP = National Council on Radiation Protection; NIOSH = National Institute for Occupational Safety and Health; NRC = Nuclear Regulatory Commission; OSHA = Occupational Safety and Health Administration; PEL = permissible exposure limit; REL = recommended exposure limit; RfC = reference concentration; RfD = reference dose; TLV = threshold limit value; TWA = time-weighted averages; WHO = World Health Organization

<sup>&</sup>lt;sup>c</sup>Air immersion DAC values: based on a stochastic dose limit of 5 rems (0.05 Sv) per year or a nonstochastic (organ) dose limit of 50 rems (0.5 Sv) per year

<sup>&</sup>lt;sup>d</sup>Class W: all compounds except those given for Y

eClass Y: oxides, hydroxides, halides, and nitrates

<sup>&</sup>lt;sup>f</sup>Column I: gas concentration

<sup>&</sup>lt;sup>9</sup>Column II: liquid and solid concentration

COBALT 291

#### 9. REFERENCES

Aage HK, Korsbech U, Bargholz K, et al. 1999. A new technique for processing airborne gamma ray spectrometry data for mapping low level contaminations. Appl Radiat Isot 51:651-662.

\*Abbasi SA, Nipaney PC, Soni R. 1989. Environmental status of cobalt and its micro determination with 7-nitroso-8-hydroxyquinoline-5-sulfonic acid in waters, aquatic weeds and animal tissues. Anal Lett 22(1):225-235.

Abraham JL. 1990. The spectrum of pulmonary pathologic reaction and lung dust burden in 30 cases of cobalt pneumonitis (hard metal disease; giant cell interstitial pneumonia). Am Rev Respir Dis 141:A248.

\*Abraham JL, Hunt A. 1995. Environmental contamination by cobalt in the vicinity of a cemented tungsten carbide tool grinding plant. Environ Res 69:67-74.

Abramson DH, Ellsworth RM, Kitchin FD. 1980. Osteogenic sarcoma of the humorous after cobalt plaque treatment for retinoblastoma. Am J Ophthalmol 90:374-376.

ACGIH. 1999a. Cobalt: 1999 TLVs and BEIs: Threshold limit values for chemical substances and physical agents biological exposure indices. Cincinnati, OH: American Conference of Governmental Industrial Hygienists.

ACGIH. 1999b. Radioactive cobalt: 1999 TLVs and BEIs: Threshold limit values for chemical substances and physical agents biological exposure indices. Cincinnati, OH: American Conference of Governmental Industrial Hygienists.

\*ACGIH. 2000. Cobalt: 2000 TLVs and BEIs: Threshold limit values for chemical substances and physical agents biological exposure indices. Cincinnati, OH: American Conference of Governmental Industrial Hygienists.

Adachi S, Takemoto K, Ohshima S, et al. 1991. Metal concentration in lung tissue of subjects suffering from lung cancer. Int Arch Occup Environ Health 63:193-197.

Adamis Z, Tatrai E, Honma K, et al. 1997. A study on lung toxicity of respirable hard metal dusts in rats. Ann Occup Hyg 41(5):515-526.

- \*Adeloju SB, Bond AM, Briggs MH. 1985. Multi element determination in biological materials by differential pulse voltammetry. Anal Chem 57:1386-1390.
- \*Adinolfi M. 1985. The development of the human blood-CSF-brain barrier. Dev Med Child Neurol 27:532-537.
- \*Adlercreu'tz H. 1995. Phytoestrogens: Epidemiology and a possible role in cancer protection. Environ Health Perspect Suppl 103(7):103-112.

Adriano DC, Delaney M, Paine D. 1977. Availability of cobalt-60 to corn and bean seedlings as influenced by soil types, lime, and DTPA. Commun Soil Sci Plant Anal 8(8):615-628.

\_

<sup>\*</sup>Cited in text

# COBALT 292 9. REFERENCES

Ahmad S, Waheed S, Mannan A, et al. 1994. Evaluation of trace elements in wheat and wheat by-products. J AOAC Int 77(1):11-18.

Aiken G, Cotsaris E. 1995. Soil and hydrology: their effect on NOM. J Am Water Works Assoc January:36-45.

\*Alaux-Negrel G, Beaucaire C, Michard G, et al. 1993. Trace-metal behavior in natural granitic waters. J Contam Hydrol 13:309-325.

Alessio L, Dell'Orto A. 1988. Biological monitoring of cobalt. In: Biological monitoring of toxic metals. New York, NY: Plenum Press, 407-417.

\*Alexander CS. 1969. Cobalt and the heart. Ann Intern Med 70:411-413.

\*Alexander CS. 1972. Cobalt-beer cardiomyopathy: A clinical and pathological study of twenty-eight cases. Am J Med 53:395-417.

\*Alexandersson R. 1988. Blood and urinary concentrations as estimators of cobalt exposure. Arch Env Health 43(4):299-303.

Alexeeva-Popova NV, Igoshina TI, Drosdova IV. 1995. Metal distribution in the Arctic ecosystems of the Chukotka Peninsula, Russia. Sci Total Environ 160/161:643-652.

Alexiou D, Grimanis AP, Grimani M, et al. 1977. Trace elements (zinc, cobalt, selenium, rubidium, bromine, gold) in human placenta and newborn liver at birth. Pediatr Res 11:646-648.

Alfy SE, Abdel-Rassoul AA. 1993. Trace metal pollutants in El Manzala Lakes by inductively coupled plasma spectroscopy. Water Res 27(7):1253-1256.

Al-Jaloud AA, Hussain G, Al-Saati AJ, et al. 1995. Effect of wastewater irrigation on mineral composition of corn and sorghum plants in a pot experiment. J Plant Nutr 18(8):1677-1692.

Allen MJ, Myer BJ, Millett PJ, et al. 1997. The effects of particulate cobalt, chromium and cobalt-chromium alloy on human osteoblast-like cells in vitro. J Bone Jt Surg Am 79-B:475-482.

\*Alomar A, Conde-Salazar L, Romaguera C. 1985. Occupational dermatosis from cutting oils. Contact Dermatitis 12:129-138.

Alonso MT, Sanchez A, Garcia-Sancho J. 1990. Arachidonic acid-induced calcium influx in human platelets: Comparison with the effect of thrombin. Biochem J 272:435-443.

Al-Saleh IA. 1996. Trace elements in drinking water coolers collected from primary schools, Riyadh, Saudi Arabia. Sci Total Environ 181:215-221.

Al-Tawil NG, Marcusson JA, Moller E. 1985. HLA-class II restriction of the proliferative T lymphocyte responses to nickel, cobalt and chromium compounds. Tissue Antigens 25:163-172.

\*Altman PL, Dittmer DS. 1974. In: Biological handbooks: Biology data book. Vol. III. 2<sup>nd</sup> ed. Bethesda, MD: Federation of American Societies for Experimental Biology, 1987-2008, 2041.

Ambrosini MV, Principato GB, Giovannini E, et al. 1979. Acid-base balance changes and erythropoietin production in the early stages of hypoxia or after CoCl<sub>2</sub> treatment in the rabbit. Acta Hematol 62:32-40.

# COBALT 293 9. REFERENCES

- \*Amiard JC, Amiard-Triquet C. 1979. Distribution of cobalt 60 in a mollusc, a crustacean and a freshwater teleost: Variations as a function of the source of pollution and during elimination. Environ Pollut 20(3):199-213.
- \*Amundsen CE, Hanssen JE, Semb A, et al. 1992. Long-range atmospheric transport of trace elements to Southern Norway. Atmos Environ 26A(7):1309-1324.
- Anard D, Kirsch-Volders M, Elhajouji A, et al. 1997. *In vitro* genotoxic effects of hard metal particles assessed by alkaline single cell gel and elution assays. Carcinogenesis 18(1):177-184.
- \*Andersen O. 1983. Effects of coal combustion products and metal compounds on sister chromatid exchange (SCE) in a macrophage like cell line. Environ Health Perspect 47:239-253.
- \*Andersen ME, Krishnan K. 1994. Relating in vitro to in vivo exposures with physiologically based tissue dosimetry and tissue response models. In: Salem H, ed. Animal test alternatives: Refinement, reduction, replacement. New York: Marcel Dekker, Inc., 9-25.
- \*Andersen ME, Clewell HJ III, Gargas ML, et al. 1987. Physiologically based pharmacokinetics and the risk assessment process for methylene chloride. Toxicol Appl Pharmacol 87:185-205.
- Anderson MB, Pedigo N, George WJ. 1986. Reproductive effects of chronic oral administration of cobaltous chloride in male mice. Biol Reprod 34:186.
- \*Anderson MB, Pedigo NG, Katz RP, et al. 1992. Histopathology of testes from mice chronically treated with cobalt. Reprod Toxicol 6:41-50.
- \*Anderson MB, Lepak K, Farinas V, et al. 1993. Protective action of zinc against cobalt-induced testicular damage in the mouse. Reprod Toxicol 7:49-54.
- Anderson PR, Christensen TH. 1988. Distribution coefficients of Cd, Co, Ni and Zn in soils. J Soil Sci 39:15-22.
- \*Andre S, Metivier H, Masse R. 1989. An interspecies comparison of the lung clearance of inhaled monodisperse cobalt oxide particles- part III: Lung clearance of inhaled cobalt oxide particles in baboons. J Aerosol Sci 20(2):205-217.
- Andreev G, Simenov V. 1990. Distribution and correlation of elements in waters, suspensions, sediments and marine organisms from the Black Sea. Toxicol Environ Chem 28:1-9.
- Andreu V, Gimeno-Garcia E. 1996. Total content and extractable fraction of cadmium, cobalt, copper, nickel, lead, and zinc in calcareous orchard soils. Commun Soil Sci Plant Anal 27:2633-2648.
- \*Andrzejewski SW, Zawisza B, Wybrzak-Wrobel T. 1980. Dose-related <sup>60</sup>Co γ-ray-induced oxygen uptake and citrulline production in liver mitochondria of whole-body irradiated rats. Biochem Med 23:282-292.
- Angelidis M, Grimanis AP. 1989. Geochemical partitioning of Co, Cr, Fe, Sc and Zn in polluted and non-polluted marine sediments. Environ Pollut 62:31-46.
- Angerer J, Heinrich-Ramm R, Lehnert G. 1989. Occupational exposure to cobalt and nickel: Biological monitoring. Int J Environ Anal Chem 35:81-88.

# COBALT 294 9. REFERENCES

ANL. 2000. Environmental monitoring at Argonne National Laboratory. Argonne, IL: Argonne National Laboratory. <a href="http://www.anl.gov/OPA/env/EMfacts.html">http://www.anl.gov/OPA/env/EMfacts.html</a>. May 15, 2000.

\*Antilla S, Sutitnen S, Paananen M, et al. 1986. Hard metal lung disease: A clinical, histological, ultra structural and x-ray micro analytical study. Eur J Respir Dis 69:83-94.

\*APHA. 1998. Standard methods for the examination of water and wastewater, 20th edition. Washington, DC: American Public Health Association.

\*Apostoli P, Porru S, Alessio L. 1994. Urinary cobalt excretion in short time occupational exposure to cobalt powders. Sci Total Environ 150:129-132.

Apostoli P, Giusti S, Bartoli D, et al. 1998. Multiple exposure to arsenic, antimony, and other elements in art glass manufacturing. Am J Ind Med 34:65-72.

Arai F, Yamamura Y, Yoshida M, et al. 1994. Blood and urinary levels of metals (Pb, Cr, Mn, Sb, Co and Cu) in cloisonne workers. Ind Health 32:67-78.

Archer RD. 1979. Coordination compounds. In: Kirk RE, Othmer DF, Grayson M, et al., ed. Kirk-Othmer encyclopedia of chemical technology. New York, NY: John Wiley and Sons, 793-797.

\*Arimoto R, Duce RA, Ray BJ, et al. 1985. Atmospheric trace elements at Enewetak Atoll: 2. Transport to the ocean by wet and dry deposition. J Geophys Res 90(D1):2391-2408.

Arizono K, Okanari E, Ueno K, et al. 1991. Heme oxygenase activity and cytochrome P-450 content associated with induced metallothionein in the liver of rats treated with various metals. J Environ Sci Health Part A 26(6):941-951.

Arkhipova OG, Golubovidh EY, Spiridonova VI. 1965. Effect of chelating agents on cobalt elimination and glycylglycine dipeptidase activity. Fed Proc 25(1):T93-T94.

\*Arlauskas A, Baker RSU, Bonin AM, et al. 1985. Mutagenicity of metal ions in bacteria. Environ Res 36:379-388.

Ashraf W, Jaffar M, Mohammad D. 1994. Trace metal contamination study on scalp hair of occupationally exposed workers. Bull Environ Contam Toxicol 53:516-523.

\*Ashraf W, Jaffar M, Mohammad D. 1995. Levels of selected trace metals in hair of urban and rural adult male population of Pakistan. Bull Environ Contam Toxicol 54:207-213.

\*Asmuß M, Mullenders LH, Hartwig A. 2000. Interference by toxic metal compounds with isolated zinc finger DNA repair proteins. Toxicol Lett 112-113:227-231.

\*ASTM. 1999. Annual Book of ASTM Standards, vol. 11.02. American Society for Testing of Materials. Philadelphia, PA: ASTM, 290-300.

Astrup A, Tuchsen F. 1990. Cobalt exposure and cancer risk. Crit Rev Toxicol 20(6):427-437.

\*ATSDR. 1989. Decision guide for identifying substance-specific data needs related to toxicological profiles; Notice. Agency for Toxic Substances and Disease Registry. Federal Register 54(174):37618-37634.

# COBALT 295 9. REFERENCES

- \*ATSDR. 1990. Biomarkers of organ damage or dysfunction for the renal, hepatobiliary, and immune systems. Subcommittee on Biomarkers of Organ Damage and Dysfunction, Agency for Toxic Substances and Disease Registry, Atlanta, GA.
- \*ATSDR. 1999. Toxicological profile for ionizing radiation. Agency for Toxic Substances and Disease Registry, Atlanta, GA.
- ATSDR. 2001. Public health assessment: Blackbird Mine, cobalt, Lemhi county, Idaho: ATSDR. <a href="http://atsdr.edc.gov/HAC/PHA/blackbird/bla*pl.html">http://atsdr.edc.gov/HAC/PHA/blackbird/bla<i>pl.html*</a>. May 29, 2001.
- Auchincloss JH, Abraham JL, Gilbert R, et al. 1992. Health hazard of poorly regulated exposure during manufacture of cemented tungsten carbides and cobalt. Br J Ind Med 49:832-836.
- \*Augsburger JJ, Shields JA. 1985. Cataract surgery following cobalt-60 plaque radiotherapy for posterior uveal malignant melanoma. Ophthalmology 92:815-822.
- Avery EL, Dunstan RH, Nell JA. 1996. The detection of pollutant impact in marine environments: Condition index, oxidative DNA damage, and their associations with metal bioaccumulation in the sydney rock oyster *saccostrea commercialis*. Arch Environ Contam Toxicol 31:192-198.
- \*Ayala-Fierro F, Firriolo JM, Carter DE. 1999. Disposition, toxicity, and intestinal absorption of cobaltous chloride in male Fischer 344 rats. J Toxicol Environ Health, Part A 56:571-591.
- \*Badsha KS, Goldspink CR. 1988. Heavy metal levels in three species of fish in Tjeukemeer, a Dutch polder lake. Chemosphere 17(2):459-463.
- \*Baes CF, Sharp RD. 1983. A proposal for estimation of soil leaching and leaching constants for use in assessment models. J Environ Qual 12(1):17-28.
- \*Bailey MR, Kreyling WG, Andre S, et al. 1989. An interspecies comparison of the lung clearance of inhaled monodisperse cobalt oxide particles- Part 1: Objectives and summary of results. J Aerosol Sci 20(2):169-188.
- \*Baker DH, Czarnecki-Maulden GL. 1987. Pharmacologic role of cysteine in ameliorating or exacerbating mineral toxicities. J Nutr 117:1003-1010.
- Balakrishnan S, Rao SB. 1999. Cytogenetic analysis of peripheral blood lymphocytes of occupational workers exposed to low levels of ionizing radiation. Mutat Res 442:37-42.
- Baldetorp L. 1977. Effect of 50 kV roentgen rays and cobalt-60 gamma rays in the activity of ciliated cells. Acta Radiologica Therapy Physics Biology 16:406-416.
- Balogh I, Rozsalyi K, Kovach A, et al. 1987. Endothelial cell injuries on the isolated rat heart after perfusion with trace elements. J Mol Cell Cardiol 19(III):S4.
- Banchereau J, Dubos M, Agneray J, et al. 1982. A direct evidence for the early membrane desialylation in cobalt-irradiated mouse lymphocytes. Biochem Biophys Res Commun 104(2):512-516.
- Banerjee RK, Datta AG. 1971. Effect of cobalt and vitamin  $B_{12}$  on the peroxides and iodinating activity of mouse thyroid and submaxillary gland: In vitro stimulation of vitamin  $B_{12}$  coenzyme on the iodination of tyrosine. Endocrinology 88:1456-1464.

# COBALT 296 9. REFERENCES

\*Baratta EJ, Apidianakis JC, Ferri ES. 1969. Cesium-137, lead-210 and polonium-210 concentrations in selected human tissues in the United States. Am Ind Hyg Assoc J 30:443-448.

Barbera R, Farre R. 1988. Determination of cobalt in foods by flame and electro thermal atomization-atomic absorption spectrometry. A comparative study. Atom Spectrosc 9(1):6-8.

\*Barborik M, Dusek J. 1972. Cardiomyopathy accompanying industrial cobalt exposure. Br Heart J 34:113-116.

\*Barceloux DG. 1999. Cobalt. Clin Toxicol 37(2):201-216.

\*Bargagli R. 2000. Trace metals in Antarctica related to climate changes and increasing human impact. Rev Environ Contam Toxicol 166:129-173.

\*Bargagli R, Barghigiani C, Siegel BZ, et al. 1991. Trace metal anomalies in surface soils and vegetation on two active island volcanos: Stromboli and Vulcano (Italy). Sci Total Environ 102:209-222.

Barman SC, Bhargave SK. 1997. Accumulation of heavy metals in soil and plants in industrially polluted field. In: Cheremisinoff PN, ed. Ecological issues and environmental impact assessment. Houston, TX: Gulf Publishing Company, 289-314.

\*Barnaby CF, Smith T. 1971. Calibration of a whole-body counter suitable for use in routine clinical investigations. Phys Med Biol 16:97-104.

\*Barnaby CF, Smith T, Thompson BD. 1968. Dosimetry of the radioisotopes of cobalt. Phys Med Biol 13(3):421-433.

\*Barnes DG, Dourson M. 1988. Reference dose (RfD): Description and use in health risk assessments. Regul Toxicol Pharmacol 8:471-486.

\*Barnes JE, Kanapilly GM, Newton GJ. 1976. Cobalt-60 oxide aerosols: Methods of production and short-term retention and distribution kinetics in the beagle dog. Health Phys 30:391-398.

Barnhart S, Daniell W, Stebbins A, et al. 1991. Occurrence of hard metal pneumoconiosis at exposure levels below the permissible exposure limit. Am Rev Respir Dis 143:A263.

Barton JC, Conrad ME, Holland R. 1981. Iron, lead, and cobalt absorption: Similarities and dissimilarities. Proc Soc Exp Biol Med 166:64-69.

Basaham AS, Al-Lihaibi SS. 1993. Trace elements in sediments of the western gulf. Mar Pollut Bull 27:103-107.

Basketter DA, Briatico-Vangosa G, Kaestner W, et al. 1993. Nickel, cobalt and chromium in consumer products: A role in allergic contact dermatitis? Contact Dermatitis 28:15-25.

\*Bassant MH, Court L. 1978. Effects of whole-body irradiation on the activity of rabbit hippocampal neurons. Radiat Res 75:593-606.

\*Baudin JP, Fritsch AF. 1987. Retention of ingested <sup>60</sup>Co by a freshwater fish. Water Air Soil Pollut 36:207-217.

# COBALT 297 9. REFERENCES

\*Baudin JP, Fritsch AF. 1989. Relative contributions of food and water in the accumulation of <sup>60</sup>Co by a freshwater fish. Water Res 23(7):817-823.

Baudin JP, Nucho R. 1992. <sup>60</sup>Co accumulation for sediment and planktonic algae by midge larvae (chironomus luridus). Environ Pollut 76:133-140.

\*Baudin JP, Fritsch AF, Georges J. 1990. Influence of labeled food type on the accumulation and retention of <sup>60</sup>Co by a freshwater fish, Cyprinus carpio L. Water Air Soil Pollut 51:261-270.

\*Baumgardt B, Jackwerth E, Otto H, et al. 1986. Trace analysis to determine heavy metal load in lung tissue: A contribution to substantiation of occupational hazards. Int Arch Occup Environ Health 58:27-34.

Bearden LJ. 1976. The toxicity of two prosthetic metals (cobalt and nickel) to cultured fibroblasts. Diss Abstr Int B 37(4):1785.

\*Beaugelin-Seiller K, Baudin JP, Brottet D. 1994. Use of aquatic mosses for monitoring artificial radionuclides downstream of the nuclear power plant of Bugey (River Rhone, France). J Environ Radioact 24:217-233.

\*Becker DE, Smith SE. 1951. The level of cobalt tolerance in yearling sheep. J Anim Sci 10:266-271.

Becker G, Osterloh K, Schafer S, et al. 1981. Influence of fucoidan on the intestinal absorption of iron, cobalt, manganese and zinc in rats. Digestion 21:6-12.

Beckett WS, Figueroa S, Gerstenhaber B, et al. 1992. Pulmonary fibrosis associated with occupational exposure to hard metal at a metal-coating plant - Connecticut, 1989. MMWR Morb Mortal Wkly Rep 41(4):65-67.

\*Bedello PG, Goitre M, Alovisi V, et al. 1984. Contact dermatitis caused by cobalt naphthenate. Contact Dermatitis 11:247-264.

\*Behrooz A, Ismail-Beigi F. 1997. Dual control of glut1 glucose transporter gene expression by hypoxia and by inhibition of oxidative phosphorylation. J Biol Chem 272(9):5555-5562.

\*Beijer K, Jernelov A. 1986. Sources, transport and transformation of metals in the environment. In: Handbook on the toxicology of metals: Volume I: General aspects. New York, NY: Elsevier Science Publishing Co., Inc., 68-84.

\*BEIR V. 1990. Health effects of exposure to low levels of ionizing radiation. Biological Effects of Ionizing Radiations. Washington, DC: National Academy Press.

Beitler JJ, McCormick B, Ellsworth RM, et al. 1990. Ocular melanoma: Total dose and dose rate effect with Co-60 plaque therapy. Radiology 176:275-278.

\*Beleznay E, Osvay M. 1994. Long-term clearance of accidentally inhaled <sup>60</sup>Co aerosols in humans. Health Phys 66:392-399.

\*Bellet-Barthas M, Barthelemy L, Bellet M. 1980. Effects of <sup>60</sup>Co radiation on the rabbit lung surfactant system. Int J Radiat Oncol Biol Phys 6:1169-1177.

# COBALT 298 9. REFERENCES

- \*Bencko V, Wagner V, Wagnerova M, et al. 1983. Immuno-biochemical findings in groups of individuals occupationally and non-occupationally exposed to emissions containing nickel and cobalt. J Hyg Epidemiol Microbiol Immunol 27(4):387-394.
- \*Bencko V, Wagner V, Wagnerova M, et al. 1986. Human exposure to nickel and cobalt: Biological monitoring and immunobiochemical response. Environ Res 40:399-410.
- Benes P, Cernik M. 1992. Kinetics of radionuclide interaction with suspended solids in modeling the migration of radionuclides in rivers: II. Effect of concentration of the solids and temperature. J Radioanal Nucl Chem 159(2):187-200.
- \*Benes P, Jurak M, Crenik M. 1989a. Factors affecting interaction of radiocobalt with river sediments: II. Composition and concentration of sediment temperature. J Radioanal Nucl Chem 132(2):225-239.
- \*Benes P, Jurak M, Kunkova M. 1989b. Factors affecting interaction of radiocobalt with river sediments: I. pH and composition of water and contact time. J Radioanal Nucl Chem 132(2):209-223.
- Benes P, Kuncova M, Slovak J, et al. 1988. Analysis of the interaction of radionuclides with solid phase in surface waters using laboratory model experiments: Methodical problems. J Radioanal Nucl Chem 125(2):295-315.
- \*Benjamin SA, Lee AC, Angleton GM, et al. 1998a. Mortality in beagles irradiated during prenatal and postnatal development. I. Contribution of non-neoplastic diseases. Radiat Res 150:316-329.
- \*Benjamin SA, Lee AC, Angleton GM, et al. 1998b. Mortality in beagles irradiated during prenatal and postnatal development. II. Contribution of benign and malignant neoplasia. Radiat Res 150:330-348.
- \*Benjamin SA, Saunders WJ, Angleton GM, et al. 1991. Radiation carcinogenesis in dogs irradiated during prenatal postnatal development. J Radiat Res 2(Suppl.):86-103.
- \*Benjamin SA, Saunders WJ, Lee AC, et al. 1997. Non-neoplastic and neoplastic thyroid disease in beagles irradiated during prenatal and postnatal development. Radiat Res 147:422-430.
- Bennett JE. 1968. Treatment of carcinoma of the prostate. Radiology 90:532-535.
- \*Berg JW, Burbank F. 1972. Correlations between carcinogenic trace metals in water supplies and cancer mortality. Ann NY Acad Sci 199:249-264.
- \*Berger GS. 1994. Epidemiology of endometriosis. In: Berger GS, ed. Endometriosis: Advanced management and surgical techniques. New York, NY: Springer-Verlag.
- \*Berger ME, Hurtado R, Dunlap J, et al. 1997. Accidental radiation injury to the hand: anatomical and physiological considerations. Health Physics 72(3):343-348.
- Berkenstock OL. 1992. Issues concerning possible cobalt-chromium carcinogenicity: A literature review and discussion. Contemporary Orthopaedics 24(3):265-278.
- \*Bernstein H-G, Keilhoff G, Kirschke H, et al. 1986. Cathepsins B and D in rat brain glia during experimentally induced neuropathological defects. An immunocytochemical approach. Biomed Biochem Acta 45:1461-1464.

# COBALT 299 9. REFERENCES

Bertine KK, Goldberg ED. 1972. Trace elements in clams, mussels, and shrimp. Limnol Oceanogr 17(6):877-884.

\*Beskid M. 1963. The effect of administration of cobalt chloride on the pancreas in the guinea-pig. Folia Histochem Cytochem 1(1):95-102.

Betti M, Giannarelli S, Hiernaut T, et al. 1996. Detection of trace radioisotopes in soil, sediment and vegetation by glow discharge mass spectrometry. Fresenius J Anal Chem 355:642-646.

Beyersmann D. 1994. Interactions in metal carcinogenicity. Toxicol Lett 72:333-338.

Beyersmann D, Hartwig A. 1992. The genetic toxicology of cobalt. Toxicol Appl Pharmacol 115:137-145

\*Bezek S, Trnovec T, Scasnar V, et al. 1990. Irradiation of the head by <sup>60</sup>Co opens the blood-brain barrier for drugs in rats. Experientia 46:1017-1020.

\*Bhat IS, Hedge AG, Chandramouli S, et al. 1973. Evaluation of internal exposure to radionuclides of I, Cs, and Co, during maintenance operations on primary steam leak in a nuclear power station. Health Phys 25:135-139.

\*Bibak A, Behrens A, Sturup S, et al. 1998a. Concentration of 55 major trace elements in Danish agricultural crops measured by inductively coupled plasma mass spectrometry. 2. Pea (*pisum sativum* ping pong). J Agric Food Chem 46:3146-3149.

\*Bibak A, Behrens A, Sturup S, et al. 1998b. Concentrations of 63 major and trace elements in Danish agricultural crops measured by inductively coupled plasma mass spectrometry. 1. Onion (*Allium cepa Hysam*). J Agric Food Chem 46:3139-3145.

Bieger W, Seybold J, Kern HF. 1975. Studies on intracellular transport of secretory proteins in the rat exocrine pancreas: III. Effect of cobalt, lanthanum, and antimycin A. Virchows Arch A Pathol Anat Histol 368:329-345.

\*Biego GH, Joyeux M, Hartemann P, et al. 1998. Daily intake of essential minerals and metallic micropollutants from foods in France. Sci Total Environ 217:27-36.

Bingham D, Harrison JD, Phipps AW. 1997. Biokinetics and dosimetry of chromium, cobalt, hydrogen, iron and zinc radionuclides in male reproductive tissues of the rat. Int J Radiat Biol 72(2):235-248.

\*Bird GA, Hesslein RH, Mills KH, et al. 1998a. Bioaccumulation of radionuclides in fertilized Canadian Shield Lake basins. Sci Total Environ 218:67-83.

Bird GA, Mills KH, Schwartz WJ. 1999. Accumulation of <sup>60</sup>Co and <sup>134</sup>Cs in lake whitefish in a Canadian shield lake. Water Air Soil Pollut 114:303-322.

\*Bird GA, Schwartz WJ, Motycka M, et al. 1998b. Behavior of <sup>60</sup>Co and <sup>134</sup>Cs in a Canadian shield lake over 5 years. Sci Total Environ 212:115-135.

Blalock TL, Hill CH. 1986. Mechanism of alleviation of Zn, Cd, V, Ni and Co toxicities by dietary iron. Fed Proc 45:369.

# COBALT 300 9. REFERENCES

Blume HP, Brummer G. 1991. Prediction of heavy metal behavior in soil by means of simple field tests. Ecotoxicol Environ Saf 22:164-174.

\*BNA. 2001. Environment and Safety Library on the Web States and Territories. Washington, DC: Bureau of National Affairs, Inc. Http://www.esweb.bna.com/. June 06, 2001.

\*Boccolini A, De Franceschi L, Gentili A, et al. 1976. <sup>60</sup>Co in air. Health Phys 31:175-176.

Bode P, De Bruin M, Aalbers TG, et al. 1990. Plastics from household waste as a source of heavy metal pollution. Biol Trace Elem Res 27:377-384.

\*Boikat U, Fink A, Bleck-Neuhaus J. 1985. Cesium and cobalt transfer from soil to vegetation on permanent pastures. Radiation and Environmental Biophysics 24:287-301.

\*Bond AM, Wallace GG. 1984. Liquid chromatography with electrochemical and/or spectrophotometric detection for automated determination of lead, cadmium, mercury, cobalt, nickel, and copper. Anal Chem 56:2085-2090.

Borg H, Johansson K. 1989. Metal fluxes to Swedish forest lakes. Water Air Soil Pollut 47:427-440.

\*Bouman AA, Platenkamp AJ, Posma FD. 1986. Determination of cobalt in urine by flameless atomic absorption spectrometry. Comparison of direct analysis using Zeeman background correction and indirect analysis using extraction in organic solution. Ann Clin Biochem 23:346-350.

\*Bourg WJ, Nation JR, Clark DE. 1985. The effects of chronic cobalt exposure on passive-avoidance performance in the adult rat. Bulletin of the Psychonomic Society 23(6):527-530.

\*Brady ME, Hayton WL. 1977a. GI drug absorption in rats exposed to cobalt-60  $\gamma$ -radiation I: Extent of absorption. J Pharm Sci 66(3):361-365.

\*Brady ME, Hayton WL. 1977b. GI drug absorption in rats exposed to cobalt-60 γ-radiation II: *In vivo* rate of absorption. J Pharm Sci 66(3):366-370.

Braham HW, Sacher GA. 1978. Metabolic and thermoregulatory effects of acute <sup>60</sup>Co radiation in myomorph rodents. Radiat Res 75:108-120.

Braham JL. 1987. Lung pathology in 22 cases of giant cell interstitial pneumonia (GIP) suggests GIP os pathognomic of cobalt (hard metal) disease. Chest 91(2):312.

Brasch J, Geier J. 1997. Patch test results in schoolchildren: Results from the information network of departments of dermatology (IVDK) and the German Contact Dermatitis Research Group (DKG). Contact Dermatitis 37:286-293.

Braselmann H, Schmid E, Bauchinger M. 1994. Chromosome aberrations in nuclear power plant workers: the influence of dose accumulation and lymphocyte life-time. Mutat Res 306:197-202.

Breccia A, Balducci R, Stagni G. 1982. Electrochemical studies on nitroimidazole sensitizers: Interaction with Co(II), Zn(II), and Fe(III) in biological media. Int J Radiat Oncol Biol Phys 8:423-426.

\*Bregman B, Le Saux F, Trottier S, et al. 1985. Chronic cobalt-induced epilepsy: Noradrenaline ionophoresis and adrenoceptor binding studies in the rat cerebral cortex. J Neural Transm 63:109-118.

# COBALT 9. REFERENCES 301

- \*Brewer G. 1940. A statistical study of cobalt polycythemia in the dog. Am J Physiol 128:345-348.
- \*Brizzee KR, Ordy JM, Kaak B, et al. 1978. Prenatal cobalt-60 irradiation effects on early postnatal development of the squirrel monkey offspring. DOE Symp Ser 47:204-227.
- \*Bronstein AC, Currance PL. 1988. Emergency care for hazardous materials exposure. St. Louis, MO: CV Mosby Company.
- \*Brooks AL, Carsten AL, Mead DK, et al. 1974. Effect of <sup>60</sup>Co exposure or continuous intake of tritiated water on the liver chromosomes of hale-stoner brookhaven mice. In: Inhal. Toxicol. Research Institute Annual Report 1973-1974. Albuquerque, New Mexico: Lovelace Foundation for Medical Educational Research, 182-185.
- \*Brooks AL, Mead DK, Peters RF. 1971a. Effect of chronic exposure to <sup>60</sup>Co on the frequency of metaphase chromosome aberrations in the liver cells of the Chinese hamster (in vivo). Int J Radiat Biol 20:(6)599-604.
- \*Brooks AL, Peters RF, Rollag MD. 1971b. Metaphase chromosome aberrations in Chinese hamster liver cells in vivo after single acute <sup>60</sup>Co exposure. Radiat Res 45:191-201.
- \*Brooks SC, Herman JS, Hornberger GM, et al. 1998. Biodegradation of cobalt-citrate complexes: Implications for cobalt mobility in groundwater. J Contam Hydrol 32:99-115.
- \*Bruce BW, McMahon PB. 1996. Shallow ground-water quality beneath a major urban center: Denver, Colorado, USA. J Hydrol 186:129-151.
- Bruckner-Tuderman L, Konig A, Schnyder UW. 1992. Patch test results of the dermatology clinic Zurich in 1989: Personal computer-aided statistical evaluation. Dermatology 184:29-33.
- \*Brugman L. 1988. Some peculiarities of the trace-metal distribution in Baltic waters and sediments. Mar Chem 23:425-440.
- \*Brune D, Kjaerheim A, Paulsen G, et al. 1980. Pulmonary deposition following inhalation of chromium-cobalt grinding dust in rats and distribution in other tissues. Scand J Dent Res 88:543-551.
- \*Bruner A. 1977. Immediate changes in estimated cardiac output and vascular resistance after <sup>60</sup>Co exposure in monkeys: Implications for performance decrement. Radiat Res 70:391-405.
- \*Bruni JE, Persaud TVN, Froese G, et al. 1994. Effect of *in utero* exposure to low dose ioinizing on development in the rat. Histol Histopathol 9:27-33.
- \*Brusseau ML, Zachara JM. 1993. Transport of Co<sup>2+</sup> in a physically and chemically heterogeneous porous medium. Environ Sci Technol 27:1937-1939.
- \*Bryan SE, Bright JE. 1973. Serum protein responses elicited by iron, cobalt and mercury. Toxicol Appl Pharmacol 26:109-117.
- \*Bucher JR, Elwell MR, Thomson MB, et al. 1990. Inhalation toxicity studies of cobalt sulfate in F344/N rats and B6C3F1 mice. Fundam Appl Toxicol 15:357-372.
- \*Bucher JR, Hailey JR, Roycroft JR, et al. 1999. Inhalation toxicity and carcinogenicity studies of cobalt sulfate. Toxicol Sci 49:56-67.

# COBALT 302 9. REFERENCES

- \*Buchholz BA, Landsberger S. 1995. Leaching dynamics studies of municipal solid waste incinerator ash. J Air Waste Manage Assoc 45:579-590.
- \*Buchter B, Davidoff B, Amacher MC, et al. 1989. Correlation of freundlich Kd and n retention parameters with soils and elements. Soil Sci 148(5):370-379.
- \*Budavari S. 1996. The Merck index. 12th edition. Merck and Co., Inc., 412-414.
- \*Bulinksi R, Kot A, Bloniarz J, et al. 1986. [Study on some trace elements in homemade food stuffs: Part VII. Lead, cadmium, zinc, copper, vanadium, and cobalt content in vegetables and fruits]. Bromatol Chem Toksykol 19:21-26. (Polish).
- Bunzl K, Schimmack W. 1989. Associations between the fluctuations of the distribution coefficients of Cs, Zn, Sr, Co, Cd, Ce, Ru, Tc and I in the upper two horizons of a podzol forest soil. Chemosphere 18:2109-2120.
- \*Burba P, Rocha J, Klockow D. 1994. Labile complexes of trace metals in aquatic humic substances: Investigations by means of an ion exchange-based flow procedure. Fresenius J Anal Chem 349:800-807.
- \*Burger J, Gochfeld M. 1988. Metals in tern eggs in a New Jersey estuary: A decade of change. Environ Monit Assess 11:127-135.
- Burke DH, Brooks JC, Ryan RP, et al. 1979. p-Chloroamphetamine antagonism of cobaltous chloride-induced hypothermia in mice. Eur J Pharmacol 60:241-243.
- Burke DH, Brooks JC, Treml SB. 1983. Cobaltous chloride-induced hypothermia in mice III: Effect of pretreatment with 5-hydroxytrypaminergic agents. J Pharm Sci 72(7):824-826.
- Burrows BA, Chalmers TC. 1990. Cesium-137/potassium-40 ratios in firewood ashes as a reflection of worldwide radioactive contamination of the environment. Ann NY Acad Sci 609:334-339.
- \*Burr G, Sinks TH. 1989. Health hazard evaluation--report no. HETA 85-295-1907. General Electric Carboloy Systems, Detroit, Michigan. Cincinnati, OH: U.S. Department of Health and Human Services, Public Health Service, Centers for Disease Control, National Institute for Occupational Safety and Health. NTIS PB89-121008.
- \*Burt C. 1966. The Genetic determination of differences in intelligence: A study of monozygotic twins reared together and apart.
- \*Byczkowski JZ, Gearhart JM, Fisher JW. 1994. "Occupational" exposure of infants to toxic chemicals via breast milk. Nutrition 10(1):43-48.
- Byrd JT, Lee KW, Lee DS, et al. 1990. The behavior of trace metals in the Geum Estuary, Korea. Estuaries 13(1):8-13.
- \*CA Air Resources Board. 2000. California Air Toxics Program. Toxic air contaminant identification program. <a href="http://www.arb.ca.gov/toxics/toxics.htm">http://www.arb.ca.gov/toxics/toxics.htm</a>. March 16, 2000.
- \*CA EPA. 2000. State of California Environmental Protection Agency. Chemicals known to cause cancer. <a href="http://www.oehha.ca.gov/prop65/prop65"><u>Http://www.oehha.ca.gov/prop65/prop65 list/newlist.html.</u></a> January 8, 2000.

\*Cahill DF, Harvey HD, McCurry DC, et al. 1972. Radiological surveys of Pearl Harbor, Hawaii, and environs. Radiation Data and Reports 13:323-334.

Caicedo A, Kungel M, Pujol R, et al. 1998. Glutamate-induced Co<sup>2+</sup> uptake in rat auditory brainstem neurons reveals developmental changes in Ca<sup>2+</sup> permeability of glutamate receptors. Eur J Neurosci 10:941-954.

\*Camean A, Lopez-Artiguez M, Roca I, et al. 1998. Determination of cobalt, manganese, and alcohol content in beers. J Food Prot 61(1):129-131.

Camner P, Johansson A. 1992. Reaction of alveolar macrophages to inhaled metal aerosols. Environ Health Perspect 97:185-188.

\*Camner P, Boman A, Johansson A, et al. 1993. Inhalation of cobalt by sensitized guinea pigs: Effects on the lungs. Br J Ind Med 50:753-757.

Capar SG, Cunningham WC. 2000. Element and radionuclide concentrations in food: FDA total diet study 1991-1996. J AOAC Int 83(1):157-177.

\*Capomazza C, Botta A. 1991. Cobalt chloride induces micronuclei in human lymphocytes. Med Sci Res 19:219-220.

Cardarelli J, Elliott L, Hornung R, et al. 1997. Proposed model for estimating dose to inhabitants of <sup>60</sup>Co contaminated buildings. Health Phys 72(3):351-360.

Carnes BA, Olshansky SJ, Grahn D. 1998. An interspecies prediction of the risk of radiation-induced mortality. Radiat Res 149:487-492.

\*Carvajal NJ, Zienius RH. 1986. Gas chromatographic analysis of trace metals isolated from aqueous solutions as diethyldithiocarbamates. J Chromatogr 355:107-116.

\*Carvalho FP. 1987. Comparative uptake from sea water and tissue distribution of <sup>60</sup>Co in marine mollusks. Health Phys 53(1):73-81.

\*Casarett and Doull. 1986. Toxicology: The basic science of poisons. 3rd ed. New York, NY: Macmillan Publishing Company, 56-57.

\*Cassidy RM, Elchuk S, McHugh JO. 1982. Determination of metals in groundwaters by trace enrichment and liquid chromatography. Anal Chem 54:727-731.

Castiglioni G, Carosso A, Manzoni S, et al. 1992. Results of routine patch testing of 834 patients in Turin. Contact Dermatitis 27:182-185.

Cavelier C, Foussereau J, Gille P, et al. 1989. Allergy to nickel or cobalt: tolerance to nickel and cobalt samples in man and in the guinea pig allergic or sensitized to these metals. Contact Dermatitis 21:72-78.

\*CDC. 2001. National Report on Human Exposure to Environmental Chemicals, National Health and Nutrition Examination Survey, 1999. NCEH Pub. No. 01-0164, March 2001. Center for Disease Control, National Center for Environmental Health.

CEA. 1985. Behavior of cesium 137, chromium 51, cobalt 60, manganese 54, sodium 22 and zinc 65 in simulated estuarine environments. Effects if suspended mineral particles and dissolved organic matters.

Saint Paul Les Durance, France: Commissariat A L'Energie Atomique, Centre D'Etudes Nucleaires De Cadarache. CEA-R-5319.

Centeno JA, Pestaner JP, Mullick FG, et al. 1996. An analytical comparison of cobalt cardiomyopathy and idiopathic dilated cardiomyopathy. Biol Trace Elem Res 55:21-30.

Cereda C, Redaelli ML, Canesi M, et al. 1994. Widia tool grinding: The importance of primary prevention measures in reducing occupational exposure to cobalt. Sci Total Environ 150:249-251.

Chadwick JK, Wilson HK, White MA. 1997. An investigation of occupational metal exposure in thermal spraying processes. Sci Total Environ 199:115-124.

Chang MC, Hunt DM. 1960. Effects of in vitro radiocobalt irradiation of rabbit ova on subsequent development in vivo with special reference to the irradiation of maternal organism. Anat Rec 137:511-519.

\*Chang MC, Hunt DM, Harvey EB. 1963. Effects of radiocobalt irradiation of pregnant rabbits on the development of fetuses. Anat Rec 145:455-466.

Chang MC, Hunt DM, Romanoff EB. 1957. Effects of radiocobalt irradiation of rabbit spermatozoa in vitro on fertilization and early development. Anat Rec 129:211-229.

Chang MG, Hunt DM, Romanoff EB. 1958. Effects of radiocobalt irradiation of unfertilized or fertilized rabbit ova in vitro on subsequent fertilization and development in vivo. Anat Rec 132:161-179.

\*Chang WP, Chan CC, Wang JD. 1997. <sup>60</sup>CO contamination in recycled steel resulting in elevated civilian radiation doses: Causes and challenges. Health Phys 73:(3)465-472.

\*Chang WP, Hwang JS, Hung MC, et al. 1999a. Chronic low-dose  $\gamma$ -radiation exposure and the alteration of the distribution of lymphocyte subpopulations in residents of radioactive buildings. Int J Radiat Biol 75(10):1231-1239.

\*Chang WP, Lin YP, Hwang PT, et al. 1999b. Persistent leukocyte abnormalities in children years after previous long-term low-dose radiation exposure. Br J Haematol 106:954-959.

Chang WP, Tsai M-S, Hwang J-S, et al. 1999c. Follow-up in the micronucleus frequencies and its subsets in human population with chronic low-dose  $\gamma$ -irradiation exposure. Mutat Res 428:99-105.

Chauncey DM, Hagan PL, Halpern SE, et al. 1978a. Distribution of <sup>137</sup>Cs, <sup>201</sup>T1, <sup>203</sup>Hg, <sup>203</sup>Pb and <sup>57</sup>Co in a rat hematoma model. Comparison with <sup>67</sup>Ga. Invest Radiol 13(1):40-45.

Chauncey DM, Hagan PL, Halpern SE, et al. 1978b. The distribution of cadmium-115m chloride, cobalt-57 bleomycin, iodine-125 human serum albumin, selenium-75 selenite and selenomethionine-75 in a rat hepatoma model. Eur J Nucl Med 3:243-248.

Chave TA, Warin AP. 1999. Allergic contact dermatitis from cobalt in a beauty product. Contact Dermatitis 41:236.

\*Cheam V, Li EX. 1988. Ion chromatographic determination of low level cadmium(II), cobalt(II) and manganese(II) in water. J Chromatogr 450:361-371.

Chesnokov AV, Fedin VI, Govorun AP, et al. 1997. Collimated detector technique for measuring a <sup>137</sup>Cs deposit in soil under a clean protected layer. Appl Radiat Isot 48(9):1265-1272.

\*Chester R, Berry AS, Murphy KJT. 1991. The distributions of particulate atmospheric trace metals and mineral aerosols over the Indian Ocean. Mar Chem 34:261-290.

\*Chetty KN, Rao DSVS, Drummond L, et al. 1979. Cobalt induced changes in immune response and adenosine triphosphatase activities in rats. J Environ Sci Health B 14(5):525-544.

Chiappino G. 1994. Hard metal disease: clinical aspects. Sci Total Environ 150:65-68.

Chiavarini S, Galletti M, Michetti I, et al. 1994. Environmental monitoring at Terra Nova Bay Station from 1989-1991. Int J Environ Anal Chem 55:331-340.

Chillrud SN, Bopp RF, Simpson HJ, et al. 1999. Twentieth century atmospheric metal fluxes into Central Park Lake, New York City. Environ Sci Technol 33(5):657-662.

Chin JH, Delorenzo RJ. 1985. Cobalt ion enhancement of 2-chloro[<sup>3</sup>H]adenosine binding to a novel class of adenosine receptors in brain: antagonism by calcium. Brain Res 348:381-386.

Chocholova L. 1976. Effect of diazepam on the electroencephalographic pattern and vigilance of unanaesthetized and curarized rats with a chronic cobalt-gelatin focus. Physiol Bohemoslov 25(2):129-137.

Christensen JM, Poulsen OM. 1994. A 1982-1992 surveillance programed on Danish pottery painters. Biological levels and health effects following exposure to soluble or insoluble cobalt compounds in cobalt blue dyes. Sci Total Environ 150:95-104.

\*Christensen JM, Poulsen OM, Thomsen M. 1993. A short-term cross-over study in oral administration of soluble and insoluble cobalt compounds: Sex differences in biological levels. Int Arch Occup Environ Health 65:233-240.

Christensen TH, Kjeldsen P, Albrechtsen HJ, et al. 1994. Attenuation of landfill leachate pollutants in aquifers. Crit Rev Environ Sci 24:119-202.

\*Cikrt M, Tich M. 1981. Biliary excretion of cobalt in rats. J Hyg Epidemiol Microbiol Immunol 25(4):364-368.

\*Cirla AM. 1994. Cobalt-related asthma: Clinical and immunological aspects. Sci Total Environ 150:85-94.

\*Clewell HJ III, Andersen ME. 1985. Risk assessment extrapolations and physiological modeling. Toxicol Ind Health 1(4):111-131.

\*Clifford D, Subramonian S, Sorg TJ. 1986. Removing dissolved inorganic contaminants from water. Environ Sci Technol 20(11):1072-1080.

\*Clyne N, Lins L-E, Pehrsson SK, et al. 1988. Distribution of cobalt in myocardium, skeletal muscle and serum in exposed and unexposed rats. Trace Elem Med 5(2):52-54.

Clyne N, Persson B, Havu N, et al. 1990a. The intracellular distribution of cobalt in exposed and unexposed rat myocardium. Scand J Clin Lab Invest 50:605-609.

Clyne N, Wibom R, Havu N, et al. 1990b. The effect of cobalt on mitochondrial ATP-production in the rat myocardium and skeletal muscle. Scand J Clin Lab Invest 50:153-159.

\*Coakley JP, Nagy E, Serodes JB. 1993. Spatial and vertical trends in sediment-phase contaminants in the upper estuary of the St. Lawrence River. Estuaries 16(3B):653-669.

\*Cobalt Development Institute. 2000. Cobalt in Catalysts and Chemicals. Batteries. <a href="http://www.cobaltdevinstitute.com/">http://www.cobaltdevinstitute.com/</a>. April 16, 2000.

Cockerham LG, Cerveny TJ, Hampton JD. 1986. Postradiaiton regional cerebral blood flow in primates. Aviat Space Environ Med June: 578-582.

Cockerham LG, Prell GD. 1989. Prenatal radiation risk to the brain. Neurotoxicology 10:467-474.

\*CO Dept Public Health and Environment. 2000. Air quality. Colorado Department of Public Health and Environment. <a href="http://www.cdphe.state.co.us/cdphereg.asp">http://www.cdphe.state.co.us/cdphereg.asp</a>. April 4, 2000.

Colasanti BK, Craig CR. 1992. Reduction of seizure frequency by clonazepam during cobalt experimental epilepsy. Brain Res Bull 28(2):329-331.

\*Coleman ME, Elder RS, Basu P. 1992. Trace metals in edible tissues of livestock and poultry. J AOAC Int 75(4):615-625.

\*Collecchi P, Esposito M, Brera S, et al. 1986. The distribution of arsenic and cobalt in patients with laryngeal carcinoma. J Appl Toxicol 6(4):287-289.

\*Collier CG, Bailey MR, Hodgson A. 1989. An interspecies comparison of the lung clearance of inhaled monodisperse cobalt oxide particles- part V: Lung clearance of inhaled cobalt oxide particles in hamsters, rats and guinea-pigs. J Aerosol Sci 20(2):233-247.

\*Collier CG, Hodgson A, Gray SA, et al. 1991. The lung clearance kinetics of <sup>57</sup>Co<sub>3</sub>O<sub>4</sub>. J Aerosol Sci 22(4):537-549.

\*Collins JF, Johanson WG, McCullough B, et al. 1978. Effects of compensatory lung growth in irradiation-induced regional pulmonary fibrosis in the baboon. Am Rev Respir Dis 117:1079-1089.

\*Comar and Davis. 1947. Cobalt metabolism studies III. Excretion and tissue distribution of radioactive cobalt administered to cattle. Archives of Biochem 12:257-266.

Conde-Salazar L, Guimaraens D, Villegas C, et al. 1995. Occupational allergic contact dermatitis in construction workers. Contact Dermatitis 33:226-230.

Conrad CH, Brooks WW, Ingwall JS, et al. 1984. Inhibition of hypoxic myocardial contracture by cobalt in the rat. J Mol Cell Cardiol 16:345-354.

Coombs M. 1996. Biological monitoring of cobalt oxide workers. Int Arch Occup Environ Health 68:511-512.

Coquerelle TM, Weibezahn KF, Lucke-Huhle C. 1987. Rejoining of double strand breaks in normal human and ataxia-telangiectasia fibroblasts after exposure to 60Co y-rays, 241Am a-particles or bleomycin. Int J Radiat Biol 51(2):209-218.

\*Corisco JAG, Carreiro MCV. 1999. Co-60 transfer from water to the freshwater planktonic algae *Selenastrum capricornutum* Prinz. In: Anagnostoopoulos P, Brebbia CA, ed. Water polution V: modeling, measuring, and prediction. Progress in water resources 1. Boston: WIT Press, 427-436.

\*Corrier DE, Mollenhauer HH, Clark DE, et al. 1985. Testicular degeneration and necrosis induced by dietary cobalt. Vet Pathol 22:610-616.

\*Costa M, Heck JD, Robison S. 1982. Selective phagocytosis of crystalline metal sulfide particles and DNA strand breaks as a mechanism for the induction of cellular transformation. Cancer Res 42:2757-2763.

\*Cotton FA, Wilkinson G. 1980. Advanced inorganic chemistry. 4th ed. New York: John Wiley & Sons

Courtois A. 1972. Motor phenomenology of cobalt experimental epileptic focus in the motor cortex of the cat during various stages of vigilance. Electroencephalogr Clin Neurophysiol 32:259-267.

\*Cox AB, Keng PC, Glass NL, et al. 1981. Effects of heavy ions on rabbit tissues: alopecia. Int J Radiat Biol 40(6):645-657.

Craig CR, Colasanti BK. 1992. Reduction of frequency of seizures by carbamazepine during cobalt experimental epilepsy in the rat. Pharmacol Biochem Behav 41:813-816.

Craig CR, Chiu P, Colasanti K. 1976. Effects of diphenylhydantoin and trimethadione on seizure activity during cobalt experimental epilepsy in the rat. Neuropharmacology 15:485-489.

\*Croudace IW, Cundy AB. 1995. Heavy metal and hydrocarbon pollution in recent sediments from Southampton water, Southern England: A geochemical and isotopic study. Environ Sci Technol 29:1288-1296.

Cugell DW. 1992. The hard metal diseases. Clinical Chest Medicine 13(2):269-279.

Cugell DW, Morgan WKC, Perkins DG, et al. 1990. The respiratory effects of cobalt. Arch Intern Med 150:177-183.

Cui J-Q, Xu G-L. 1989. Protection of experimental cobalt cardiomyopathy in the rat by selenium pretreatment. In: Wendel A, ed. Selenium in biology and medicine. Berlin: Springer-Verlag, 194-198.

Cummings KB, Taylor WJ, Correa RJ, et al. 1976. Observations on definitive cobalt 60 radiation for cure in bladder carcinoma: 15-year followup. J Urol 115:152-154.

\*Cunningham GR, Huckins C. 1978. Serum FSH, LH, and testosterone in <sup>60</sup>Co γ-irradiated male rats. Radiat Res 76:331-338.

\*Cushing CE, Watson DG, Scott AJ, et al. 1981. Decrease of radionuclides in Columbia River biota following closure of hanford reactors. Health Physics 41:59-67.

\*Cyr F, Mehra MC, Mallet VN. 1987. Leaching of chemical contaminants from a municipal landfill site. Bull Environ Contam Toxicol 38:775-782.

Czeizel AE, Hegedus S, Timar L. 1999. Congenital abnormalities and indicators of germinal mutations in the vicinity of an acrylonitrile producing factory. Mutat Res 427:105-123.

- \*Czyscinski KS, Pietrzak RF, Weiss AJ. 1982. Evaluation of isotope migration-land burial: Water chemistry at commercially operated low-level radioactive waste disposal sites. Nuclear Regulatory Commission, Office of Nuclear Regulatory Research, Washington, DC. NTIS/NUREG/CR-2124.
- \*Dabeka RW. 1989. Survey of lead, cadmium, cobalt and nickel in infant formulas and evaporated milks and estimation of dietary intakes of the elements by infants 1-12 months old. Sci Total Environ 89:279-289.
- \*Dabeka RW, McKenzie AD. 1995. Survey of lead, cadmium, fluoride, nickel, and cobalt in food composites and estimation of dietary intakes of these elements by Canadians in 1986-1988. J AOAC Int 78(4):897-909.
- D'Adda F, Borleri D, Migliori M, et al. 1994. Cardiac function study in hard metal workers. Sci Total Environ 150:179-186.
- \*Dalvi RR, Robbins TJ. 1978. Comparative studies on the effect of cadmium, cobalt, lead, and selenium on hepatic microsomal monooxygenase enzymes and glutathione levels in mice. J Environ Pathol Toxicol 1:601-607.
- Dameron GW, Beck ML, Maurer JK, et al. 1997. Early clinical chemistry changes associated with short-term exposure to cobalt in rats. Clin Chem 43:6.
- \*Darwezah N, Maruyama Y, Feola JM, et al. 1988. Six- and thirty-day LD50 data for acute Co-60, Cs-137, and Cf-252 in total body-irradiated BALB/C mice. Int J Radiat Oncol Biol Phys 15(Suppl. 1):252.
- \*Dasch JM, Wolff GT. 1989. Trace inorganic species in precipitation and their potential use in source apportionment studies. Water Air Soil Pollut 43:401-412.
- Dauvalter V. 1994. Heavy metals in lake sediments of the Kola Peninsula, Russia. Sci Total Environ 158:51-61.
- Davidson JS, Franco SE, Millar RP. 1993. Stimulation by Mn<sup>2+</sup> and inhibition by Cd<sup>2+</sup>, Zn2+, Ni<sup>2+</sup>, and Co<sup>2+</sup> ions of lutenizing hormone exocytosis at an intracellular site. Endocrinology 132(6):2654-2658.
- \*Davis JE. 1937. Cobalt polycythemia in the dog. Proc Soc Exp Biol Med 37:96-99.
- \*Davis JE, Fields JP. 1958. Experimental production of polycythemia in humans by administration of cobalt chloride. Proc Soc Exp Biol Med 99:493-495.
- Davis ME. 1982. Cobaltous chloride effects on hexachlorobutadiene (HCBD) nephrotoxicity. Fed Proc 41(4):1053.
- \*Davis SD, Yankelevitz DF, Henschke CI. 1992. Radiation effects on the lung: Clinical features, pathology, and imaging findings. AJR Am J Roentgenol 159:1157-1164.
- \*Davison AG, Haslam PL, Corrin B, et al. 1983. Interstitial lung disease and asthma in hard-metal wokers: bronchoalveolar lavage, ultrastructural, and analytical findings and results of bronchial provocation tests. Thorax 38:119-128.
- \*DE Air Quality Management. 2000. Chemicals and reportable quantities in pounds by CAS number. <a href="http://www.dnrex.state.de.us/air/aqmpage/regs.htm">http://www.dnrex.state.de.us/air/aqmpage/regs.htm</a>. April 10, 2000.

\*De Boeck M, Lison D, Kirsh Volders M. 1998. Evaluation of the in vitro direct and indirect genotoxic effects of cobalt compounds using the alkaline comet assay. Influence of interdonor and interexperimental variability. Carcinogenesis 19:2021-2129.

De Boeck M, Saaristo M, Van Goethem F, et al. 1997. Mutagenic and antimutagenic effects of cobalt compounds measured by the comet assay. Mutat Res 379:S129.

Decaestecker AM, Marez T, Jdaini J, et al. 1990. Hypersensitivity to dichromate among asymptomatic workers in a chromate pigment factory. Contact Dermatitis 23:52-53.

\*De Franceschi L, Gentilis A, Guidi P, et al. 1976. <sup>60</sup>Co in the marine mollusc, Pinna nobilis. Health Phys 31:376-377.

\*Deka NC, Sehgal AK, Chhuttani PN. 1981. Absorption and transport of radioactive <sup>57</sup>cobalt vitamin B<sub>12</sub> in experimental giardiasis in rats. Indian J Med Res 74:675-679.

De La Cuadra J, Grau-Massanes M. 1991. Occupational contact dermatitis from rhodium and cobalt. Contact Dermatitis 25:182-184.

De Matteis F, Gibbs AH. 1976. The effect of cobaltous on liver haem metabolism in the rat: Evidence for inhibition of haem synthesis and for increased haem degradation. Ann Clin Res 8(Suppl. 17):13-197.

\*De Matteis F, Gibbs AH. 1977. Inhibition of haem synthesis caused by cobalt in rat liver. Biochem J 162:213-216.

\*Demedts M, Gheysens B, Lauweryns J, et al. 1984a. "Hard-metal" lung disease due to cobalt in diamond polishers. Am Rev Respir Dis 129:A155.

\*Demedts M, Gheysens B, Nagels J, et al. 1984b. Cobalt lung in diamond polishers. Am Rev Respir Dis 130:130-135.

\*Deng JF, Sinks T, Elliott L, et al. 1991. Characterization of respiratory health and exposures are a sintered permanent magnet manufacturer. Br J Ind Med 48:609-615.

Desrosiers MF. 1991. In vivo assessment of radiation exposure. Health Phys 61(6):859-861.

Deur CJ, Stone MJ, Frenkel EP. 1981. Trace metals in hematopoiesis. Am J Hematol 11:309-331.

\*Devi UP, Baskar R, Hande MP. 1994. Effect of exposure to low-dose gamma radiation during the late organogenesis in the mouse fetus. Radiat Res 138:133-138.

\*Devi UP, Hossain M, Bisht KS. 1998. Effect of gamma radiation on the foetal haemopoietic system in the mouse. Int J Radiat Biol 74(5):639-646.

\*Devi U, Saini MR, Saharan BR, et al. 1979. Radioprotective effect of 2-mercaptopropionylglycine on the intestinal crypt of Swiss albino mice after cobalt-60 irradiation. Radiat Res 80:214-220.

Dewar AJ, Dow RC, McQueen JK. 1972. RNA and protein metabolism in cobalt-induced epileptogenic lesions in rat brain. Epilepsia 13:552-560.

\*Dick HLH, Saylor CB, Reeves MM, et al. 1979. Chronic cardiac arrhythmias produced by focused cobalt-60 gamma irradiation of the canine atria. Radiat Res 78:390-403.

- Diediker LP. 1999. Waste management and chemical inventories. Hartford site environmental report for calendar year 1998. PNNL-12088. <a href="http://www.hanford.gov/docs/annualrp98/index.htm.">http://www.hanford.gov/docs/annualrp98/index.htm.</a> June 8, 1999.
- \*Di Giulio C, Data PG, Lahiri S. 1991. Chronic cobalt causes hypertrophy of glomus cells in the rat carotid body. Am J Physiol 261:C102-C105.
- \*Di Guilio C, Huang WX, Lahiri S, et al. 1990. Cobalt stimulates carotid body chemoreceptors. J Appl Physiol 68(5):1844-1849.
- \*Dinehart SM, Anthony JL, Pollack SV. 1991. Basal cell carcinoma in young patients after irradiation for childhood malignancy. Med Pediatr Oncol 19:508-510.
- DOE. 1978. Prenatal cobalt-60 irradiation effects on early postnatal development of the squirrel monkey offspring. In: Developmental toxicology if energy-related pollutants; proceedings of the seventeenth annual Hanford Biology Symposium at Richland, Washington, October 17-19, 1977. U.S. Department of Energy. DOE symposium series 47.
- DOE. 1983. Long term lung retention after inhalation of cobalt-oxide and cobalt-nitrate aerosols. In: Current concepts in lung dosimetry: Proceedings of a special workshop. U.S. Department of Energy. PNL-SA 11049.
- DOE. 1988. Investigation of leaching of radionuclides and hazardous materials from low-level wastes at Oak Ridge National Laboratory. Washington, DC: U.S. Department of Energy. NTIS/DE87013363.
- \*DOE. 1991. Radioactive releases at the Savannah River site, 1954-1989. An environmental protection department summary. Washington, DC: U.S. Department of Energy. NTIS/DE92009983.
- \*DOE. 1995. National low-level waste management program radionuclide report series. Volume 12: Cobalt-60. U.S. Department of Energy. DOE/LLW-128.
- \*DOE. 1996. Evaluation of cobalt mobility in soils from the Nevada test site. Reno, NV: U.S. Department of Energy. DOE/NV/10845-58.
- \*DOE. 1999. Inventory and characteristics of spent nuclear fuel high level radioactive waste and other materials. U.S. Department of Energy. <a href="http://www.ymp.gov/deisdoc/Volume%2011/Appendix\_A.pdf">http://www.ymp.gov/deisdoc/Volume%2011/Appendix\_A.pdf</a>. January 18, 1999.
- \*DOE. 2000. Derived air concentrations (DAC), radiation standards inhalation. U.S. Department of Energy. Code of Federal Regulations. 10 CFR 835 Appendix A, C, E.
- \*Dolling JA, Boreham DR, Brown DL, et al. 1998. Modulation of radiation-induced strand break repair by cesplatin in mammalian cells. Int J Radiat Biol 74(1):61-69.
- \*Domingo JL. 1989. Cobalt in the environment and its toxicological implications. Rev Environ Contam Toxicol 108:105-132.
- Domingo JL. 1994. Metal-induced development toxicity in mammals: A review. J Toxicol Environ Health 42:123-141.
- \*Domingo JL, Llobet JM. 1984. Treatment of acute cobalt intoxication in rats with L-methionine. Rev Esp Fisiol 40:443-448.

\*Domingo JL, Llobet JM, Bernat R. 1984. A study of the effects of cobalt administered orally to rats. Arch Farmacol Toxicol 10:13-20.

\*Domingo JL, Llobet JM, Corbela J. 1983. The effects of EDTA in acute cobalt intoxication in rats. Toxicol Eur Res 5(6):251-255.

\*Domingo JL, Llobet JM, Corbella J. 1985a. The effect of L-histadine on acute cobalt intoxication in rats. Food Chem Toxicol 23:130-131.

\*Domingo JL, Paternain JL, Llobet JM, et al. 1985b. Effects of cobalt on postnatal development and late gestation in rats upon oral administration. Rev Esp Fisiol 41:293-298.

Dominiczak A, Clyde E, Bohr D. 1991. Cobalt contraction of vascular smooth muscle. FASEB J 5:A384.

\*Donaldson JD. 1986. Cobalt and cobalt compounds. In: Gerhartz W, Yamamoto YS, Campbell FT, et al., ed. Ullman's Encyclopedia of industrial chemistry. New York, NY: VCH, 281-313.

Donat JR, Bruland KW. 1988. Direct determination of dissolved cobalt and nickel in seawater by differential pulse cathodic stripping voltammetry preceded by adsorptive collection of cyclohexane-1,2-dione dioxime complexes. Anal Chem 60:240-244.

Dong ZZ, Chen P, Li X-Q. 1996. Neurobehavioral study of prenatal exposure to hyperthermia combined with irradiation in mice. Neurotoxicol Teratol 18(6):703-709.

Doody MM, Mandel JS, Boice JD. 1995. Employment practices and breast cancer among radiologic technologists. J Occup Environ Med 37(3):321-327.

\*Dooms-Goossens A, Ceuterick A, Vanmalaele N, et al. 1980. Follow-up study of patients with contact dermatitis caused by chromates, nickel, and cobalt. Dermatologica 160:249-260.

\*DOT. 2001a. U.S. Department of Transportation. 40CFR173.435. Activity values for radionuclides. Http://www.dot.gov. June 18, 2001.

\*DOT. 2001b. U.S. Department of Transportation. 40CFR172.101. Superfund reportable quantity. <a href="http://www.dot.gov"><u>Http://www.dot.gov.</u></a>. June 18, 2001.

\*Down JD, Easton DF, Steel GG. 1986. Repair in the mouse lung during low dose-rate irradiation. Radiother Oncol 6:29-42.

Dreizen S, Levy BM, Niedermeier W, et al. 1970. Comparative concentrations of selected trace metals in human and marmoset saliva. Arch Oral Biol 15:179-188.

Dressler RL, Storm GL, Tzilkowski WM, et al. 1986. Heavy metals in cottontail rabbits on mined lands treated with sewage sludge. J Environ Qual 15(3):278-281.

Drosselmeyer E, Muller HL, Pickering S. 1989. An interspecies comparison of the lung clearance of inhaled monodisperse cobalt oxide particles - part VII: Lung clearance of inhaled cobalt oxide particles in Sprague-Dawley rats. J Aerosol Sci 20(2):257-260.

\*Duckham JM, Lee HA. 1976a. Cobalt cardiomyopathy. Lancet 1:1350.

\*Duckham JM, Lee HA. 1976b. The treatment of refractory anaemia of chronic renal failure with cobalt chloride. Q J Med 178:277-294.

Dufresne A, Loosereewanich P, Armstrong B, et al. 1996. Inorganic particles in the lungs of five a luminum smelter workers with pleuro-pulmonary cancer. Am Ind Hyg Assoc J 57:370-375.

\*Duncan RE, Bennett DW, Evans AT, et al. 1977. Radiation-induced bladder tumors. J Urol 118:43-45.

Duncan WRH, Morrison ER, Garton GA. 1981. Effects of cobalt deficiency in pregnant and post-parturient ewes and their lambs. Br Med J 46:337-344.

\*Dzubay TG, Morosoff N, Whitaker GL, et al. 1988. Polymer film standards for x-ray fluorescence spectrometers. Journal of Trace and Microprobe Techniques 5(4):327-341.

Eaton RP. 1972. Cobalt chloride-induced hyperlipemia in the rat: Effects on intermediary metabolism. Am J Physiol 222(6):1550-1557.

\*Eaton RP, Pommer I. 1973. Glucagon secretion and activity in the cobalt chloride-treated rat. Am J Physiol 225:67-72.

\*Eckel WP, Jacob TA. 1988. Ambient levels of 24 dissolved metals in U.S. surface and ground waters. In: Proceedings of the 196th meeting of the American Chemical Society, Division of Environmental Chemistry. New York, NY: American Chemical Society, 317-372.

Edel J, Pozzi G, Sabbioni E, et al. 1994. Metabolic and toxicological studies on cobalt. Sci Total Environ 150:233-244.

Edmondson PW, Batchelor AL. 1971. Acute lethal responses of goats and sheep to bilateral or unilateral whole-body irradiation by gamma-rays and fission neutrons. Int J Radiat Biol 20(3):269-290.

\*Eisenbud M. 1987. Environmental Radioactivity. 3<sup>rd</sup> ed. New York: Academic Press, Inc.

Elinder CG. 1984. Health hazards from exposure to cobalt, with special reference to carcinogenic, mutagenic and teratogenic effects. Toxicol Environ Chem 7:251-256.

\*Ellenhorn MJ, Schonwald S, Ordog G, et al., eds. 1997. Medical toxicology: Diagnosis and treatment of human poisoning. 2<sup>nd</sup> edition. Baltimore, MD: Williams & Wilkins. 1682-1723.

Elliott JE, Scheuhammer AM. 1997. Heavy metal and metallothionein concentrations in seabirds from the pacific coast of Canada. Mar Pollut Bull 34(10):794-801.

Elliott WC, Koski J, Houghton DC, et al. 1982. Bis(2,3-dibromopropyl) phosphate nephrotoxicity: Effect of sex and CoCl2 pretreatment. Life Sci 32:1107-1117.

El-Sewedy SM, Abdel-Tawab GA, El-Zoghby SM, et al. 1974. Studies with tryptophan metabolites in vitro. Effect of zinc, manganese, copper and cobalt ions on kynurenine hydrolase and kynurenine aminotransferase in normal mouse liver. Biochem Pharmacol 23:2557-2565.

Emtestam L, Zetterquist H, Olerup O. 1993. HLA-DR, -DQ and -DP alleles in nickel, chromium, and/or cobalt-sensitive individuals: Genomic analysis based on restriction fragment length polymorphisms. J Invest Dermatol 100:271-274.

\*Endo A, Kano Y, Mihara K, et al. 1993. Alteration in the retinoblastoma gene associated with immortalization of human fibroblasts treated with <sup>60</sup>Co gamma rays. J Cancer Res Clin Oncol 119:522-526.

EPA. 1980. Prescribed procedures for measurement of radioactivity in drinking water. Cincinnati, OH: U.S. Environmental Protection Agency. EPA-600/4-80-032.

EPA. 1986. Broad scan analysis if the FY82 national human adipose tissue survey specimens volume I-executive summary: Final report. Washington, DC: U.S. Environmental Protection Agency. EPA-560/5-86-035.

EPA. 1987. Reference dose (RfD): Description and use in health risk assessments. Volume I, Appendix A: Integrated risk information system supportive documentation. Washington, DC: U.S. Environmental Protection Agency. EPA/600/8-86/032a.

EPA. 1988. Analysis of clean water act effluent guidelines: Pollutants. Summary of the chemicals regulated by industrial point source category. Environmental Protection Agency. Federal Register. 40 CFR Parts 400-475.

EPA. 1989a. Reportable quantity adjustments: Delisting of ammonium thiosulfate. Final rules. U.S. Environmental Protection Agency. Federal Register 54:33417. 40 CFR parts 116, 117 and 302.

EPA. 1989b. Reportable quantity adjustments: radionuclides. Final rules. U.S. Environmental Protection Agency. Federal Register 54:22524-22543. 40 CFR parts 202 and 355.

\*EPA. 1990. Interim methods for development of inhalation reference concentrations. Washington, DC: U.S. Environmental Protection Agency, Office of Health and Environmental Assessment, Office of Research and Development, Environmental Criteria and Assessment Office. EPA 600/8-90/066A.

EPA. 1994. State tribal and site identification center, NPL site narrative at listing. Http://www.epa.gov/superfund/sites/npl/nar1369.htm. May 29, 1994.

\*EPA. 1997a. Special report on environmental endocrine disruption: An effects assessment and analysis. Washington, DC: U.S. Environmental Protection Agency, Risk Assessment Forum. EPA/630/R-96/012.

\*EPA. 1997b. Health effects assessment summary tables, FY 1997 update. Washington, DC: U.S. Environmental Protection Agency, Risk Assessment Forum. EPA/540/R/97/036.

EPA. 1999a. Designation of hazardous substances. Code of Federal Regulations. 40 CFR 302.4.

EPA. 1999b. Table 6 - VHAP or potential concern. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 63 Subpart JJ.

EPA. 1999c. NPDES permit application testing requirements for organic toxic pollutants by industrial category for existing dischargers. U.S. Environmental Protection Agency. Code of Federal Regulations 40 CFR 122, Appendix D.

EPA. 1999d. Designation of hazardous substances. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 116.4.

- EPA. 1999e. Toxic chemical release reporting; Community right-to-know. Sub-part D Specific toxic chemical listings. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 372.65.
- EPA. 1999f. Health and safety data reporting: Scope and compliance. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 716.1.
- EPA. 1999g. Designation of hazardous substances and reportable quantities. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 302.4, 40 CFR 302.5.
- EPA. 1999h. Compliance procedures methods for determining compliance with subpart I. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 61, Appendix E.
- \*EPA. 2000. Drinking water standards and health advisories. U.S. Environmental Protection Agency. EPA 822-B-00-001.
- \*EPA. 2001a. Annual possession quantities. U.S. Environmental Protection Agency. 40CFR61. <u>Http://www.epa.gov/epacfr40/chapt-I.info/chi-toc.htm</u>. March 13, 2001.
- \*EPA. 2001b. BPT effluent limitations. U.S. Environmental Protection Agency. 40CFR415.652. <u>Http://www.epa.gov/epacfr40/chapt-I.info/chi-toc.htm</u>. March 13, 2001.
- \*EPA. 2001c. Community right-to-know, release reporting. U.S. Environmental Protection Agency. 40CFR372.65. Http://www.epa.gov/epacfr40/chapt-I.info/chi-toc.htm. February 22, 2001.
- \*EPA. 2001d. Groundwater monitoring. U.S. Environmental Protection Agency. 40CFR264. <u>Http://www.epa.gov/epacfr40/chapt-I.info/chi-toc.htm.</u> March 13, 2001.
- \*EPA. 2001e. Hazardous waste identification and listing. U.S. Environmental Protection Agency. 40CFR261.38. Http://www.epa.gov/epacfr40/chapt-I.info/chi-toc.htm. February 22, 2001.
- \*EPA. 2001f. Municipal solid waste landfills. U.S. Environmental Protection Agency. 40CFR258. Http://www.epa.gov/epacfr40/chapt-I.info/chi-toc.htm. February 22, 2001.
- \*EPA. 2001g. NPEDS permit application testing requirements. U.S. Environmental Protection Agency. 40CFR122. <a href="http://www.epa.gov/epacfr40/chapt-I.info/chi-toc.htm">http://www.epa.gov/epacfr40/chapt-I.info/chi-toc.htm</a>. March 23, 2001.
- \*EPA. 2001h. Reported quantity, cobalt compounds. U.S. Environmental Protection Agency. 40CFR302.4. Http://www.epa.gov/epacfr40/chapt-I.info/chi-toc.htm. March 23, 2001.
- \*EPA. 2001i. Superfund reportable quantities. U.S. Environmental Protection Agency. 40CFR302.4. Http://www.epa.gov/epacfr40/chapt-I.info/chi-toc.htm. March 13, 2001.
- \*EPA. 2001j. Test methods. U.S. Environmental Protection Agency. 40CFR61. <u>Http://www.epa.gov/epacfr40/chapt-I.info/chi-toc.htm.</u> February 22, 2001.
- \*EPA. 2001k. TSCA health and safety data reporting. U.S. Environmental Protection Agency. 40CFR716.120. <a href="http://www.epa.gov/epacfr40/chapt-I.info/chi-toc.htm">http://www.epa.gov/epacfr40/chapt-I.info/chi-toc.htm</a>. February 22, 2001.
- \*Erlandsson B, Ingemansson T, Mattsson S. 1983. Comparative studies of radionuclides from global fallout and local sources in ground level air and sewage sludge. Water Air Soil Pollut 20:331-346.

\*Esclapez M, Trottier S. 1989. Changes in GABA-immunoreactive cell density during motor focal epilepsy induced by cobalt in the rat. Exp Brain Res 76:369-385.

\*Evans GJ, Jervis RE. 1987. Hair as a bio-indicator: Limitations and complications in the interpretation of results. J Radioanal Nucl Chem 110(2):613-625.

\*Evans GJ, Tan PV. 1998. The fate elements in residential composters. Arch Environ Contam Toxicol 34:323-329.

\*Evans RD, Andrews D, Cornett RJ. 1988. Chemical fractionation and bioavailability of cobalt-60 to benthic deposit-feeders. Can J Fish Aquat Sci 45:228-236.

Facchini A, Maraldi NM, Bartoli S, et al. 1976. Changes in membrane receptors of B and T human lymphocytes exposed to <sup>60</sup>CO gamma rays. Radiat Res 68:339-348.

Fan Z, Hiraoka M. 1989. Depression of delayed outward K<sup>+</sup> current by Co<sup>2+</sup> in guinea pig ventricular myocytes. J Mol Cell Cardiol 21(Suppl. II):S55.

Farah SB. 1983. The in vivo effect of cobalt chloride on chromosomes. Rev Bras Genet 6(3):433-442.

Farquhar SJ. 1997. Self dosing with cobalt or selenium by farmers. N Z Med J 110:237.

Fatemi SH, Antosh M, Cullan GM, et al. 1985. Late ultrastructural effects of heavy ions and gamma irradiation in the gastrointestinal tract of the mouse. Virchows Arch B 48:325-340.

Fawade MM, Pawar SS. 1983. Effect of NiCl<sub>2</sub>, CoCl<sub>2</sub> & cycloheximide on microsomal drug metabolism & ALA-synthesis during thiodemeton toxicity. Indian J Exp Biol 21:343-346.

\*FDA. 1999. Ionizing radiation in animal feed and pet food. U.S. Food and Drug Administration. Code of Federal Regulations. 21 CFR 579.40.

\*FDA. 2000a. Drug products withdrawn or removed from the market for reasons of safety or effectiveness. U.S. Food and Drug Administration. Code of Federal Regulations. 21 CFR 216.24.

\*FDA. 2000b. Certain drugs accorded new drug status through rulemaking procedures. U.S. Food and Drug Administration. Code of Federal Regulations. 21 CFR 310.502.

\*FDA. 2000c. Sources of radiation used for inspection of food, for inspection of packaged food, and for controlling food processing. U.S. Food and Drug Administration. Code of Federal Regulations. 21 CFR 179.21.

\*FDA. 2000d. Requirements regarding certain radioactive drugs. U.S. Food and Drug Administration. Code of Federal Regulations. 21 CFR 310.503.

\*FDA. 2000e. OTC warning label. U.S. Food and Drug Administration. Code of Federal Regulations. 21 CFR 369.20. <a href="http://www.access.gpo.gov"><u>Http://www.access.gpo.gov.</u></a> March 13, 2001.

\*FDA. 2000f. Substances recognized as safe. U.S. Food and Drug Administration. Code of Federal Regulations. 21 CFR 582.20. <a href="http://www.access.gpo.gov"><u>Http://www.access.gpo.gov.</u></a> March 13, 2001.

\*FDA. 2000g. Prohibited use in human food. U.S. Food and Drug Administration. Code of Federal Regulations. 21 CFR 189.120. <a href="http://www.access.gpo.gov">http://www.access.gpo.gov</a>. March 13, 2001.

- \*FDRL. 1984a. Acute oral LD<sub>50</sub> study of cobalt sulphate lot no. S88336/A in Sprague-Dawley rats. FDRL study no. 8005D. Food and Drug Research Laboratories, Inc., Waverly, NY. April 11, 1984.
- \*FDRL. 1984b. Study of cobalt (II) carbonate tech gr. CoCo<sub>3</sub>, lot #030383 in Sprague-Dawley rats. Food and Drug Research Laboratories, Inc., Waverly, NY. April 12, 1984.
- \*FDRL. 1984c. Acute oral toxicity study of cobalt oxide tricobalt tetraoxide in Sprague-Dawley rats. Food and Drug Research Laboratories, Inc., Waverly, NY. April 5, 1984.
- \*FDRL. 1984d. Acute oral LD<sub>50</sub> study of cobalt-325 MESH t3N in Sprague-Dawley rats. FDRL study no. 8005B. Food and Drug Research Laboratories, Inc., Waverly, NY. April 11, 1984.
- \*FEDRIP. 2000. Federal Research In Progress Database. National Technical Information Service, Springfield, VA.
- \*Feinendegen LE, Henneberg P, Tislgar-Lentulis G. 1977. DNA strand breakage and repair in human kidney cells after exposure to incorporated iodine-125 and cobalt-60 γ-rays. Curr Top Radiat Res Q 12:436-452.
- Fenech M, Morley AA. 1989. Kinetochore detection in micronuclei: An alternative method for measuring chromosome loss. Mutagenesis 4(2):98-104.
- \*Feng MR, Rossi DT, Strenkoski C, et al. 1998. Disposition kinetics of cobalt mesoporphyrin in mouse, rat, monkey and dog. Xenobiotica 28(4):413-426.
- Feola J, Maruyama Y, Magura C, et al. 1986. Response of lymphoid organs to low dose rate Cf-252, Cs-137 and acute Co-60. Nucl Sci Appl 2:787-796.
- \*Ferdenzi P, Giaroli C, Mori P, et al. 1994. Cobalt powder sintering industry (stone cutting diamond wheels): A study of environmental-biological monitoring, workplace improvement and health surveillance. Sci Total Environ 150:245-248.
- \*Fergusson JE, Ryan DE. 1984. The elemental composition of street dust from large and small urban areas related to city type, source and particle size. Sci Total Environ 34:101-116.
- Fernandez MA, Martinez L, Segarra M, et al. 1992. Behavior of heavy metals in the combustion gases of urban waste incinerators. Environ Sci Technol 26(5):1040-1047.
- Fernandez-Turiel JL, Lopez-Soler A, Liorens JF, et al. 1995. Environmental monitoring using surface water, river sediments, and vegetation: A case study in the Famatina Range, La Rioja, NW Argentina. Environ Int 21(6):807-820.
- Ferrans VJ, Hibbs RG, Weilbaecher DG. 1964. Alcoholic cardiomyopathy: a histochemical and electron microscopic study. Am J Cardiol 13:106-107.
- Ferri F, Candela S, Bedogni L, et al. 1994. Exposure to cobalt in the welding process with stellite. Sci Total Environ 150:145-147.
- Feuer G, Roomi MW, Stuhne-Sekalec L, et al. 1985. Association between progesterone binding and cytochrome P-450 content of heoatic microsomes in the rat treated with cobalt-haem. Xenobiotica 15(5):407-412.

Fiedler H, Hoffman HD. 1970. [The action of nickel(II)-L-glutamate and of different cobalt complexes on the behavior of several lipid components in rabbits]. Acta Biol Med Ger 25:389-398.

\*Figueroa S, Gerstenhaber B, Welch L, et al. 1992. Hard metal interstitial pulmonary disease associated with a form of welding in a metals parts coating plant. Am J Ind Med 21:363-373.

\*Finney BP, Huh C-A. 1989. History of metal pollution in the southern California bight: An update. Environ Sci Technol 23:294-303.

\*Firriolo JM, Ayala-Fierro F, Snipes IG, et al. 1999. Absorption and disposition of cobalt naphthenate in rats after a single oral dose. J Toxicol Environ Health, Part A 58:383-395.

\*Fischer T, Rystedt I. 1983. Cobalt allergy in hard metal workers. Contact Dermatitis 9:115-121.

Fisher DR, Dunavant BG. 1978. Internal decontamination of radiocobalt. Health Phys 35(2):279-285.

Fisher GE, MacPherson A. 1991. Effect of cobalt deficiency in the pregnant ewe on reproductive performance and lamb viability. Res Vet Sci 50:319-327.

\*Fisher NS, Fowler SW, Boisson F, et al. 1999. Radionuclide bioconcentration factors and sediment partition coefficients in arctic seas subject to contamination from dumped nuclear wastes. Environ Sci Technol 33(12):1979-1982.

\*Fisher NS, Teyssie JL, Fowler SW, et al. 1996. Accumulation and retention of metals in mussels from food and water: A comparison under field and laboratory conditions. Environ Sci Technol 30:3232-3242.

Fishman MJ, Perryman GR, Schroder LJ, et al. 1986. Determination of trace metals in low ionic strength waters using Zeeman and Deuterium background correction for graphite furnace absorption spectrometry. J Assoc Off Anal Chem 69(4):704-708.

\*Fishman ML, Bean SC, Cogan DG. 1976. Optic atrophy following prophylactic chemotherapy and cranial radiation for acute lymphocytic leukemia. Am J Ophthalmol 82(4):571-576.

\*Flaten TP. 1991. A nation-wide survey of the chemical composition of drinking water in Norway. Sci Total Environ 102:35-73.

Fleet JC, Golemboski KA, Dietert RR, et al. 1990. Induction of hepatic metallothionen by intraperitoneal metal injection: an associated inflammatory response. Am J Physiol 258:G926-G933.

Flegal ARE, Smith GJ, Gill GA, et al. 1991. Dissolved trace element cycles in the San Francisco Bay estuary. Mar Chem 36:329-363.

\*Fomon SJ. 1966. Body composition of the infant: Part I: The male "reference infant". In: Falkner F, ed. Human development. Philadelphia, PA: WB Saunders, 239-246.

\*Fomon SJ, Haschke F, Ziegler EE, et al. 1982. Body composition of reference children from birth to age 10 years. Am J Clin Nutr 35:1169-1175.

\*Forbes RM, Cooper AR, Mitchell HH. 1954. On the occurrence of beryllium, boron, cobalt, and mercury in human tissues. J Biol Chem 209:857-865.

Fordham PJ, Gramshaw JW, Crews HM, et al. 1995. Element residues in food contact plastics and their migration into food stimulants, measured by inductively-coupled plasma-mass spectrometry. Food Addit Contam 12(5):651-669.

Forni A. 1994. Bronchoalveolar lavage in the diagnosis of hard metal disease. Sci Total Environ 150:69-76.

Fortoul TI, Osorio LS, Tovar AT, et al. 1996. Metals in lung tissue from autopsy cases in Mexico City residents: Comparison of cases from the 1950s and the 1980s. Environ Health Perspect 104(6):630-632.

\*Foster PP, Pearman I, Ramsden D. 1989. An interspecies comparison of the lung clearance of inhaled monodisperse cobalt oxide particles- part II: Lung clearance of inhaled cobalt oxide in man. J Aerosol Sci 20(2):189-204.

Fowler SW. 1986. Trace metal monitoring of palagic organisms from the open Mediterranean Sea. Environ Monit Assess 7:59-78.

Franchi A, Prens EP, Ferrara GB, et al. 1996. Allergy to cobalt is associated with the activation of cobalt-specific HLA-DR-restricted CD4+ T-cells. Euro Respir J 9(Suppl. 23):895.

Franchini I, Bocchi MC, Giaroli C, et al. 1994. Does occupational cobalt exposure determine early renal changes? Sci Total Environ 150:149-152.

Francis AJ, Dodge CJ. 1988. Anaerobic microbial dissolution of transition and heavy metal oxides. Appl Environ Microbiol 54(4):1009-1014.

Francis AJ, Dodge CJ. 1990. Anaerobic microbial remobilization of toxic metals coprecipitated with iron oxide. Environ Sci Technol 24:373-378.

\*Francis CW, Davis EC, Goyert JC. 1985. Plant uptake of trace elements from coal gasification ashes. J Environ Qual 14(4):561-569.

Frank R, Stonefield KI, Luyken H, et al. 1986. Survey of elemental contents in two organs of slaughtered bovine, porcine and avian specimens, Ontario, Canada 1980-83. Environ Monit Assess 6:259-265.

\*Freitas ACS, Guimaraes JRD, Gouvea VA, et al. 1988. Laboratory experiments on <sup>60</sup>CO bioaccumulation by tropical seaweeds. In: Seeliger U, de Lacerda LD, Patchineelam SR, ed. Metals in coastal environments of Latin America. Berlin, Germany: Springer-Verlag, 147-154.

Frias-Espericueta MG, Osuna-Lopez JI, Sandoval-Salazar G, et al. 1999. Distribution of trace metals in different tissues in the rock oyster crassostrea iridescens: Seasonal variation. Bull Environ Contam Toxicol 63:73-79.

Fried W, Kilbridge T. 1969. Effect of testosterone and of cobalt on erythroprotein production by anephric rats. J Lab Clin Med 74(4):623-629.

\*Friedman HA, Kelmers AD. 1988. Investigation of leaching of radionuclides and hazardous materials from low-level wastes at Oak Ridge National Laboratory. Department of Energy, Washington, DC. NTIS/DE87013363.

\*FSTRAC. 1995. Summary of state and federal drinking water standards and guidelines 1993-1995. Federal-State Toxicology and Risk Analysis Committee. U.S. Environmental Protection Agency.

\*FSTRAC. 1999. Summary of state and federal drinking water standards and guidelines 1998-1999. Federal-State Toxicology and Risk Analysis Committee. U.S. Environmental Protection Agency.

Fuge R, Laidlaw IMS, Perkins WT, et al. 1991. The influence of acidic mine and spoil drainage on water quality in the mid-Whales area. Environ Geochem Health 13(2):70-75.

\*Fukunaga M, Kurachi Y, Mizuguchi Y. 1982. Action of some metal ions on yeast chromosomes. Chem Pharm Bull 30(8):3017-3019.

Fuller CC, Harvey JW. 2000. Reactive uptake of trace metals in the hyporheic zone of a mining-contaminated stream, Pinal Creek, Arizona. Environ Sci Technol 34:1150-1155.

Furuno K, Suetsugu T, Sugihara N. 1996. Effects of metal ions on lipid peroxidation in cultured rat hepatocytes loaded with  $\alpha$ -linolenic acid. J Toxicol Environ Health 48:121-129.

Gagnon WF, Horton JL. 1979. Physical factors affecting absorbed dose to the skin from cobalt-60 gamma rays and 25-MV x rays. Med Physics 6(4):285-290.

Gallagher MJ, Alade PI, Dominiczak AF, et al. 1994. Cobalt contraction of vascular smooth muscle is calcium dependent. J Cardiovasc Pharmacol 24:293-297.

\*Gallorini M, Edel J, Pietra R, et al. 1994. Cobalt speciation in urine of hard metal workers. A study carried out by nuclear and radioanalytical techniques. Sci Total Environ 150:153-160.

\*Garcia-Silva J, Velasco-Benito JA, Pena-Penabad C, et al. 1996. Basal cell carcinoma in a girl after cobalt irradiation to the cranium for acute lymphoblastic leukemia: Case report and literature review. Pediatric Dermatology 13(1):54-57.

\*Garg AN, Weginwar RG, Chutke NL. 1993. Radiochemical neutron activation analysis of Fe, Co, Zn, Sb, and Se in biomedical and environmental samples. Sci Total Environ 139/140:421-430.

Garnham GW, Codd GA, Gadd GM. 1993. Uptake of cobalt and cesium by microalgal-and cyanobacterial-clay mixtures. Microb Ecol 25:71-82.

\*Gautier MA. 1983. Manual of analytical methods for radiobioassay, DOE report no. LA-9763-M (National Technical Information Services, Springfield, Virginia).

Gawkrodger DJ, Lewis FM. 1993. Isolated cobalt sensitivity in an etcher. Contact Dermatitis 28:46.

Genicot J-L. 1997. Room-temperature semiconductor detectors for in vivo monitoring of internal contamination. Environ Health Perspect Suppl 105(6):1423-1426.

\*Gennart J, Lauwerys R. 1990. Ventilatory function of workers exposed to cobalt and diamond containing dust. Int Arch Occup Environ Health 62:333-336.

Gennart JP, Baleux C, Verellen-Dumoulin C, et al. 1993. Increased sister chromatid exchanges and tumor markers in workers exposed to elemental chromium-, cobalt- and nickel-containing dusts. Mutat Res 299:55-61.

Gerhardsson L, Nordberg GF. 1993. Lung cancer in smelter workers - interactions of metals as indicated by tissue levels. Scand J Work Environ Health 19(Suppl. 1):90-94.

\*Gerhardsson L, Brune D, Nordberg GF, et al. 1988. Multielemental assay of tissues of deceased smelter workers and controls. Sci Total Environ 74:97-110.

\*Gerhardsson L, Wester PO, Nordberg GF, et al. 1984. Chromium, cobalt and lanthanum in lung, liver and kidney tissue from deceased workers. Sci Total Environ 37:233-246.

\*Gerritse RG, Vriesema R, Dalenberg JW, et al. 1982. Effect of sewage sludge on trace element mobility in soils. J Environ Qual 11(3):359-364.

Geuniche A, Viac J, Lizard G, et al. 1994. Effect of various metals on intercellular adhesion molecule-1 expression and tumor necrosis factor alpha production by normal human keratinocytes. Arch Dermatol Res 286:466-470.

\*Gheysens B, Auwerx J, Van den Eeckhout A, et al. 1985. Cobalt-induced bronchial asthma in diamond polishers. Chest 88:740-744.

\*Gibbs RJ. 1994. Metals in the sediments along the Hudson River Estuary. Environ Int 20(4):507-516.

\*Gilman JPW. 1962. Metal carcinogenesis: II. A study on the carcinogenic activity of cobalt, copper, iron, and nickel compounds. Cancer Res 22:158-162.

\*Gilman JPW, Ruckerbauer GM. 1962. Metal carcinogenesis: I. Observations on the carcinogenicity of a refinery dust, cobalt oxide, and colloidal thorium dioxide. Cancer Res 22:152-157.

\*Gilot-Delhalle J, Moutschen J, Garsou J. 1988. Induction of translocations in mouse spermatogonia after fractionated exposure to  $^{60}$ Co  $\gamma$ -rays. Mutat Res 207:29-31.

Giulio CD, Data PG, Lahiri S. 1991. Chronic cobalt causes hypertrophy of glomulus cells in the rat carotid body. Am J Physiol 261:C102-C105.

Giusti L, Yang Y-L, Hewitt CN, et al. 1993. The solubility and partitioning of atmospherically derived trace metals in artificial and natural waters: A review. Atmos Environ 27A(10):1567-1578.

\*Giwercman A, Carlsen E, Keiding N, et al. 1993. Evidence for increasing incidence of abnormalities of the human testis: A review. Environ Health Perspect Suppl 101(2):65-71.

Glasgow GP, Corrigan KW. 1995. Installation of <sup>60</sup>Co 100 cm source-to-axis distance teletherapy units in vaults designed for 80-cm units. Health Phys 68(3):411-415.

\*Glooschenko WA, Capocianco J, Coburn J, et al. 1981. Geochemical distribution of trace metals and organochlorine contaminants of a lake Ontario shoreline marsh. Water Air Soil Pollut 15:197-213.

Godleski JJ, Kreyling WG. 1990. Localization of cobalt in the matrix of airway cartilage. Am Rev Respir Dis 141:A525.

Goebeler M, Meinardus-Hager G, Roth J, et al. 1993. Nickel chloride and cobalt chloride, two common contact sensitizers, directly induce expression of intercellular adhesion molecule-1 (ICAM-1), vascular cell adhesion molecule-1 (VCAM-1), and endothelial leukocyte adhesion molecule (ELAM-1) by endothelial cells. J Invest Dermatol 100:759-765.

Goebeler M, Roth J, Brocker E-B, et al. 1995. Activation of nuclear factor-kB and gene expression in human endothelial cells by the common haptens nickel and cobalt. J Immunol 155:2459-2467.

\*Goh CL, Gan SL, Ngui SJ. 1986. Occupational dermatitis in a prefabrication construction factory. Contact Dermatitis 15:235-240.

\*Goldberg MA, Schneider TJ. 1994. Similarities between the oxygen-sensing mechanisms regulating the expression of vascular endothelial growth factor and erythropoietin. J Biol Chem 269(6):4355-4359.

\*Goldberg MA, Dunning SP, Bunn HF. 1988. Regulation of the erythropoietin gene: Evidence that the oxygen sensor is a heme protein. Science 242:1412-1415.

\*Goldfrank LR, Flomenbaum NE, Weisman RS, et al. 1990. Cobalt. In: Goldfrank LR, Flomenbaum NE, Weisman RS, et al., ed. Goldfrank's toxicologic emergencies. Norwalk, Connecticut: Appleton and Lange, 654-655.

\*Goldfrank, LR, Flomenbaum, NE, Lewin, NA, et al. eds. 1998. Toxicological emergencies. 6<sup>th</sup> edition. Connecticut: Appleton & Lange, 481t, 489, 490t, 1338-1339.

\*Goldner MG, Volk BW, Lazarus SS. 1952. The effect of cobaltous chloride on the blood sugar and alpha cells in the pancreatic islets of the rabbit. Metabolism 1:544-548.

\*Golomb D, Ryan D, Eby N, et al. 1997. Atmospheric deposition of toxics onto Massachusetts Bay--I. Metals. Atmos Environ 31(9):1349-1359.

Gomaa MA, Aziz A, El-Assaly FM, et al. 1983. Biologically and physically recorded doses after an accidental exposure to <sup>60</sup>Co-γ rays. Health Phys 44:409-411.

\*Gomez-de-Segura I, Grande AG, De Miguel E. 1998. Antiemetic effects of lerisetron in radiation-induced emesis in the dog. Acta Oncol 37:759-763.

Gonsior SJ, Sorci JJ, Zoellner MJ, et al. 1997. The effects of EDTA on metal solubilization in river sediment/water systems. J Environ Qual 26:957-966.

\*Goodwin DA, Meares CF. 1976. Radiolabeled antitumor agents. Seminars in Nuclear Medicine 6(4):389-396.

Gopfert T, Eckardt K-U, Gess B, et al. 1995. Cobalt exerts opposite effects on erythropoietin gene expression in rat hepatocytes in vivo and in vitro. Am J Physiol 269:R995-R1001.

\*Grahn D, Carnes BA, Farrington BH. 1988. Genetic injury in hybrid male mice exposed to low doses of <sup>60</sup>Co γ-rays or fission neutrons. Mutat Res 162:81-89.

\*Grahn D, Lee CH, Farrington BF. 1983. Interpretation of cytogenetic damage induced in the germ line of male mice exposed for over 1 year to <sup>239</sup>Pu alpha particles, fission neutrons, or <sup>60</sup>Co gamma rays. Radiat Res 95:566-583.

Grant FW. 1976. Chromogenic response of aqueous cobalt thiocyanate to lipophilic drugs. J Chromatogr 116:230-234.

- \*Greathouse DG, Craun GF. 1978. Cardiovascular disease study occurrence of inorganics in household tap water and relationships to cardiovascular mortality rates. In: Proceedings of the 12th annual conference on trace substances on environmental health. Columbia, MO: University of Missouri, 31-39.
- \*Greenberg DM, Copp DH, Cuthbertson EM. 1943. Studies in mineral metabolism with the aid of artificial radioactive isotopes: VII. The distribution and excretion, particularly by way of the bile, of iron, cobalt, and manganese. J Biol Chem 147:749-756.
- \*Gregus Z, Klaassen CD. 1986. Disposition of metals in rats: A comparative study of fecal, urinary, and biliary excretion and tissue distribution of eighteen metals. Toxicol Appl Pharmacol 85:24-38.
- \*Greig RA, Jones J. 1976. Nondestructive neutron activation analysis of marine organisms collected from ocean dump sites of the middle eastern United States. Arch Environ Contam Toxicol 4(4):420-434.
- Greig RA, Sennefelder G. 1985. Metals and PCB concentrations in mussels from Long Island Sound. Bull Environ Contam Toxicol 35:331-334.
- \*Grice HC, Goodman T, Munro IC, et al. 1969. Myocardial toxicity of cobalt in the rat. Ann Acad Sci NY 156:189-194.
- Griffin MO, Levere RD, Abraham NC. 1991. Differential effect of DMSO and cobalt chloride on gene expression during erythropropoiesis. Exp Hematol 19:486.
- \*Guieu C, Martin JM, Thomas AJ, et al. 1991. Atmospheric versus river inputs of metals to the Gulf of Lions. Total concentrations, partitioning and fluxes. Mar Pollut Bull 22(4):176-183.
- \*Gumgum B, Unlu E, Tez Z, et al. 1994. Heavy metal pollution in water, sediment and fish from the Tigris River in Turkey. Chemosphere 29(1):111-116.
- \*Gutenmann WH, Rutzke M, Kuntz HT, et al. 1994. Elements and polychlorinated biphenyls in sewage sludge of large cities in the United States. Chemosphere 28(4):725-728.
- Guzelian PS, Bissell DM. 1976. Effect of cobalt on synthesis of heme and cytochrome P-450 in the liver. J Biol Chem 251(14):4421-4427.
- \*Guzelian PS, Henry CJ, Olin SS, eds. 1992. Similarities and differences between children and adults: Implications for risk assessment. Washington, DC: International Life Sciences Institute Press.
- \*Haddad E, Winchester JF. 1990. Clinical management of poisoning and drug overdose, 2nd ed. Philadelphia, PA: W.B. Saunders Co., 1030.
- \*Haddad E, Zikovsky L. 1985. Determination of Al, As, Cr, Cs, Fe, Mn, Sb, Sc, W and Zn in the workroom air by instrumental neutron activation analysis. J Radioanal Nucl Chem 93(6):371-378.
- \*Haga Y, Clyne N, Hatori N, et al. 1996. Impaired myocardial function following chronic cobalt exposure in an isolated rat heart model. Trace Elem Electrolytes 13(2):69-74.
- \*Hakanson R, Lundquist I, Sundler F. 1974. elevated levels of insulin-like activity and 5-hydroxytryptamine in guinea pig pancreas following CoCl<sub>2</sub> treatment. Endocrinology 94:318-324.
- \*Hamilton BF, Benjamin SA, Angleton GM, et al. 1989. The effect of perinatal  $^{60}$ Co  $\gamma$  radiation on brain weight in beagles. Radiat Res 119:366-379.

- \*Hamilton EI. 1994. The geobiochemistry of cobalt. Sci Total Environ 150:7-39.
- \*Hamilton-Koch W, Snyder RD, Lavelle JM. 1986. Metal-induced DNA damage and repair in human diploid fibroblasts and Chinese hamster ovary cells. Chem Biol Interact 59:17-28.
- \*Hanford. 1999. Hanford site environmental report for calendar year 1998. Sec 2.5. Waste Management. PNNL-12088, Richland, WA: Pacific Northwest National Laboratory. Http://www.hanford.gov/docs/annualrp98/sec2.5. February 16, 1999.
- \*Hanks GE, Ainsworth EJ, Leong GF, et al. 1966. Injury accumulation and recovery in sheep exposed to protracted cobalt-60 gamma radiation. Radiat Res 29:211-221.
- \*Hanna RGM. 1992. The level of heavy metals in the Red Sea after 50 years. Sci Total Environ 125:417-448.
- \*Hansen HS, Rygard J, Engelholm SA. 1976. Clinical use of combined bleomycin and radiation therapy for head and neck tumors and testicular cancers. Bull Cancer 63(3):371-378.

Hanson WF, Grant W. 1974. Dose to the skin from cobalt-60 tangential chest wall therapy. Phys Med Biol 19(2):260-261.

\*Hansson H-C, Ekholm A-KP, Ross HB. 1988. Rainwater analysis: A comparison between proton-induced x-ray emission and graphite furnace atomic absorption spectroscopy. Environ Sci Technol 22:527-531.

\*Harding HE. 1950. Nores on the toxicology of cobalt metal. Brit J Ind Med 7:76-78.

Haritonidis S, Malea P. 1995. Seasonal and local variation of Cr, Ni and Co concentrations in *Ulva rigida* C. Agardh and *Enteromorpha linza* (Linnaeus) from Theremaikos Gulf, Greece. Environ Pollut 89(3):317-327.

Harmuth-Hoene AE, Schelenz R. 1980. Effect of dietary fiber on mineral absorption in growing rats. J Nutr 110:1774-1784.

\*Harp MJ, Scoular FI. 1952. Cobalt metabolism of young college women on self-selected diets. J Nutr 47:67-72.

Harrison RM, Jones M. 1995. The chemical composition of airborne particles in the UK atmosphere. Sci Total Environ 168:195-214.

Harrow JAC. 1976. Subcellular basis of the cardiotoxic effects of cobalt, nickel and manganese. Diss Abstr Int B 37(11):5541-5542.

- \*Hartman ER, Colasanti BK, Craig CR. 1974. Epileptogenic properties of cobalt and related metals applied directly to cerebral cortex of rat. Epilepsia 15:121-129.
- \*Hartung M, Schaller K-H, Brand E. 1982. On the question of the pathogenetic importance of cobalt for hard metal fibrosis of the lung. Int Arch Occup Environ Health 50:53-57.

Hartwig A. 1998. Carcinogenicity of metal compounds: Possible role of DNA repair inhibition. Toxicol Lett 102-103:235-239.

Hartwig A, Kasten U, Boakye-Dankwa K, et al. 1990. Uptake and genotoxicity of micromolar concentrations of cobalt chloride in mammalian cells. Toxicol Environ Chem 28:205-215.

Hartwig A, Schlepegrell R, Dally H, et al. 1996. Interaction of carcinogenic metal compounds with deoxyribonucleic acid repair processes. Ann Clin Lab Sci 26(1):31-38.

\*Hartwig A, Snyder RD, Schlepegrell R, et al. 1991. Modulation by Co(II) of UV-induced DNA repair, mutagenesis and sister-chromatid exchanges in mammalian cells. Mutat Res 248:177-185.

\*Harvey EB, Chang MC. 1962. Effects of radiocobalt irradiation of pregnant hamsters on the development of embryos. J Cell Comp Physiol 59:293-305.

\*Hasanen E, Lipponen M, Minkkinen P, et al. 1990. Element concentrations of aerosol samples from the Baltic Sea area. Chemosphere 21(3):339-347.

\*Hashimoto M, Mitsuyasu Y. 1967. Subacute and chronic histological changes in the irradiated bone marrow. Acta Pathol Jpn 17(3):328-329.

Hatori N, Pehrsson SK, Clyne N, et al. 1993. Acute exposure and oxygen radical scavengers in the rat myocardium. Biochem Biophys Acta 1181:257-260.

Hatta T, Ishimoto F, Shinohara H, et al. 1990. Interference of MNNG and cobalt in teratogenicity. Teratology 42:46A.

Hattori Y, Moriwaki A, Hayashi Y, et al. 1985. Regional difference in depolarization-elicited accumulation of cyclic amp in cobalt-induced epileptic cortex of the rat. Acta Med Okayama 39(6):489-492.

Hattori Y, Moriwaka A, Hayashi Y, et al. 1992. Increased responses to adenosine and 2-chloroadenosine of cyclic AMP-generating systems in the primary cortical region of cobalt-induced epilepsy in the rat. Jpn J Physiol 42:151-157.

\*Hattori Y, Moriwaki A, Hayashi Y, et al. 1993. Involvement of adenosine-sensitive cyclic AMP-generating systems in cobalt-induced epileptic activity in the rat. J Neurochem 61:2169-2174.

Haux F, Lasfargues G, Lauwerys R, et al. 1995. Lung toxicity of hard metal particles and production of interleukin-1, tumor necrosis factor-α, fibronectin, and cystatin-c by lung phagocytes. Toxicol Appl Pharmacol 132:53-62.

Hayward DG, Petreas MX, Winkler JJ, et al. 1996. Investigation of a wood treatment facility: Impact on an aquatic ecosystem in the San Joaquin River, Stockton, California. Arch Environ Contam Toxicol 30:30-39.

\*HazDat. 2001. Agency for Toxic Substances and Disease Registry (ATSDR), Atlanta, GA.

\*Heath JC. 1956. The production of malignant tumors by cobalt in the rat. Br J Cancer 10:668-673.

\*Heath JC. 1960. The histogenesis of malignant tumors induced by cobalt in the rat. Br J Cancer 15:478-482.

\*Heath JC, Daniel MR. 1962. The production of malignant tumors by cobalt in the rat: Intrathoracic tumors. Br J Cancer 1:473-478.

\*Heath JC, Webb M, Caffrey M. 1969. The interaction of carcinogenic metals with tissues and body fluids. Cobalt and horse serum. Br J Cancer 23:153-166.

\*Heaton RW, Rahn KA, Lowenthal DH. 1990. Determination of trace elements, including regional tracers, in Rhode Island precipitation. Atmos Environ 24A(1):147-153.

Heinrich R, Angerer J. 1984. Determination of cobalt in biological materials by voltammetry and electro thermal atomic absorption spectrometry. Int J Environ Anal Chem 16:305-314.

\*Hellou J, Fancey LL, Payne JF. 1992a. Concentrations of twenty-four elements in bluefin tuna, Thunnus thynnus from the northwest Atlantic. Chemosphere 24(2):211-218.

Hellou J, Warren WG, Payne JF, et al. 1992b. Heavy metals and other elements in three tissues of cod, Gadus morhua from the Northwest Atlantic. Mar Pollut Bull 24(9):452-458.

\*Helmers E, Schrems O. 1995. Wet deposition of metals to the tropical north and the south Atlantic ocean. Atmos Environ 29(18):2475-2484.

\*Henquin J-C, Lambert AE. 1975. Cobalt inhibition of insulin secretion and calcium uptake by isolated rat islets. Am J Physiol 228(6):1669-1677.

\*Henquin J-C, Schmeer W, Meissner HP. 1983. Forskolin, and activator of adenylate cyclase, increase Ca2+-dependent electrical activity induced by glucose in mouse pancreatic B cells. Endocrinology 112(6):2218-2220.

Henrichs K, Newhaus R, Roth W. 1997. The monitoring of potential incorporations of occupationally exposed workers in Germany: II. Monitoring intakes of employees servicing nuclear power plants. Kerntechnik 62(1):51-52.

\*Henshaw JM, Heithmar EM, Hinners TA. 1989. Inductively coupled plasma mass spectrometric determination of trace elements in surface waters subject to acidic deposition. Anal Chem 61:335-342.

Herndon BL, Jacob RA, McCann J. 1979. Physiological effects. In: Smith IC, Carson BL, ed. Trace elements in the environment. Ann Arbor, MI: Ann Arbor Science Publishers, 925-1075.

\*Hewitt PJ. 1988. Accumulation of metals in the tissues of occupationally exposed workers. Environ Geochem Health 10(3-4):113-116.

Hicks M, Wharton G, Murphy WR, et al. 1997. Assessing the sequence specificity in the binding of CO(III) to DNA via a thermodynamic approach. Biopolymers 42:549-559.

HI Dept of Health. 2000. Environmental health: Clean air rules. Air pollution controls. Hawaii Department of Health. Http://www.hawaii.gov/doh/rules/emd/carule.htm. June 18, 2000.

Hildebrand HF, Veron C, Martin P. 1989. Nickel, chromium, cobalt dental alloys and allergic reactions: an overview. Biomaterials 10:545-548.

Hilgertova J, Ostra A, Sonka J. 1975. Formiminoglutamate excretion in rats exposed to x-rays and <sup>60</sup>Co gamma radiation. J Nucl Biol Med 19(1):1-4.

\*Hillerdal G, Hartung M. 1983. Short communication on cobalt in tissues from hard metal workers. Int Arch Occup Environ Health 53:89-90.

- \*Hiraide M, Sakurai K, Mizuike A. 1984. Radiochemical separation of cobalt-60 in seawater using continuous-flow coprecipitation-flotation. Anal Chem 56:2851-2853.
- \*Hirobe T. 1994. Effects of y-irradiation on the yield of mid-ventral white spots in mice in different genetic backgrounds and at different times during development. Mutat Res 322:213-220.
- \*Hirobe T, Zhou X. 1990. Effects of  $\gamma$ -irradiation on the differentiation of mouse melanocytes in the hair follicles. Mutat Res 234:91-96.
- Hirose K. 1990. Chemical speciation of trace metals in seawater: Implication of particulate trace metals. Mar Chem 28:267-274.
- Hobel M, Maroske D, Wegener K, et al. 1972. Uber die toxische wirkung von CoCl<sub>2</sub>, Co[Co-EDTA] oder Na<sub>2</sub>[Co-EDTA] enthaltender aerosole auf die ratte und die verteilung von [Co-EDTA]-- in organen des meerschweinchens. Arch Int Pharmacodyn 198:213-222.
- \*Hocherman S, Reichenthal E. 1983. Induction of semichronic epileptic foci using cobalt oxide. Surg Neurol 20:417-421.
- \*Hodge FG. 1993. Cobalt and cobalt alloys. In: Kroschwitz JI, Howe-Grant M, ed. Kirk-Othmer Encyclopedia of chemical technology. New York, NY: John Wiley & Sons, 760-777.
- \*Hoel DG, Davis DL, Miller AB, et al. 1992. Trends in cancer mortality in 15 industrialized countries, 1969-1986. J Natl Cancer Inst 84(5):313-320.
- Hoffman P, Dedik AN, Deutsch F, et al. 1997. Solubility of single chemical compounds from an atmospheric aerosol in pure water. Atmos Environ 31(17):2777-2785.
- \*Holcombe LJ, Eynon BP, Switzer P. 1985. Variability of elemental concentrations in power plant ash. Environ Sci Technol 19:615-620.
- \*Hollins JG, McCullough RS. 1971. Radiation dosimetry of internal contamination by inorganic compounds of cobalt: An analysis of cobalt metabolism in rats. Health Phys 21:233-246.
- \*Holly RG. 1955. Studies on iron and cobalt metabolism. JAMA, 158:1349-1352.
- Honda K, Nasu T, Tatsukawa R. 1984. Metal distribution in the earthworm, Pheretima hilgendorfi, and their variations with growth. Arch Environ Contam Toxicol 13:427-432.
- Horn EM, Dilloin GH, Fan Y-P, et al. 1999. Developmental aspects and mechanisms of rat caudal hypothalamic neuronal responses to hypoxia. Journal of Neurophysiology 81:1949-1959.
- \*Horowitz SF, Fischbein A, Matza D, et al. 1988. Evaluation of right and left ventricle function in hard metal workers. Brit J Ind Med 45:742-746.
- Horvath Z, Laszitty A, Varga I. 1992. The role of spectrochemicalkanalysis in the determination of the composition of atmospheric precipitation and aerosol samples in remote environments. Microchem J 46:130-135
- \*Horwitz C, Van Der Linden SE. 1974. Cadmium and cobalt in tea and coffee and their relationship to cardiovascular disease. S Afr Med J 48:230-233.

\*Houk AEH, Thomas AW, Sherman HC. 1946. Some interrelationships of dietary iron, copper and cobalt in metabolism. J Nutr 31:609-620.

\*House RA, Sax SE, Rumack ER, et al. 1992. Medical management of three workers following a radiation exposure incident. Am J Ind Med 22:249-257.

Howie DW, Rogers SD, McGee MA, et al. 1996. Biological effects of cobalt chrome in cell and animal models. Clin Orthop Relat Res 329S:S217-S232.

HSDB. 1989. Hazardous Substance Data Bank. National Library of Medicine, National Toxicology Information Program, Bethesda, MD.

\*HSDB. 2001. Hazardous Substance Data Bank. National Library of Medicine, National Toxicology Information Program, Bethesda MD.

HSE. 1996. Cobalt and cobalt compounds in air. Methods for the determination of hazardous substances 30/2. Sudbury, UK: Health and Safety Executives.

Huang C-Y, Lee J-D, Tseng C-L, et al. 1994. A rapid method for the determination of <sup>137</sup>Cs in environmental water samples. Anal Chim Acta 294:221-226.

Huck DW. 1976. The study of cobalt toxicity in pigs and rats. Diss Abstr Int B 37(1):159.

Huy ND, Morin PJ, Mohiuddin SM, et al. 1973. Acute effects of cobalt on cardiac metabolism and mechanical performance. Can J Physiol Pharmacol 51(1):46-51.

\*IAEA. 1962. Whole-body counting, International Atomic Energy Agency. Vienna: IAEA Publication No. STI/PUB/47.

\*IAEA. 1970. Directory of whole-body radioactivity monitors, International Atomic Energy Agency. Vienna: IAEA Publication No. STI/PUB/213.

\*IAEA. 1972. Assessment of radioactive contamination in man, International Atomic Energy Agency. Vienna: IAEA Publication No. STI/PUB/290.

\*IAEA. 1976. Diagnosis and treatment of incorporated radionuclides, International Atomic Energy Agency. Vienna: IAEA Publication No. STI/PUB/411.

\*IAEA. 1985. Assessment of radioactive contamination in man 1984, International Atomic Energy Agency. Vienna: IAEA Publication No. STI/PUB/674.

IAEA. 1988. The radiological accident in Goiania. International Atomic Energy Agency. Vienna: IAEA Publication No. STI/PUB/815.

\*IARC. 1991. IARC monographs on the evaluation of carcinogenic risks to humans. Vol. 52: Chlorinated drinking-water; chlorination by-products; some other halogenated compounds; cobalt and cobalt compounds. World Health Organization, Lyon, France.

\*IARC. 2000. Cobalt. International Agency for Research on Cancer. <a href="http://193.51.164.11/htdocs/Directory/index.html">http://193.51.164.11/htdocs/Directory/index.html</a>. February 17, 2000.

- \*IARC. 2001a. Carcinogenicity classification Group 2B, cobalt and cobalt compounds. International Agency for Research on Cancer. <a href="http://www.iarc.fr/pageroot/top1.html"><u>Http://www.iarc.fr/pageroot/top1.html</u></a>. June 7, 2001.
- \*IARC. 2001b. Some internally deposited radionuclides. International Agency for Research on Cancer. <a href="http://193.51.164.11/htdocs/Monographs/Vol78/Vol78-radionuclides.html">http://193.51.164.11/htdocs/Monographs/Vol78/Vol78-radionuclides.html</a>. June 7, 2001.
- \*Ichikawa Y, Kusaka Y, Goto S. 1985. Biological monitoring of cobalt concentrations in blood and urine. Int Arch Occup Environ Health 55:269-276.
- Ichikawa Y, Kusaka Y, Ogawa Y, et al. 1988. Changes of blood and urinary levels of cobalt during single exposure to cobalt. Jpn J Ind Health 30(3):208-209.
- \*ICRP. 1979. Limits for intakes of radionuclides by workers. International Commission of Radiological Protection. ICRP Publication 30. New York: Pergamon Press.
- \*ICRP. 1983. Radionuclide transformations: Energy and intensity of emissions. The International Commission on Radiological Protection. ICRP publication 30. New York, NY: Pergamon Press, 54-66.
- \*ICRP. 1991. 1990 Recommendations of the International Commission on Radiological Protection. International Commission on Radiological Protection.
- \*ICRP. 1993. Age-dependent doses to members of the public from intake of radionuclides: Part 2 ingestion dose coefficients. The International Commission on Radiological Protection. ICRP publication 67. New York, NY: Pergamon Press.
- \*ICRP. 1994. Age-dependent doses to members of the public from intake of radionuclides: Part 2, ingestion dose coefficients. The International Commission on Radiological Protection. ICRP publication 67. New York, NY: Pergamon Press.
- \*ICRP. 1995. Age-dependent doses to members of the public from intake of radionuclides: Part 4 ingestion dose coefficients. The International Commission on Radiological Protection. ICRP publication 71. New York, NY: Pergamon Press.
- \*ID Dept of Environmental Quality 2000. Air pollution control. Idaho Department of Environmental Quality. <a href="http://www2.state.id.us/adm/adminrules/rules/IDAPA58/58INDEX.htm">http://www2.state.id.us/adm/adminrules/rules/IDAPA58/58INDEX.htm</a>. March 13, 2000.
- Igarashi J, Hayashi N, Kikuchi G. 1978. Effects of administration of cobalt chloride and cobalt proroporphyrin on σ-aminolevulinate synthesis in rat liver. J Biochem 84:997-1000.
- \*Ikarashi Y, Ohno K, Tsuchiya T, et al. 1992a. Differences of draining lymph node cell proliferation among mice, rats and guinea pigs following exposure to metal allergens. Toxicology 76:283-292.
- \*Ikarashi Y, Tsuchiya T, Nakamura A. 1992b. Detection of contact sensitivity of metal salts using the murine local lymph node assay. Toxicol Lett 62:53-61.
- \*IL EPA. 2000a. Toxic air contaminants. Illinois Pollution Control Board. Illinois Environmental Protection Agency. <a href="http://www.ipcb.state.il.us/title35/35conten.html">http://www.ipcb.state.il.us/title35/35conten.html</a>. June 12, 2000.
- \*IL EPA. 2000b. Ground water quality. Illinois Pollution Control Board. Illinois Environmental Protection Agency. <a href="http://www.ipcb.state.il.us/title35/35conten.html">http://www.ipcb.state.il.us/title35/35conten.html</a>. June 12, 2000.

- \*IL EPA. 2000c. Radiation hazards. Illinois Pollution Control Board. Illinois Environmental Protection Agency. <a href="http://www.ipcb.state.il.us/title35/35conten.html">http://www.ipcb.state.il.us/title35/35conten.html</a>. June 12, 2000.
- \*Imbrogno P, Alborghetti F. 1994. Evaluation and comparison of the levels of occupational exposure to cobalt during dry and/or wet hard metal sharpening. environmental and biological monitoring. Sci Total Environ 150:259-262.
- \*Inaba J, Suzuki-Yasumoto M. 1979. A kinetic study of radionuclide absorption through damaged and undamaged skin of the guinea pig. Health Phys 37(4):592-595.
- \*Inaba J, Nishimura Y, Ichikawa R. 1980. Comparative metabolism of <sup>54</sup>Mn, <sup>59</sup>Fe, <sup>60</sup>Co and <sup>65</sup>Zn incorporated into Chlorella and in inorganic form in rats. Health Phys 39:611-617.
- \*Inano H, Ishii-Ohba H, Suzuki K, et al. 1990. Reasons for reduced activities of  $17\chi$ -hydrolase and  $C_{17}$ - $C_{20}$  lyase in spite of increased contents of cytochrome P-450 in mature rat testis fatally irradiated with  $^{60}$ Co. J Steroid Biochem 35(6):711-714.
- Inano H, Suzuki K, Ishii-Ohba H, et al. 1989. Steroid hormone production on testis, ovary, and adrenal gland of immature rats irradiated in utero with  $^{60}$ Co. Radiat Res 117:293-303.
- \*INEL. 2000. Isotope report. National Low-Level Waste Management Program, Idaho National Environmental Laboratory. Manifest Information Management System (MIMS). <a href="http://mims.inel.gov"><u>Http://mims.inel.gov</u></a>. June 12, 2000.
- \*Inoue T, Ohta Y, Sadaie Y, et al. 1981. Effect of cobaltous chloride on spontaneous mutation induction in a Bacillus subtilis mutator strain. Mutat Res 91:41-45.
- \*Institute of Medicine. 2000. Dietary reference intakes for thiamine, riboflavin, niacin, vitamin B<sub>6</sub>, folate, vitamin B<sub>12</sub>, pantothenic acid, biotin and choline. Washington DC: National Academy Press, 306-356. <a href="http://www.nap.edu/books/0309065542/html/index.html">http://www.nap.edu/books/0309065542/html/index.html</a>. June 25, 2000.
- \*IRIS. 2000. Cobalt. Integrated Risk Information System. <a href="http://www.epa.gov/iris/subst/index.htm"><u>Http://www.epa.gov/iris/subst/index.htm</u></a>. April 6, 2000.
- \*IRIS. 2001. Cobalt. Integrated Risk Information System. U.S. Environmental Protection Agency. <a href="http://www.epa.gov/iris/subst/index.htm">http://www.epa.gov/iris/subst/index.htm</a>. April 3, 2001.
- Isaacs RD, Wattie WJ, Wells AU, et al. 1987. Massive haemoptysis as a late consequence of pulmonary irradiation. Thorax 42:77-78.
- \*Ishihara N, Koizumi M, Yoshida A. 1987. Metal concentrations in human pancreatic juice. Arch Env Health 42(6):356-360.
- Isom GE, Way JL. 1974. Alteration of in vivo glucose metabolism by cobaltous chloride. Toxicol Appl Pharmacol 27:131-139.
- \*Jacobziner H, Raybin HW. 1961. Poison control... accidental cobalt poisoning. Arch Pediatr 78:200-205.
- Jagadeesan V, Sivaramakrishnan VM. 1969. Fate of cobalt-60 1-nitroso 2-naphthol chelate in albino rats after intravenous administration. Indian J Exp Biol 7:217-220.

- \*Jansen HML, Knollema S, van der Duin LV, et al. 1996. Pharmacokinetics and dosimetry of cobalt-55 and cobalt-57. J Nucl Med 37(12):2082-2086.
- \*Jarvis JQ, Hammond E, Meier R, et al. 1992. Cobalt cardiomyopathy: A report of two cases from mineral assay laboratories and a review of the literature. J Occup Med 34(6):620-626.
- \*Jenkins DW. 1980. Biological monitoring of toxic trace metals: Volume 1. Biological monitoring and surveillance. NTIS PB81-103475.
- Jensen AA, Tuchsen F. 1990. Cobalt exposure and cancer risk. Crit Rev Toxicol 20:427-437.
- Jimenez JS, Benitez MJ, Lechuga CG, et al. 1995. Casein kinase 2 inactivation by Mh<sup>2+</sup>, Mn<sup>2+</sup> and Co<sup>2+</sup> ions. Mol Cell Biochem 152:1-6.
- Johansen OJ, Carlson DA. 1976. characterization of sanitary landfill leachates. Water Res 10:1129-1134.
- \*Johanson CE. 1980. Permeability and vascularity of the developing brain: Cerebellum vs cerebral cortex. Brain Res 190:3-16.
- \*Johansson A, Curstedt T, Camner P. 1991. Lung lesions after combined inhalation of cobalt and nickel. Environ Res 54:24-38.
- \*Johansson A, Curstedt T, Rasool O, et al. 1992. Rabbit lung after combined exposure to soluble cobalt and trivalent chromium. Environ Res 58:80-96.
- \*Johansson A, Curstedt T, Robertson B, et al. 1984. Lung morphology and phospholipids after experimental inhalation of soluble cadmium, copper, and cobalt. Environ Res 34:295-309.
- \*Johansson A, Robertson B, Camner P. 1987. Nodular accumulation of type II cells and inflammatory lesions caused by inhalation of low cobalt concentrations. Environ Res 43:227-243.
- \*Jones P, Williams T, Ebdon L. 1989. Determination of cobalt at picogram levels by high-performance liquid chromatography with chemiluminescence detection. Anal Chim Acta 217:157-163.
- Jones WA, Miller EV, Sullivan LD, et al. 1980. RE: Severe prostatic calcification after radiation therapy for cancer. J Urol 123:135-136.
- \*Jordan C, Whitman RD, Harbut M, et al. 1990. Memory deficits in workers suffering from hard metal disease. Toxicol Lett 54:241-243.
- \*Jorhem L, Sundstrom B. 1993. Levels of lead, cadmium, zinc, copper, nickel, chromium, manganese, and cobalt in foods on the Swedish market, 1983-1990. J Food Comp Anal 6:223-241.
- Joseph MH, Emson PC. 1976. Taurine and cobalt induced epilepsy in the rat: A biochemical and electrocorticographic study. J Neurochem 27:1495-1501.
- \*Juraskova V, Drasil V. 1987. The level of chromosome aberrations and sister chromatid exchanges in continuously irradiated LS/BL lymphosarcoma cells. Studia Biophys 118:125-134.

\*Kada T, Shirasu Y, Ikekawa N, et al. 1986. Detection of natural bio-antimutagens and in vivo and in vitro analysis of their action. In: Genetic toxicology of environmental chemicals, part A: Basic principles and mechanisms of action: Alan Liss, Inc.

\*Kadiiska MB, Maples KR, Mason RP. 1989. A comparison of cobalt(II) and iron(II) hydroxyl and superoxide free radical formation. Arch Biochem Biophys 275(1):98-111.

Kahkonen MA, Suominen KP, Manninen PKG, et al. 1998. 100 years of sediment accumulation history of organic halogens and heavy metals in recipient and nonrecipient lakes of pulping industry in Finland. Environ Sci Technol 32(12):1741-1746.

\*Kakinuma J, Orii H. 1982. DNA interaction with <sup>57</sup>Co-bleomycin. Nucl Med 21:232-235.

Kamendulis LM, Jiang J, Xu Y, et al. 1999. Induction of oxidative stress and oxidative damage in rat glial cells by acrylonitrile. Carcinogenesis 20(8):1555-1560.

Kamiya K, Inoh A, Fujii Y, et al. 1985. High mammary carcinogenicity of neutron irradiation in rats and its promotion by prolactin. Jpn J Cancer Res 76:449-456.

\*Kanematsu N, Hara M, Kada T. 1980. Rec assay and mutagenicity studies on metal compounds. Mutat Res 77:109-116.

\*Kanerva L, Estlander T, Jolanki R. 1988. Occupational skin disease in Finland. Int Arch Occup Environ Health 60:89-94.

\*Kanerva L, Estlander T, Jolanki R. 1998. Bank clerk's occupational allergic nickel and cobalt contact dermatitis from coins. Contact Dermatitis 38:217-218.

\*Kapstad B. 1978. Treatment of squamous cell carcinomas of the head and neck region with cobalt and bleomycin. Int J Radiat Oncol Biol Phys 4:91-94.

\*Kapstad B. 1979. Cobalt and bleomycin against carcinomas of head and neck: A controlled clinical study. Acta Otolaryngol Suppl (Stockh) 360:171-173.

Karube Y, Iwamoto K, Miura J, et al. 1989. Radioactive metal complexes with affinity for tumors. II. Biodistribution of radioactivity in cellular and subcellular fractions of tumor tissues. Chem Pharm Bull 37(7):1874-1876.

\*Kasprzak KS, Zastawny TH, North SL, et al. 1994. Oxidative DNA base damage in renal, hepatic, and pulmonary chromatin of rats after intraperitoneal injection of cobalt(II) acetate. Chem Res Toxicol 7:329-335.

Kasten U, Hartwig A, Beyersmann D. 1992. Mechanisms of cobalt(II) uptake into V79 Chinese hamster cells. Arch Toxicol 66:592-597.

\*Kasten U, Mullenders LH, Hartwig A. 1997. Cobalt(II) inhibits the incision and the polymerization step of nucleotide excision repair in human fibroblasts. Mutat Res 383:81-90.

\*Katsarou A, Baxevanis C, Armenaka M, et al. 1997. Study of persistence and loss of patch test reactions to dichromate and cobalt. Contact Dermatitis 36:87-90.

Katsuoka Y, Beckman B, George WJ, et al. 1983. Increased levels of erythropoietin in kidney extracts of rats treated with cobalt and hypoxia. Am J Physiol 244(13):F129-F133.

Katz RP, George WJ, Anderson MB. 1988. Ultrastructural evaluation of the toxic effect of cobalt on the murine testis. Anat Rec 220(4):51A.

\*Kawakami Y, Koyama I. 1992. Changes in the strength of recurrent inhibition in cobalt-induced epilepsy. Epilepsia 33(3):428-434.

\*Kawanishi S, Inoue S, Yamamoto K. 1994. Active oxygen species in DNA damage induced by carcinogenic metal compounds. Environ Health Perspect Suppl 102(3):17-20.

Kawanishi S, Yamamoto K, Inoue S. 1989. Site-specific DNA damage induced by sulfite in the presence of cobalt(II) ion. Biochem Pharmacol 38(20):3491-3496.

Kawakami Y, Ishikawa T, Koyama I. 1990. Seizure elicited by VPL stimulation in cobalt induced epilepsy model. Jpn J Psychiatry Neurol 44(2):422-423.

Kazantzis G. 1981. Role of cobalt, iron, lead, manganese, mercury, platinum, selenium, and titanium in carcinogenesis. Environ Health Perspect 40:143-161.

\*Keener HA, Percival GP, Morrow KS, et al. 1949. Cobalt tolerance in young dairy cattle. J Dairy Sci 32:527-533.

Kempron S, Sterritt RM, Lester JN. 1987. Heavy metal removal in primary sedimentation II. The influence of metal speciation and particle size. Sci Total Environ 63:247-258.

Kempton S, Sterritt RM, Lester JN. 1987. Heavy metal removal in primary sedimentation I. The influence of metal solubility. Sci Total Environ 63:231-246.

Kent B, Spycher N. 1994. Major chemical parameters in groundwater control. In: Environmental science and pollution control. Groundwater contamination and control. New York, NY: Dekker, M, 479-495.

\*Kent NL, McCance RA. 1941. The absorption and excretion of 'minor' elements by man. Biochem J 35:877-883.

\*Kerfoot EJ. 1975. Semi-chronic inhalation study on cobalt. Diss Abstr Int B 35:6054-6055.

\*Keys HM, Reed W. 1980. Severe prostactic calcification after radiation therapy for cancer. J Urol 123:(1)135-1366.

\*Kharab P, Singh I. 1985. Genotoxic effects of potassium dichromate, sodium arsenite, cobalt chloride and lead nitrate in diploid yeast. Mutat Res 155:117-120.

Kiec-Swierczynska M. 1990a. Allergy to chromate, cobalt and nickel in Lodz 1977-1988. Contact Dermatitis 22:229-231.

Kiec-Swierczynska M. 1990b. Occupational dermatoses and allergy to metals in Polish construction workers manufacturing prefabricated building units. Contact Dermatitis 23:27-32.

\*Kiek-Swierczynska M, Krecisz B. 2000. Occupational skin diseases among the nurses in the region of Lodz. Int J Occup Med Environ Health 13(3):179-184.

Kilinc K, Rouhani R. 1992. Cobaltous ion inhibition of lipid peroxidation in biological membranes. Biochem Biophys Acta 1125:189-195.

\*Killey RWD, McHugh JO, Champ DR, et al. 1984. Subsurface cobalt-60 migration from a low-level waste disposal site. Environ Sci Technol 18:148-157.

\*Kim EY, Goto R, Tanabe S, et al. 1998a. Distribution of 14 elements in tissues and organs of oceanic seabirds. Arch Environ Contam Toxicol 35:638-645.

Kim SH, Chung CY, Son CH. 1998b. Cell death by apoptosis in the neonatal mouse cerebellum following gamma-irradiation. Anticancer Res 18:1629-1632.

Kimberly MM, Bailey GG, Paschal DC. 1987. Determination of urinary cobalt using matrix modification and graphite furnace atomic absorption spectrometry with Zeeman-effect background correction. Analyst 112:287-290.

\*Kincaid JF, Strong JS, Sunderman FW. 1954. Toxicity studies of cobalt carbonyls. Arch Ind Hyg Occup Med 10:210-212.

\*King GL. 1988a. Characterization of radiation-induced emesis in the ferret. Radiat Res 114:599-612.

\*King JN, Fritz JS. 1987. Determination of cobalt, copper, mercury, and nickel as bis(2-hydroxyethyl)dithiocarbamate by high-performance liquid chromatography. Anal Chem 59:703-708.

\*King LD. 1988b. Retention of metals by several soils of the southeastern United States. J Environ Qual 17(2):239-246.

\*Kinoshita K, Fujita T. 1972. Metabolism of <sup>57</sup>Co-methylcobalamin in rat and guinea pig. Chem Pharm Bull 20(12):2561-2569.

\*Kirchgessner M, Reuber S, Kreuzer M. 1994. Endogenous excretion and true absorption of cobalt as affected by the oral supply of cobalt. Biol Trace Elem Res 41:175-189.

Kitahara J, Yamanaka K, Kato K, et al. 1996. Mutagenicity of cobalt and reactive oxygen producers. Mutat Res 370:133-140.

\*Kitamori T, Suzuki K, Sawada T, et al. 1986. Determination of sub-part-per-trillion amounts of cobalt by extraction and photoacoustic spectroscopy. Anal Chem 58:2275-2278.

Klaassen CD, Amdur MO, Doull J. 1986. Casarett and Doull's toxicology: The basic science of poisons. 3rd ed. New York, NY: Macmillon Publishing Company.

Klavins M, Rodinov V, Vereskuns G. 1998. Metals and organochlorine compounds in fish from Latvian lakes. Bull Environ Contam Toxicol 60:538-545.

\*Klener V, Tuscany R, Velupkova J, et al. 1986. Long-term follow-up after accidental y irradiation from a 60 Co source. Health Phys 51(5):601-607.

\*Kloke A, Sauerbeck DR, Vetter H. 1984. The contamination of plants and soils with heavy metals and the transport of metals in terrestrial food chains. In: Nriagu JO, ed. Changing metal cycles and human health. Berlin Heidelberg: Springer-Verlag, 113-141.

\*Knauer GA, Martin JH, Gordon RM. 1982. Cobalt in north-east Pacific waters. Nature 297:49-51.

Knulst J, Sodergren A. 1994. Occurrence and toxicity of persistent pollutants in surface microlayers near an incinerator plant. Chemosphere 29(6):1339-1347.

\*Knutson AB, Klerks PL, Levinton JS. 1987. The fate of metal contaminated sediments in Foundry Cove, New York. Environ Pollut 45:291-304.

Kobayashi M, Shimizu S. 1999. Cobalt proteins. Eur J Biochem 261:1-9.

Koethals E, Obersztyn A, Dominikowski M. 1967. Pathological changes in the teeth and tooth appendages of the rat in radiation sickness. Pol Med J 6(5):1198-1205.

Kohlhardt M, Haap K. 1980. On the mechanism underlying the cobalt-induced inhibition of slow inward current in mammalian ventricular myocardium. J Mol Cell Cardiol 12:1075-1090.

Kohlhardt M, Bauer B, Krause H, et al. 1973. Selective inhibition of the transmembrane Ca conductivity of mammalian myocardial fibres by Ni, Co and Mn ions. Pflugers Arch 338:115-123.

\*Kokelj F, Daris F, Lutmann A, et al. 1994. Nickel, chromate and cobalt in toilet soaps analyzed by inductively coupled plasma mass spectrometry. Contact Dermatitis 31:270.

\*Koksal G, Dalci DO, Pala FS. 1996. Micronuclei in human lymphocytes: The Co-60 gamma-ray doseresponse. Mutat Res 359:151-157.

\*Koksal G, Pala FS, Dalci DO. 1995. In vitro dose-response curve for chromosome aberrations induced in human lymphocytes by <sup>60</sup>Co γ-radiation. Mutat Res 329:57-61.

Komeda H, Kobayashi M, Shimizu S. 1997. A novel transporter involved in cobalt uptake. Proc Natl Acad Sci U S A 94:36-41.

\*Komori M, Nishio K, Kitada M, et al. 1990. Fetus-specific expression of a form of cytochrome P-450 in human livers. Biochemistry 29:4430-4433.

\*Koponen M, Gustafsson T, Kalliomaki P-L. 1982. Cobalt in hard metal manufacturing dusts. Am Ind Hyg Assoc J 43(9):645-651.

Koyama I. 1992. A morphological study of the cortical pyramidal neuron in the cobalt-induced epileptogenic focus of the cat. Jpn J Psychiatry Neurol 46(2):351-352.

Koyama I, Ueda K, Sekino Y, et al. 1988. A morphological study comparing cortical neurons of focal epilepsy in humans with those of cobalt-induced focal epilepsy in cats. Jpn J Psychiatry Neurol 42(3):653-655.

Kozubek S, Krasavin EA, Amirtayev KG, et al. 1989. The induction of reverants by heavy particles and y-rays in salmonella tester strains. Mutat Res 210:221-226.

- \*Krasovskii GN, Fridlyand SA. 1971. Experimental data for the validation of the maximum permissible concentration of cobalt in water bodies. Hyg Sanit 26:277-279.
- \*Kratchler M, Rossipal SLE, Irgolic KJ. 1998. Changes in the concentrations of trace elements in human milk during lactation. J Trace Elements Med Biol 12:159-176.
- Kreyling WG, Cox C, Ferron GA, et al. 1993. Lung cancer in Long-Evans rats after inhalation of porous, monodisperse cobalt oxide particles. Exp Lung Res 19:445-467.
- Kreyling W, Ferron GA, Haider B. 1980. Analysis of the long term lung retention of cobalt oxide nitrate aerosols in dogs. In: Hochrainer D, ed. Aerosols in science, medicine and technology: Physical and chemical properties of aerosols. Schmallenberg, Germany: Gesellschaft für Aerosolforschung, 251-258.
- \*Kreyling WG, Ferron GA, Haider B. 1984a. The dependency of the lung retention on cobalt aerosol parameters. J Aerosol Sci 15(3):229-232.
- \*Kreyling WG, Ferron GA, Haider B. 1984b. Lung retention and clearance of cobalt oxide particles depending on their physicochemical parameters. EUR 9384:141-146.
- \*Kreyling W, Ferron G, Haider B, et al. 1985. Total and regional lung retention of monodisperse cobalt compound aerosols after a single inhalation. Z Erkr Atmungsorgane 164:60-66.
- \*Kreyling WG, Ferron GA, Haider B. 1986. Metabolic fate of inhaled Co aerosols in beagle dogs. Health Phys 51(6):773-795.
- Kreyling WG, Ferron GA, Haider B. 1989. An interspecies comparison of the lung clearance of inhaled monodisperse cobalt oxide particles- part IV: Lung clearance of inhaled cobalt oxide particles in beagle dogs. J Aerosol Sci 20(2):219-232.
- \*Kreyling WG, Godleski JJ, Kariya ST, et al. 1990. In vitro dissolution of uniform cobalt oxide particles by human and canine alveolar macrophages. Am J Resp Cell Mol Biol 2:413-422.
- \*Krishnan K, Andersen ME. 1994. Physiologically based pharmacokinetic modeling in toxicology. In: Hayes AW, ed. Principles and methods of toxicology. 3rd ed. New York, NY: Raven Press, Ltd., 149-188.
- \*Krishnan K, Andersen ME, Clewell HJ III, et al. 1994. Physiologically based pharmacokinetic modeling of chemical mixtures. In: Yang RSH, ed. Toxicology of chemical mixtures: Case studies, mechanisms, and novel approaches. San Diego, CA: Academic Press, 399-437.
- \*Kriss JP, Carnes WH, Ross RT. 1955. Hypothyroidism and thyroid hypoplasia in patients treated with cobalt. JAMA 157(2):117-121.
- KS Dept of Health and Environment. 2000. Ambient air quality standards and air pollution control. Rules and Regulations. Http://www.kdhe.state.ks.us/. May 16, 2000.
- \*Kumagai S, Kusaka Y, Goto S. 1996. Cobalt exposure level and variability in the hard metal industry of Japan. Am Ind Hyg Assoc J 67:365-369.
- Kumagai S, Kusaka Y, Goto S. 1997. Log-normality of distribution of occupational exposure concentrations to cobalt. Ann Occup Hyg 41(3):281-286.

Kumar GP, Laloraya M, Laloraya MM. 1990. Powerful anti-sperm motility action of cobaltous ions and its recovery by a sulfhydryl compound. Contraception 41(6):633-639.

Kureishy T, Gupta RS, Mesquita A, et al. 1993. Heavy metals in some parts of Antarctica and the southern Indian Ocean. Mar Pollut Bull 26(11):651-652.

Kurishita A, Ihara T. 1990. Inhibitory effects of cobalt chloride and cinnamaldehyde on 5-azacytidine-induced digital malformations in rats. Teratology 41:161-166.

\*Kuroda Y, Inoue T. 1988. Antimutagenesis by factors affecting DNA repair in bacteria. Mutat Res 202:387-391.

\*Kusaka Y, Ichikawa Y, Shirakawa T, et al. 1986a. Effect of hard metal dust in ventilatory function. Brit J Ind Med 43:486-489.

\*Kusaka Y, Iki M, Kumagai S, et al. 1996a. Decreased ventilatory function in hard metal workers. Occup Environ Med 53:194-199.

\*Kusaka Y, Iki M, Kumagai S, et al. 1996b. Epidemiological study of hard metal asthma. Occup Environ Med 53:188-193.

Kusaka Y, Kumagai S, Kyono H, et al. 1992. Determination of exposure to cobalt and nickel in the atmosphere in the hard metal industry. Ann Occup Hyg 36(5):497-507.

\*Kusaka Y, Yokoyama K, Sera Y, et al. 1986b. Respiratory diseases in hard metal workers: An occupational hygiene study in a factory. Brit J Ind Med 43:474-485.

Kusama T, Itoh S, Yoshizawa Y. 1986. Absorption of radionuclides through wounded skin. Health Phys 51(1):138-141.

\*Kyono H, Kusaka Y, Homma K, et al. 1992. Reversible lung lesions in rats due to short-term exposure to ultrafine cobalt particles. Ind Health 30:103-118.

\*Lacy PE, Cardeza AF. 1958. Electron microscopy of guinea pig pancreas. Diabetes 7(5):368-374.

\*Lacy SA, Merritt K, Brown SA, et al. 1996. Distribution of nickel and cobalt following dermal and systematic administration with in vitro and in vivo studies. J Biomed Mater Res 32:279-283.

\*Ladoux A, Frelin C. 1994. Cobalt stimulates the expression of vascular endothelial growth factor and mRNA in rat cardiac cells. Biochem Biophys Res Commun 204(2):794-798.

\*Lafuma C, Wegrowski J, Labat-Robert J, et al. 1987. Parallel increase of plasma fibronectin and perchlorosoluble serum glycoproteins in radiation-induced lung damage. Clin Physiol Biochem 5:61-69.

\*Lafuma J, Chmelevsky D, Chameaud J, et al. 1989. Lung carcinomas in sprague-dawley rats after exposure to low doses of radon daughters, fission neutrons, or y rays. Radiat Res 118:230-245.

Lahaye D, Demedts M, Van Den Oever R, et al. 1984. Lung diseases among diamond polishers due to cobalt? Lancet:156-157.

Laissue JA, Bally E, Joel DD, et al. 1983. Protection of mice from whole-body gamma radiation by deuteration of drinking water. Radiat Res 96:59-64.

\*Lammintausta K, Pitkanen O-P, Kalimo K, et al. 1985. Interrelationship of nickel and cobalt contact sensitization. Contact Dermatitis 13:148-152.

\*Lantzy RJ, Mackenzie FT. 1979. Atmospheric trace metals: Global cycles and assessment of man's impact. Geochemica et Cosmochimica Acta 43:511-525.

\*Laporte P, Viguier-MArtinez M-C, Zongo D, et al. 1985. Changes in testicular fluid production and plasma hormones in the adult rat after testicular <sup>60</sup>Co irradiation. Reprod Nutr Dev 25(2):355-366.

\*Lasfargues G, Lardot C, Delos M, et al. 1995. The delayed lung responses to single and repeated intratracheal administration of pure cobalt and hard metal powder in the rat. Environ Res 69:108-121.

Lasfargues G, Lison D, Maldague P, et al. 1992. Comparative study of the acute lung toxicity of pure cobalt powder and cobalt-tungsten carbide mixture in rat. Toxicol Appl Pharmacol 112(1):41-50.

\*Lasfargues G, Wild P, Moulin JJ, et al. 1994. Lung cancer mortality in a French cohort of hard-metal workers. Am J Ind Med 26:585-595.

Lauwerys R, Lison D. 1994. Health risks associated with cobalt exposure - an overview. Sci Total Environ 150:1-6.

\*Lazarus SS, Goldner MG, Volk BW. 1953. Selective destruction of pancreatic alpha cells by cobaltous chloride in the dog. Metabolism 2:513-520.

LBNL. 2000. The Isotopes Project, Ernest Orlando Lawrence Berkeley National Laboratory, <a href="http://ie.lbl.gov/"><u>Http://ie.lbl.gov/</u></a>. Collaborative Project with Lund Nuclear Data WWW Service, Lund University, Sweden, update 5/30/99, <a href="http://nucleardata.nuclear.lu.se/nucleardata/"><u>Http://nucleardata.nuclear.lu.se/nucleardata/</u></a>. June 21,2001.

Ledney GD, Exum ED, Jackson WE. 1985. Wound-induced alterations in survival of <sup>60</sup>Co irradiated mice: importance of wound timing. Experientia 41:614-616.

\*Lee AC, Angleton GM, Benjamin SA. 1989. Hypodontia in the beagle after perinatal whole-body  $^{60}$ Co  $\gamma$  irradiation. Radiat Res 118:467-475.

\*Lee C, Malpeli JG. 1986. Somata-selective lesions induced by cobaltous chloride: A parametric study. Brain Res 364:396-399.

Lee JY, Watanabe H, Komatsu K, et al. 1997. Developmental anomalies and embryo lethality of  $^{60}$ Co  $\gamma$ -ray irradiation on the embryonic development scid mice. Teratology 55(1):67-68.

\*Leeder JS, Kearns GL. 1997. Pharmcogenetics in pediatrics: Implications for practice. Pediatr Clin North Am 44(1):55-77.

Leghissa P, Ferrari MT, Piazolla S, et al. 1994. Cobalt exposure evaluation in dental prostheses production. Sci Total Environ 150:253-257.

\*Legrum W, Stuehmeier G, Netter KJ. 1979. Cobalt as a modifier of microsomal monooxygenases in mice. Toxicol Appl Pharmacol 48:195-204.

Lehninger AL. 1982. Principles of Biochemistry. New York: Worth Publishers, Inc., 361-466.

Leivouri M, Vallius H. 1998a. A case study of seasonal variation in the chemical composition of accumulating suspended sediments in the central Gulf of Finland. Chemosphere 36(3):503-521.

Leivouri M, Vallius H. 1998b. A case study of seasonal variation in the chemical composition of accumulating suspended sediments in the Central Gulf of Finland. Chemosphere 36(10):2417-2435.

Leonard A, Lauwerys R. 1990. Mutagenic, carcinogenicity and teratogenicity of cobalt metal and cobalt compounds. Mutat Res 239:17-27.

\*Leonard KS, McCubbin D, Harvey BR. 1993a. Chemical speciation and environmental behavior of <sup>60</sup>Co discharged from a nuclear establishment. J Environ Radioact 20:1-21.

\*Leonard KS, McCubbin D, Harvey BR. 1993b. A radiochemical procedure for the determination and speciation of radiocobalt in environmental waters. Sci Total Environ 130/131:237-251.

\*Lessard ET, Miltenberger RP, Cohn SH, et al. 1984. Protracted exposure to fallout: Rongelap and Utirik experience. Health Phys 46:511-527.

Letourneau EG, Jack GC, McCullough RS, et al. 1972. The metabolism of cobalt by the normal human male: Whole body retention and radiation dosimetry. Health Phys 22:451-459.

\*Leung H-W. 1993. Physiologically-based pharmacokinetic modelling. In: Ballentine B, Marro T, Turner P, eds. General and applied toxicology. Vol. 1. New York, NY: Stockton Press, 153-164.

Lewis CPL, Demedts M, Nemery B. 1990. Cobalt induces oxidative stress in pulmonary tissue. Amer Rev Respir Dis 141:A423.

\*Lewis CPL, Demedts M, Nemery B. 1991. Indices of oxidative stress in hamster lung following exposure to cobalt(II) ions: In vivo and in vitro studies. Am J Resp Cell Mol Biol 5:163-169.

Lewis CPL, Demedts M, Nemery B. 1992. The role of thiol oxidation in cobalt(II)-induced toxicity in hamster lung. Biochem Pharmacol 43(3):519-525.

Lewis M, Worobey J, Ramsay DS, et al. 1992. Prenatal exposure to heavy metals: Effect on childhood cognitive skills and health status. Pediatrics 89(6):1010-1015.

Li CS, Hsu LY, Chuang YYT. 1993. Elemental profiles of indoor and outdoor particulate matter less than 10um (PM10) and 2.5um (PM2.5) in Taipei. Chemosphere 27(11):2143-2154.

\*Libshitz HI. 1993. Radiation changes in the lung. Semin Roentgenol 28:303-320.

\*Licht A, Oliver M, Rachmilewitz EA. 1972. Optic atrophy following treatment with cobalt chloride in a patient with pancytopenia and hypercellular marrow. Isr J Med Sci 8:61-66.

\*Lichtenstein ME, Bartl F, Pierce RT. 1975. Control of cobalt exposures during wet process tungsten carbide grinding. Am Ind Hyg Assoc J 36:879-885.

\*Lide, DR, ed. 1994. Handbook of chemistry and physics. 75th edition. Boca Raton, FL: CRC Press, Inc., 4-3, 37-8.

\*Lide, DR, ed. 1998. Handbook of chemistry and physics. 79th edition. Boca Raton, FL: CRC Press, Inc., 4-38, 11-41, 143-4.

# COBALT 339 9. REFERENCES

Liden C, Wahlberg JE. 1994. Cross-reactivity to metal compounds studies in guinea pigs induced with chromate or cobalt. Acta Derm Venereol (Stockh) 74(5):341-343.

Lin L, Villalon P, Martasek P, et al. 1990. Regulation of heme oxygenase gene expression by cobalt in rat liver and kidney. Eur J Biochem 192:577-582.

\*Lindahl-Kiessling K, Santesson B, Book JA. 1970. Chromosome and chromatid-type aberrations induced by cobalt60 irradiation and tritiated urindine in human leukocyte cultures. Chromosoma 31:280-284.

Linnainmaa M, Kiilunen M. 1997. Urinary cobalt as a measure of exposure in the wet sharpening of hard metal and stellite blades. Int Arch Occup Environ Health 69:193-200.

\*Linnainmaa M, Kangas J, Kalliokoski P. 1996. Exposure to airborne metals in the manufacture and maintenance of hard metal and stellite blades. Am Ind Hyg Assoc J 57:196-201.

\*Lisk DJ, Gutenmann WH, Rutzke M, et al. 1992. Survey of toxicants and nutrients in composted waste materials. Arch Environ Contam Toxicol 22:190-194.

Lison D. 1996. Human toxicity of cobalt-containing dust and experimental studies on the mechanism of interstitial lung disease (hard metal disease). Crit Rev Toxicol 26(6):585-616.

Lison D, Lauwerys R. 1990. In vitro cytotoxic effects of cobalt-containing dusts on mouse peritoneal and rat alveolar macrophages. Environ Res 52:187-198.

Lison D, Lauwerys R. 1991. Biological responses of isolated macrophages to cobalt metal and tungsten carbide-cobalt powders. Pharmacol Toxicol 69:282-285.

Lison D, Lauwerys R. 1992. Study of the mechanism responsible for the elective toxicity of tungsten carbide-cobalt powder toward macrophages. Toxicol Lett 60:203-210.

Lison D, Lauwerys R. 1993. Evaluation of the role of reactive oxygen species in the interactive toxicity of carbide-cobalt mixtures on macrophages in culture. Arch Toxicol 67:347-351.

Lison D, Lauwerys R. 1994. Cobalt bioavailability from hard metal particles. Arch Toxicol 68:528-531.

Lison D, Lauwerys R. 1995. The interaction of cobalt metal with different carbides and other mineral particles on mouse peritoneal macrophages. Toxicol in Vitro 9(3):341-347.

Lison D, Buchet JP, Swennen B, et al. 1994. Biological monitoring of workers exposed to cobalt metal, salt, oxides, and hard metal dust. Occup Environ Med 51:447-450.

\*Lison D, Carbonnelle P, Mollo L, et al. 1995. Physicochemical mechanism of the interaction between cobalt metal and carbide particles to generate toxic activated oxygen species. Chem Res Toxicol 8:600-606

\*Lison D, Lauwerys R, Demedts M, et al. 1996. Experimental research into the pathogenesis of cobalt/hard metal lung disease. European Respiratory Journal 9:1024-1028.

\*Livingston, AL. 1978. Forage plant estrogens. J Toxicol Environ Health 4:301-324.

\*Llena JF, Cespedes G, Hirano A, et al. 1976. Vascular alterations in delayed radiation necrosis of the brain. Arch Pathol Lab Med 100:531-534.

Llobet JM, Domingo JL, Corbella J. 1985. Comparison of antidotal efficacy of chelating agents upon acute toxicity of Co(II) in mice. Res Commun Chem Pathol Pharmacol 50(2):305-308.

\*Llobet JM, Domingo JL, Corbella J. 1988. Comparative effects of repeated parenteral administration of several chelators on the distribution and excretion of cobalt. Res Commun Chem Pathol Pharmacol 60(2):225-233.

\*Lloyd DR, Phillips DH, Carmichael PL. 1997. Generation of putative intrastrand cross-links and strand breaks in DNA by transition metal ion-mediated oxygen radical attack. Chem Res Toxicol 10:393-400.

Lobel PB, Longerich HP, Jackson SE, et al. 1991. A major factor contributing to the high degree of unexplained variability of some elements concentrations in biological tissue: 27 elements in 5 organs of the mussel Mytilus as a model. Arch Environ Contam Toxicol 21:118-125.

\*Lofstrom A, Wigzell H. 1986. Antigen specific human T cell lines for cobalt chloride. Acta Derm Venereol (Stockh) 66:200-206.

\*Lohmann W, Denny WF, Perkins WH, et al. 1966. Influence of roentgen and <sup>60</sup> Co gamma rays on DNA synthesis in hamster organs. Acta Radiologica Therapy Physics Biology 4(1):3-6.

Lorusso GF, De Stasio G, Gilbert B, et al. 1998. High sensitivity quantitative analysis of cobalt uptake in rat cerebral granule cells with and without excitatory amino acids. Neurosci Lett 248:9-12.

\*Lucke-Huhle C, Pech M, Herrlich P. 1986. Selective gene amplification in mammalian cells after exposure to <sup>60</sup>Co γ rays, <sup>241</sup>Am χ particles, or uv light. Radiat Res 106:345-355.

\*Lucke-Huhle C, Pech M, Herrlich P. 1990. SV40 DNA amplification and reintegration in surviving hamster cells after <sup>60</sup>Co γ-irradiation. Int J Radiat Biol 58(4):577-588.

Lugowski SJ, Smith DC, McHugh AD, et al. 1991. Release of metal ions from dental implant materials in vivo: Determination of Al, Co, Cr, Mo, Ni, V, and Ti in organ tissue. J Biomed Mater Res 25:1443-1458.

Lundborg M, Falk R, Johansson A, et al. 1992. Phagolysosomal pH and dissolution of cobalt oxide particles by alveolar macrophages. Environ Health Perspect 97:153-157.

Lundborg M, Johard U, Johansson A, et al. 1995. Phagolysosomal morphology and dissolution of cobalt oxide particles by human and rabbit alveolar macrophages. Exp Lung Res 21:51-66.

\*Lux D, Kammerer L, Ruhm W, et al. 1995. Cycling of Pu, Sr, Cs, and other long living radionuclides in forest ecosystems of the 30-km zone around Chernobyl. Sci Total Environ 173/174:375-384.

Lymberis C, Makrigiorgos G, Sbonias E, et al. 1987. Radiocesium levels in human muscle samples in Greece one year after the Chernobyl accident. Appl Radiat Isot 39(2):175-176.

Lytle TF, Lytle JS. 1990. Heavy metals in the eastern oyster, Crassostrea virginica, of the Mississippi Sound. Bull Environ Contam Toxicol 44:142-148.

MacVicar BA. 1987. Morphological differentiation of cultured astrocytes is blocked by cadmium or cobalt. Brain Res 420:175-177.

Madden JD, Grodner RM, Feagley SE, et al. 1991. Minerals and xenobiotic residues in the edible tissues of wild and pond-raised Louisiana crayfish. J Food Saf 12:1-15.

Madruga MJ, Carreiro MCV. 1992. Experimental study of <sup>60</sup>Co behavior in Tejo River sediments. Hydrobiologia 235/236:661-668.

\*Maenhaut W, Zoller WH, Duce RA, et al. 1979. Concentration and size distribution of particulate trace elements in the south polar atmosphere. J Geophys Res 84(C5):2421-2431.

Mahara Y, Kudo A. 1980. Mobility and retention of <sup>60</sup>Co in soils in coastal areas. In: Radiation Protection: A systematic approach to safety. New York, NY: Pergamon Press, 1111-1142.

\*Mahara Y, Kudo A. 1981. Interaction and mobility of cobalt-60 between water and sediments in marine environments possible effects by acid rain. Water Res 15(4):413-419.

\*Maier DM, Landauer MR. 1989. Effects of acute sublethal gamma radiation exposure on aggressive behavior in male mice: A dose-response study. Aviation, Space, and Environmental Medicine, 774-778.

Maines MD, Kappas A. 1975. Study of the developmental pattern of heme catabolism in liver and the effects of cobalt on cytochrome P-450 and the rate of heme oxidation during the neonatal period. J Exp Med 141:1400-1410.

Maines MD, Janousek V, Tomio JM, et al. 1976. Cobalt inhibition of synthesis and induction of  $\delta$ -aminolevulinate synthase in liver. Proc Natl Acad Sci U S A 73(5):1499-1503.

Malanin G, Kalimo K. 1992. Occupational contact dermatitis due to delayed allergy to pig epithelia. Contact Dermatitis 26:134-135.

\*Malinski T, Fish J, Matsusiewicz H. 1988. Determining ultratrace metal concentrations by inductively coupled plasma emission spectrometry. Am Water Works Assoc 80:81-85.

Malzone WF, Wilder BJ, Mayersdorf A. 1972. A method of modifying the rapidity of cobalt-induced epileptogenesis in the cat. Epilepsia 13:643-648.

Manciet JR, Barrade A, Janssen F, et al. 1995. Contact allergy with immediate and delayed photoaggravation to chromate and cobalt. Contact Dermatitis 33:282-284.

Manninen H, Perkio A, Palonen J, et al. 1996. Trace metal emissions from co-combustion of refuse derived and packaging derived fuels in a circulating fluidized bed boiler. Chemosphere 32(12):2457-2469.

\*Mantoura RFC, Dickson A, Riley JP. 1978. The complexation of metals with humic materials in natural waters. Estuarine Coastal Shelf Sci 6:387-408.

Mao Y, Liu KJ, Jiang JJ, et al. 1996. Generation of reactive oxygen species by Co(II) from  $H_2O_2$  in the presence of chelators in relation to DNA damage and 2'-deoxuguanosine hydroxylation. J Toxicol Environ Health 47:61-75.

\*Marcussen PV. 1963. Cobalt dermatitis. Clinical picture. Acta Derm Venereol (Stockh) 43:231-234.

# COBALT 9. REFERENCES

Marks GS. 1994. Heme oxygenase: The physiological role of one of its metabolites, carbon monoxide and interactions with zinc protoporphyrin and other metalloporphyrins. Cell Mol Biol 40(7):863-870.

Marmolejo-Rivas C, Paez-Osuna F. 1990. Trace metals in tropical coastal lagoon bivalves, mytella strigata. Bull Environ Contam Toxicol 45:545-551.

Marsh GM, Gula MJ, Youk AO, et al. 1999. Mortality among chemical plant workers exposed to acrylonitrile and other substances. Am J Ind Med 36:423-436.

Marston HR. 1970. The requirement of sheep for cobalt or for vitamin  $B_{12}$ . Br Med J 24:615-633.

\*Martin RG, Ruckdeschel JC, Chang P, et al. 1975. Radiation-related pericarditis. Am J Cardiol 35:216-220.

Maruta K, Osa T, Inoue H. 1989. Comparison of Mg, Mn, and Co ions affecting the β-adrenoceptor-mediated membrane response in the guinea-pig taenia caeci. Jpn J Physiol 39:659-671.

Maruyama Y, Feola JM, Magura C, et al. 1985. Study of acute <sup>60</sup>Co, low dose rate CF-252 and CS-137 radiation on LSA ascites lymphoma in vivo. Int J Radiat Oncol Biol Phys 11:1991-1997.

\*Mascanzoni D. 1989. Long-term transfer from soil to plant of radioactive corrosion products. Environ Pollut 57:49-62.

Massone L, Anonide A, Borghi S, et al. 1991. Positive patch test reactions to nickel, cobalt, and potassium dichromate in a series of 576 patients. Cutis 47:119-122.

Mat I. 1994. Arsenic and trace metals in commercially important bivalves, anadara granosa and paphia undulata. Bull Environ Contam Toxicol 52:833-839.

Matsubara S, Horiuchi J, Okuyama T, et al. 1985. Chromosome aberrations in the peripheral lymphocytes induced by brachytherapy and external cobalt teletherapy. Int J Radiat Oncol Biol Phys 11:1085-1094.

- \*Mayr U, Butsch A, Schneider S. 1992. Validation of two in vitro test systems for estrogenic activities with zearalenone, phytoestrogens and cereal extracts. Toxicology 74:135-149.
- \*Mayfield KP, Lai J, Porreca F. 1994. Selective upregulation of opioid delta receptors in NG 108-15 cells by treatment with cobalt: Possible hypoxic regulation. Regul Peptides 54(1):183-184.
- \*Mazur L, Manowska J, Bobik R. 1991. Effects of <sup>60</sup>Co gamma-irradiation of mice on the temporal changes of acid phosphatase activity in spleen and liver. Acta Physiologica Hungarica 78:(2)135-141.
- \*McBrien MP. 1973. Vitamin  $B_{12}$  malabsorption after cobalt teletherapy for carcinoma of the bladder. Br Med J 1:648-650.
- \*McCartney M, Kershaw PJ, Woodhead DS, et al. 1994. Artificial radionuclides in the surface sediments of the Irish Sea, 1968-1988. Sci Total Environ 141:103-138.
- \*McLaren JW, Mykytiuk AP, Willie SN, et al. 1985. Determination of trace metals in seawater by inductively coupled plasma mass spectrometry with preconcentration on silica-immobilized 8-hydroxyquinoline. Anal Chem 57:2907-2911.

# COBALT 9. REFERENCES

McLaren P, Little DI. 1987. The effects of sediment transport on contaminant dispersal: An example from Milford Haven. Mar Pollut Bull 18(11):586-594.

\*McLaren RG, Lawson DM, Swift RS. 1986. Sorption and desorption of cobalt by soils and soil components. J Soil Sci 37:413-426.

\*McLean RI, Summers JK. 1990. Evaluation of transport and storage of <sup>60</sup>Co, <sup>134</sup>Cs, <sup>137</sup>Cs and <sup>65</sup>Zn by river sediments in the lower Susquehanna River. Environ Poll 63:137-153.

MDE. 1999. News release: MDE seeks court action against neutron products for decommissioning of its cobalt-60 production facility. Maryland Department of the Environment. Http://www.mde.state.md.us. April 20, 2000.

\*MDS Nordion 2000. Cobalt 60 Sources. MDS Nordion, Toronto, Canada. <a href="http://www.mds.nordion.com"><u>Http://www.mds.nordion.com</u></a>. January 16, 2000.

Meachim G, Pedley RB, Wiliams DF. 1982. A study of sarcogenicity associated with Co-Cr-Mo particles implanted in animal muscle. J Biomed Mater Res 16:407-416.

\*Meecham HM, Humphrey P. 1991. Industrial exposure to cobalt causing optic atrophy and nerve deafness: A case report. J Neurol Neurosurg Psychiatry 54(4):374-375.

Meijer C, Bredberg M, Fischer T, et al. 1995. Ear piercing and nickel and cobalt sensitization, in 520 young Swedish men doing compulsory military service. Contact Dermatitis 32:147-149.

\*Mejstrik V, Svacha J. 1988. Concentrations of Co, Cd, Ni, and Zn in crop plants cultivated in the vicinity of coal-fired power plants. Sci Total Environ 72:57-67.

\*Mele PC,Franz CG, Harrison JR. 1988. Effects of sublethal doses of ionizing radiation on schedule-controlled performance in rats. Pharmacol Biochem Behav 30:1007-1014.

Mendoza CA, Cortes G, Munoz D. 1996a. Heavy metal pollution in soils and sediments of rural developing district 063, Mexico. Environ Toxicol Water Qual 11:327-333.

Mentasti E, Abollino O, Aceto M, et al. 1998. Distribution of statistical correlations of major, minor and trace metals in lake environments of Antarctica. Int J Environ Anal Chem 71(3-4):245-255.

Meplan C, Richard M-J, Hainaut P. 2000. Metalloregulation of the tumor suppressor protein p53: zinc mediates the renaturation of p53 after exposure to metal chelators *in vitro* and in intact cells. Oncogene 19(46):5227-5236.

\*Meranger JC, Subramanian KS, Chalifoux C. 1981. Metals and other elements: Survey for cadmium, cobalt, chromium, copper, nickel, lead, zinc, calcium, and magnesium in Canadian drinking water supplies. J Assoc Off Anal Chem 64(1):44-53.

\*Merian E. 1985. Introduction on environmental chemistry and global cycles of chromium, nickel, cobalt, beryllium, arsenic, cadmium and selenium, and their derivatives. Curr Top Environ Toxicol Chem 8:3-32.

\*Mermut AR, Jain JC, Song L, et al. 1996. Trace element concentrations of selected soils and fertilizers in Saskatchewan, Canada. J Environ Qual 25:845-853.

# COBALT 9. REFERENCES

Merritt K, Crowe TD, Brown SA. 1989. Elimination of nickel, cobalt, and chromium following repeated injections of high dose metal salts. J Biomed Mater Res 23:845-862.

Meyers-Schone L, Walton BT. 1994. Turtles as monitors of chemical contaminants in the environment. Rev Environ Toxicol 135:93-153.

Mahara Y, Kudo A. 1981b. Fixation and mobilization of 60Co on sediments in coastal environments. Health Phys 41(4):645-655.

Michetti G, Mosconi G, Zanelli R, et al. 1994. Bronchoalveolar lavage and its role in diagnosing cobalt lung disease. Sci Total Environ 150:173-178.

Migliori M, Mosconi G, Michetti G, et al. 1994. Hard metal disease: Eight workers with interstitial lung fibrosis due to cobalt exposure. Sci Total Environ 150:187-196.

\*Milford JB, Davidson CI. 1985. The size of particulate trace elements in the atmosphere - a review. J Air Pollut Control Assoc 35(12):1249-1260.

Milkovic-Kraus S, Kubelka D, Vekic B. 1992. Biological monitoring of three <sup>60</sup>Co radiation incident victims. Am J Ind Med 22:243-247.

Miller ME, Howard D, Stohlman F, et al. 1974. Mechanism of erythropoietin production by cobaltous chloride. Blood 44(3):339-346.

\*Miller-Ihli NJ, Wolf WR. 1986. Characterization of a diet reference material for 17 elements. Anal Chem 58:3225-3231.

\*Miltenberger RP, Lessard ET, Greenhouse NA. 1981. <sup>60</sup>Co and <sup>137</sup>Cs long-term biological removal rate constants for the marshallese population. Health Phys 40:615-623.

Miyamoto T, Iwasaki K, Mihara Y, et al. 1997. Lymphocytoma cutis induced by cobalt. Br J Dermatol 137:467-484.

\*Mochizuki H, Kada T. 1982. Antimutagenic action of cobaltous chloride on trp-P-1-induced mutations in salmonella typhimurium TA98 and TA1538. Mutat Res 95:145-157.

Mochizuki H, Kada T. 1984. Mechanisms of antimutagenicity of cobaltous chloride: Analysis of SOS reactions in Escherichia coli B/r. Mutat Res 130:371.

\*Moger WH. 1983. Effects of the calcium-channel blockers cobalt, verapamil, and D600 on leydig cell steroidogenesis. Biol Reprod 28:528-535.

Mohapatra SP. 1988. Distribution of heavy metals in polluted creek sediment. Environ Monit Assess 10(2):157-163.

\*Mohiuddin SM, Taskar PK, Rheault M, et al. 1970. Experimental cobalt cardiomyopathy. Am Heart J 80(4):532-543.

\*Mollenhauer HH, Corrier DE, Clark DE, et al. 1985. Effects of dietary cobalt on testicular structure. Virchows Arch B 49:241-248.

Momeni MH, Worden L, Goldman M. 1974. Dosimetry and facilities of UCD outdoor-indoor <sup>60</sup>Co irradiator. Health Phys 26:469-472.

Monnet-Tschudi F, Zurich MG, Honegger P. 1993. Evaluation of the toxicity of different metal compounds in the developing brain using aggregating cell cultures as a model. Toxicol in Vitro 7(4):335-339.

Monsees TK, Winterstein U, Hayatpour J, et al. 1998. Effect of heavy metals on the secretory function of testicular cells in culture. J Trace Microprobe Tech 16(4):427-435.

Montiel C, Artalejo AR, Sanchez-Garcia P, et al. 1993. Two components in the adrenal nicotinic secretory response revealed by cobalt ramps. Eur J Pharmacol 230:77-84.

\*Moorehouse CP, Halliwell B, Grootveld M, et al. 1985. Cobalt(II) ion as a promoter of hydroxyl radical and possible 'crypto-hydroxyl' radical formation under physiological conditions. Differential effects of hydroxyl radical scavengers. Biochim Biophys Acta 843:261-268.

Moratal J, Castells J, Donaire A, et al. 1994. Interaction of cobalt ions with carboxypeptidase A. J Inorg Biochem 53:1-11.

Morel FMM, Westall JC, O'Melia CR, et al. 1975. Fate of trace metals in Los Angeles County wastewater discharge. Environ Sci Technol 9(8):756-761.

\*Morelli L, Di Giulio C, Iezzi M, et al. 1994. Effect of acute and chronic cobalt administration on carotid body chemoreceptors responses. Sci Total Environ 150:215-216.

Morgan GW, Breit SN. 1995. Radiation and the lung: A reevaluation of the mechanisms mediating pulmonary injury. Int J Radiat Oncol Biol Phys 31(2):361-369.

Morgan RM, Kundomal YR, Hupp EW. 1983. Serum lactate dehydrogenase (LDH) activity following exposures to cadmium and /or [+60]Co gamma irradiation. J Environ Sci Health Part A 18(4):483-492.

Morgan RM, Kundomal YR, Hupp EW. 1987. Serum alkaline phosphatase (SAP) activity following exposure to cadmium and/or [+60]Co gamma irradiation. J Environ Sci Health Part A 22(4):337-342.

\*Morin Y, Tetu A, Mercier G. 1971. Cobalt cardiomyopathy: Clinical aspects. Br Heart J 33:175-178.

Morita H, Noda K, Umeda M. 1985. Mutagenicities of nickel and cobalt compounds in a mammalian cell line. Mutat Res 147:265-266.

Morita H, Umeda M, Ogawa HI. 1991. Mutagenicity of various chemicals including nickel and cobalt compounds in cultured mouse FM3A cells. Mutat Res 261:131-137.

Morita Y, Mizutani M. 1987. Inhibitory effect of cobaltous chloride on mutagenesis induced by N-methyl-N'-nitro-N-nitrosoguanidine (MNNG) in FM3A cells. Mutat Res 182:367-368.

Morral FR. 1979. Cobalt compounds. In: Kirk RE, Othmer DF, Grayson M, et al., ed. Kirk-Othmer encyclopedia of chemical technology. New York, NY: John Wiley and Sons, 495-510.

Morrison RA, Zellmer DL, Dean RD. 1981. Low vs high dose-rate effects on the acute reaction of pig skin to cobalt-60 gamma rays. Int J Radiat Oncol Biol Phys 7:359-364.

# COBALT 346 9. REFERENCES

Morrison RJ, Gangaiya P, Naqasima MR, et al. 1997. Trace element studies in the Great Astrolabe Lagoon, Fiji, a pristine marine environment. Mar Pollut Bull 34(5):353-356.

\*Morselli PL, Franco-Morselli R, Bossi L. 1980. Clinical pharmacokinetics in newborns and infants: Age-related differences and therapeutic implications. Clin Pharmacokin 5:485-527.

Morsy SM, El-Assaly FM. 1970. Body elimination rates of <sup>134</sup>Cs, <sup>60</sup>Co and <sup>203</sup>Hg. Health Phys 19:769-773

\*Morvai V, Szakmary E, Tatrai E, et al. 1993. The effects of simultaneous alcohol and cobalt chloride administration on the cardiovascular system of rats. Acta Physiol Hung 81(3):253-261.

\*Mosconi G, Bacis M, Leghissa P, et al. 1994a. Occupational exposure to metallic cobalt in the province of Beragmo. Results of a 1991 survey. Sci Total Environ 150:121-128.

\*Mosconi G, Bacis M, Vitali MT, et al. 1994b. Cobalt excretion in urine: Results of a study on workers producing diamond grinding tools and on a control group. Sci Total Environ 150:133-139.

Mosher BW, Winkler P, Jaffrezo JL. 1993. Seasonal aerosol chemistry at dye 3, Greenland. Atmos Environ 27A(17/18):2761-2772.

Motelica-Heino M, Coustumer PL, Thomassin JH, et al. 1998. Macro and microchemistry of trace metals in vitrified domestic wastes by laser ablation ICP-MS and scanning electron microprobe X-ray energy dispersive spectroscopy. Talanta 46:407-422.

Mothersill C, Seymour CB, Harney J, et al. 1994. High levels of stable p53 protein and the expression of c-myc in cultured human epithelial tissue after cobalt-60 irradiation. Radiat Res 137:317-322.

Moulin JJ, Wild P, Mur JM, et al. 1993. A mortality study of cobalt production workers: An extension of the follow-up. Am J Ind Med 23:281-288.

\*Moulin JJ, Wild P, Romazini S, et al. 1998. Lung cancer risk in hard-metal workers. Am J Epidemiol 148(3):241-248.

\*Mucklow ES, Griffin SJ, Delves HT, et al. 1990. Cobalt poisoning in a 6-year old. The Lancet. 335:981.

Mudroch A. 1985. Geochemistry of the Detroit River sediments. Great Lakes Res Rev 11(3):193-200.

\*Mumma RO, Raupach DC, Sahadewan K, et al. 1990. National survey of elements and radioactivity in municipal incinerator ashes. Arch Environ Contam Toxicol 19:399-404.

Mumma RO, Raupach DC, Sahadewan K, et al. 1991. Variation in elemental composition of municipal refuse incinerator ashes with time of sampling. Chemosphere 23(3):391-395.

\*Mumma RO, Raupach DC, Waldman JP, et al. 1984. National survey of elements and other constituents in municipal sewage sludges. Arch Environ Contam Toxicol 13:75-83.

Mundschenk VH. 1991. [On the behavior of the radioisotopes <sup>58</sup>Co and <sup>60</sup>Co from nuclear power plants in the case of the Rhine River]. Z Wasser Abwasser Forsch 24:268-284.

# COBALT 9. REFERENCES

- \*Munita CS, Mazzilli BP. 1986. Determination of trace elements in Brazilian cigarette tobacco by neutron activation analysis. J Radioanal Nucl Chem 108(4):217-227.
- \*Mur JM, Moulin JJ, Charruyer-Seinerra MP, et al. 1987. A cohort mortality study among cobalt and sodium workers in an electrochemical plant. Am J Ind Med 11:75-81.
- \*Muramatsu Y, Parr RM. 1988. Concentrations of some trace elements in hair, liver and kidney from autopsy subjects relationship between hair and internal organs. Sci Total Environ 76:29-40.
- \*Murdock HR. 1959. Studies on the pharmacology of cobalt chloride. J Am Pharm Assoc Sci Ed 48:140-142.
- \*Murray RL. 1994. Understanding radioactive waste, 4th edition. Battelle Pacific Northwest Laboratories, Battelle Press.
- \*Murthy GK, Rhea U, Peeler JT. 1971. Levels of antimony, cadmium, cobalt, manganese, and zinc in institutional total diets. Environ Sci Technol 5(5):436-442.
- \*Mutafova-Yambolieva V, Staneva-Stoytcheva D, Lasova L, et al. 1994. Effects of cobalt or nickel on the sympathetically mediated contractile responses in rat-isolated vas deferens. Pharmacology 48:100-110.
- \*Myskowski PL, and Safai B. 1981. Localized comedo formation after cobalt irradiation. Int Society of Tropical Dermatology Inc, 550-551.
- \*Nackerdien Z, Kasprak KS, Rao G, et al. 1991. Nickle(II)-and cobalt(II)-dependent damage by hydrogen peroxide to the DNA bases in isolated human chromatin. Cancer Res 51:5837-5842.

Nadeennko VG, Lenchenko VG, Saichenko SP, et al. 1980. [Embryotoxic action of cobalt in peroral body uptake]. Gig Sanit 2:6-8.

Nagao M, Sugaru E, Kambe T, et al. 1999. Unidirectional transport from apical to basolateral compartment of cobalt ion in polarized Madin-Darby canine kidney cells. Biochem Biophys Res Commun 257:289-294.

Nagy I, Woolf CJ, Dray A, et al. 1994. Cobalt accumulation in neurons expressing ionotropic excitatory amino acid receptors in young rat spinal cord: Morphology and distribution. J Comp Neur 344:321-335.

\*Naidu AS, Blanchard A, Kelley JJ, et al. 1997. Heavy metals in Chukchi Sea sediments as compared to selected circum-arctic shelves. Mar Pollut Bull 35:260-269.

Nakamura M, Yasukochi Y, Minakami S. 1975. Effect of cobalt on heme biosynthesis in rat liver and spleen. J Biochem 78:373-380.

Nakashima S, Sturgeon RE, Willie SN, et al. 1988. Determination of trace metals in seawater by graphite furnace atomic absorption spectrometry with preconcentration on silica-immobilized 8-hydroxyquinoline in a flow-system. Fresenius Z Anal Chem 330:592-595.

\*Namba M, Nishitani K, Fukushima F, et al. 1981. Neoplastic transformation of human diploid fibroblasts reacted with chemical carcinogens and Co-60 γ-rays. Gann Monogr Cancer Res 27:221-230.

# COBALT 348 9. REFERENCES

\*Namba M, Nishitani K, Fukushima F, et al. 1988. Multi-step neoplastic transformation of normal human fibroblasts by Co-60 gamma rays and Ha-ras oncogenes. Mutat Res 199:415-423.

\*Namba M, Nishitani K, Hyodoh F, et al. 1985. Neoplastic transformation of human diploid fibroblasts (KMST-6) by treatment with 60Co gamma rays. Indian J Cancer 35:275-280.

\*NAS. 1977. Drinking water and health. National Academy of Sciences, Washington, DC, 209-211, 247.

\*NAS/NRC. 1989. Report of the oversight committee. In: Biologic markers in reproductive toxicology. Washington, DC: National Academy of Sciences, National Research Council, National Academy Press.

Nasu T. 1992. Calcium antagonism by cobalt ions on contraction of guinea-pig taenia coli. J Pharm Pharmacol 44:879-884.

\*Nation JR, Bourgeois AE, Clark DE, et al. 1983. The effects of chronic cobalt exposure on behavior and metallothionein levels in the adult rat. Neurobehav Toxicol Teratol 5:9-15.

Nayebzadeh A, Dufresne A, Harvie S, et al. 1999. Mineralogy of lung tissue in dental laboratory technician's pneumoconiosis. Am Ind Hyg Assoc J 60:349-353.

\*Naylor GPL, Harrison JD. 1995. Gastrointestinal iron and cobalt absorption and iron status in young rats and guinea pigs. Human Exp Toxicol 14:949-954.

\*NCRP. 1987. Use of bioassay procedures for assessment of internal radionuclide deposition. National Council on Radiation Protection and Measurements. Bethesda, MD: NCRP; NCRP Report No. 87.

\*NCRP. 1993. Limitation of exposure to ionizing radiation. National Council on Radiation Protection.

\*NCRP. 1997. Deposition, retention and dosimetry of inhaled radioactive substances. National Council on Radiation Protection and Measurements. Bethesda, MD: NCRP; NCRP Report No. 125.

Neal C, Smith CJ, Jeffery HA, et al. 1996. Trace element concentrations in the major rivers entering the Humber estuary, NE England. J Hydrol 182:37-64.

Neal C, Smith CJ, Walls J, et al. 1990. Hydrogeochemical variations in Hafren Forest stream waters, Mid-Wales. J Hydrol 116:185-200.

Nellessen JE, Fletcher JS. 1993. Assessment of published literature on the uptake, accumulation, and translocation of heavy metals by vascular plants. Chemosphere 27(9):1669-1680.

\*Nemery B, Casier P, Roosels D, et al. 1992. Survey of cobalt exposure and respiratory health in diamond polishers. Am Rev Respir Dis 145:610-616.

Nemery B, Lewis CPL, Demedts M. 1994. Cobalt and possible oxidant-mediated toxicity. Sci Total Environ 150:57-64.

Nemery B, Nagels J, Verbeken E, et al. 1990. Rapidly fatal progression of cobalt lung in a diamond polisher. Am Rev Respir Dis 141(5):1373-1378.

Nemery B, Roosels D, Lahaye D, et al. 1988. Cross-sectional survey of lung function and assessment of cobalt exposure in diamond polishers. Am Rev Resp Dis 137:96.

# COBALT 9. REFERENCES

\*Nevissi AE. 1992. Measurement of actinides and long-lived radionuclides in large coral samples. J Radioanalyt Nucl Chem 156:243-251.

\*Newton D, Rundo J. 1971. The long term retention of inhaled cobalt-60. Health Phys 21:(3)377-384.

Nies DH. 1992. Resistance to cadmium, cobalt, zinc, and nickel in microbes. Plasmid 27:17-28.

\*Nimmo M, Chester R. 1993. The chemical speciation of dissolved nickel and cobalt in Mediterranean rainwaters. Sci Total Environ 135:153-160.

\*Nimmo M, Fones GR. 1997. The potential pool of Co, Ni, Cu, Pb and Cd organic complexing ligands in coastal and urban rain waters. Atmos Environ 31(5):693-702.

NIOSH. 1973. Chronic animal inhalation toxicity to cobalt. Cincinnati, OH: National Institute for Occupational Safety and Health, Center for Disease Control. PB 232 247.

NIOSH. 1989. Health hazard evaluation report no. HETA-85-295-1907. General Electric carboloy systems, Detroit, Michigan. National Institute for Occupational Safety and Health, Department of Health and Human Services.

NIOSH. 2000a. Cobalt. NIOSH pocket guide to chemical hazards. National Institute for Occupational Safety and Health. <a href="http://www.cdc.gov">http://www.cdc.gov</a>. March 13, 2000.

NIOSH. 2000b. Radioactive cobalt. NIOSH pocket guide to chemical hazards. National Institute for Occupational Safety and Health. Http://www.cdc.gov/niosh/homepage.html. March 13, 2000.

\*NIOSH. 2001. REL (TWA), cobalt metal, dust, and fume. National Institute for Occupational Safety and Health. <a href="http://www.cdc.gov/niosh/srchpage.html">http://www.cdc.gov/niosh/srchpage.html</a>. February 23, 2001.

Nishigaki I, Oku H, Noguchi M, et al. 1993. Prevention by ellagic acid of lipid peroxidation in placenta and fetus of rats irradiated with 60Co. J Clin Biochem Nutr 15:135-141.

\*Nishimura Y, Inaba J, Ichikawa R. 1978. Fetal uptake of <sup>60</sup>CoCl<sub>2</sub> and <sup>57</sup>Co-cyanocobalamin in different gestation stages of rats. J Radiat Res 19:236-245.

\*Nitta Y, Kamiya K, Yokoro K. 1992. Carcinogenic effect of in utero <sup>252</sup>Cf and <sup>60</sup>Co irradiation in C57BL/6NXC3H/He F1 (B6C3F1) mice. J Radiat Res 33:319-333.

\*Nojiri Y, Kawai T, Otsuki A, et al. 1985. Simultaneous multielement determinations of trace metals in lake waters by ICP emission spectrometry with preconcentration and their background levels in Japan. Water Res 19(4):503-509.

Nolte J. 1988. Pollution source analysis of river water and sewage sludge. Environ Technol Lett 9:857-868.

Nordberg G. 1994. Assessment of risks in occupational cobalt exposures. Sci Total Environ 150:201-207.

\*Norris WP, Poole CM. 1969. The response of ANL beagles to protracted exposure to <sup>60</sup>Co gamma rays at 5 to 35 R/day. II. Estimation of the LD50 at 35 R/day. In: Biological and Medical Research Division Annual Report. Argonne National Laboratory, IL.

# COBALT 350 9. REFERENCES

- NRC. 1982. Evaluation of isotope migration-land burial: Water chemistry at commercially operated low-level radioactive waste disposal sites. Washington, DC: Nuclear Regulatory Commission, Office of Nuclear Regulation Research. NTIS/NUREG/CR-2124.
- \*NRC. 1984. Lower limit of detection: Definition and elaboration of a proposed position for radiological effluent and environmental measurements. Nuclear Regulatory Commission. Washington, DC: NRC; U.S. Report NUREG/CR-4604.
- \*NRC. 1991. Nuclear Regulatory Commission. Washington, DC.
- \*NRC. 1993. Pesticides in the diets of infants and children. National Research Council. Washington, DC: National Academy Press.
- \*NRC. 1997. Minimum detectable concentrations with typical radiation survey instruments for various contaminants and field conditions. Nuclear Regulatory Commission. Rockville, MD: NRC; U.S. Report NUREG-1507.
- \*NRC. 1999. Annual limits on intake (ALIs) and derived air concentrations (DACs) of radionuclides for occupational exposure: Effluent concentration: Concentrations for release to sewerage. Nuclear Regulatory Commission. Code of Federal Regulations. 10 CFR 20 Sub O, Appendix B.
- \*NRC. 2000a. Quantities of radioactive materials requiring consideration of the need for an emergency lan for responding to release. Nuclear Regulatory Commission. Code of Federal Regulations. 10 CFR 30.72 Schedule C.
- \*NRC. 2000b. Quantities of licensed material requiring labeling. Nuclear Regulatory Commission. Code of Federal Regulations. 10 CFR 30, Appendix B.
- \*NRC. 2000c. Use of sources for brachytherapy. Nuclear Regulatory Commission. Code of Federal Regulations. 10 CFR 35.400.
- \*NRC. 2001a. Activity values for radionuclides. Nuclear Regulatory Commission. Code of Federal Regulations. 10 CFR 71. Http://www.nrc.gov. March 13, 2001.
- \*NRC. 2001b. Byproduct material listing. Nuclear Regulatory Commission. Code of Federal Regulations. 10 CFR 30.71. <a href="http://www.nrc.gov"><u>Http://www.nrc.gov</u></a>. March 13, 2001.
- \*NRC. 2001c. Byproduct material listing. Nuclear Regulatory Commission. Code of Federal Regulations. 10 CFR 33.100. Http://www.nrc.gov. March 23, 2001.
- \*NRC. 2001d. Byproduct material listing, electron tubes, spark gap irradiators. Nuclear Regulatory Commission. Code of Federal Regulations. 10 CFR 30.15. Http://www.nrc.gov. March 13, 2001.
- \*NRC. 2001e. Byproduct material listing, exempt concentrations. Nuclear Regulatory Commission. Code of Federal Regulations. 10 CFR 30.70. <a href="http://www.nrc.gov"><u>Http://www.nrc.gov</u></a>. March 23, 2001.
- \*NRC. 2001f. Individual monitoring. Nuclear Regulatory Commission. Code of Federal Regulations. 10 CFR 20.2206. <a href="http://www.nrc.gov"><u>Http://www.nrc.gov</u></a>. April 6, 2001.
- \*NRC. 2001g. Labeling. Nuclear Regulatory Commission. Code of Federal Regulations. 10 CFR 30. <a href="http://www.nrc.gov"><u>Http://www.nrc.gov</u></a>. April 6, 2001.

# COBALT 9. REFERENCES 351

- \*NRC. 2001h. Medical use. Nuclear Regulatory Commission. Code of Federal Regulations. 10 CFR 35.400. <a href="http://www.nrc.gov"><u>Http://www.nrc.gov.</u></a> April 6, 2001.
- \*NRC. 2001i. Quantities of radioactive materials requiring labeling. Nuclear Regulatory Commission. Code of Federal Regulations. 10 CFR 20, Appendix C. <a href="http://www.nrc.gov"><u>Http://www.nrc.gov</u></a>. April 6, 2001.
- \*NRC. 2001j. Quantities of radioactive materials requiring need for an emergency plan. Nuclear Regulatory Commission. Code of Federal Regulations. 10 CFR 30.72. <a href="http://www.nrc.gov"><u>Http://www.nrc.gov.</u></a> April 13, 2001.
- \*NRC. 2001k. Radiation standards. Nuclear Regulatory Commission. Code of Federal Regulations. 10 CFR 20. Http://www.nrc.gov. April 13, 2001.
- \*NRC. 2001. Radioactive waste classification. Nuclear Regulatory Commission. Code of Federal Regulations. 10 CFR 61.55. Http://www.nrc.gov. June 7, 2001.
- \*Nriagu JO. 1989. A global assessment of natural sources of atmospheric trace metals. Nature 338:47-49
- Nriagu JO. 1992. Toxic metal pollution in Africa. Sci Total Environ 121:1-37.
- \*Nriagu JO, Coker RD. 1980. Trace metals in humic and fulvic acids from Lake Ontario sediments. Environ Sci Technol 14:443-446.
- \*NTP. 1991. NTP report on the toxicity studies of cobalt sulfate heptahydrate in F344/N rats and B6C3F1 mice (inhalation studies). National Institutes of Health, National Toxicology Program. NIH Publication No. 91-3124.
- \*NTP. 1998. NTP report on the toxicity studies of cobalt sulfate heptahydrate in F344/N rats and B6C3F1 mice (inhalation studies). National Institutes of Health, National Toxicology Program. NIH Publication No. 471.
- Numazawa S, Oguro T, Yoshida T, et al. 1989. Synergistic induction of rat hepatic ornithine decarboxylase by multiple doses of cobalt chloride. Chem Biol Interact 72:257-267.
- \*NYS Dept of Environmental Conservation. 2000. Memorandum: DAR-1 (air guide) AGC/SGC tables. Albany, NY: New York State Department of Environmental Conservation.
- Oanh NT, Bengtsson BE, Reutergardh L, et al. 1995. Levels of contaminants in effluent, sediment, and biota from Bai Bang, a bleached kraft pulp and paper mill in Vietnam. Arch Environ Contam Toxicol 29:506-516.
- O'Brien DJ, Kaneene JB, Poppenga RH. 1993. The use of mammals as sentinels for human exposure to toxic contaminants in the environment. Environ Health Perspect 99:351-368.
- \*Ogawa HI, Liu S-Y, Sakata K, et al. 1988. Inverse correlation between combined mutagenicity in Salmonella typhimurium and strength of coordinate bond in mixtures of cobalt(II) chloride and 4-substituted pyridines. Mutat Res 204:117-121.
- Ogawa HI, Ohyama Y, Ohsumi Y, et al. 1999. Cobaltous chloride-induced mutagenesis in the *sup*F tRNA gene of Escherichia coli. Mutagenesis 14(2):249-253.

# COBALT 352 9. REFERENCES

\*Ogawa HI, Sakata K, Inouye T, et al. 1986. Combined mutagenicity of cobalt(II) salt and heteroaromatic compounds in Salmonella typhimurium. Mutat Res 172:97-104.

Ogawa HI, Shibahara T, Iwata H, et al. 1994. Genotoxic activities in vivo of cobaltous chloride and other metal chlorides as assayed in the drosophila wing spot test. Mutat Res 320:133-140.

O'Hara GP, Mann DE, Gautieri RF. 1971. Effect of cobalt chloride and sodium cobaltinitite on the growth of established epithelial tumors induced by methylcholanthrene. J Pharm Sci 60(3):473-474.

Ohba S, Hiramatsu M, Edamatsu R, et al. 1994. Metal ions affect neuronal membrane fluidity of rat cerebral cortex. Neurochem Res 19(3):237-247.

\*Ohio EPA. 2000. Toxic release inventory. Air pollution regulations. <a href="http://www.epa.ohio.gov/dapc/regs/trirules.html">http://www.epa.ohio.gov/dapc/regs/trirules.html</a>. February 22, 2000.

Olivarius F, Menne T. 1992. Skin reactivity to metallic cobalt in patients with a positive patch test to cobalt chloride. Contact Dermatitis 27:241-243.

\*Olivero S, Villani P, Botta A. 1995. Genotoxic effects of cobalt chloride, sulfate and nitrate on cultured human lymphocytes. Med Sci Res 23:339-341.

Olmez I, Sheffield AE, Gordon GE, et al. 1988. Compositions of particles from selected sources in Philadelphia for receptor modeling applications. J Air Pollut Control Assoc 38(11):1392-1402.

Olsavszky R, Rycroft RJG, White IR, et al. 1998. Contact sensitivity to chromate: comparison at a London contact dermatitis clinic over a 10-year period. Contact Dermatitis 38:329-331.

\*Ondov JM, Zoller WH, Gordon GE. 1982. Trace element emissions on aerosols from motor vehicles. Environ Sci Technol 16:318-328.

Ondov JM, Choquette CE, Zoller WH, et al. 1989. Atmospheric behavior of trace elements on particles emitted from a coal-fired power plant. Atmos Environ 23(10):2193-2204.

Ong A, Li WX, Ling CC. 1993. Low-dose-rate irradiation of rat embryo cells containing the Ha-ras oncogene. Radiat Res 134:251-255.

\*Onkelinx C. 1976. Compartment analysis of cobalt (II) metabolism in rats of various ages. Toxicol Appl Pharmacol 38(425-438):425-438.

Onozuka M, Imai S. 1990. Induction of epileptic seizure activity by a specific protein from cobalt-induced epileptogenic cortex of rats. Brain Res 507:143-145.

OSHA. 1993. Air contaminants. Occupational Safety and Health Administration. Code of Federal Regulations. 29 CFR 1910.

OSHA. 1999a. Air contaminants. Occupational Safety and Health Administration. Code of Federal Regulations. 29 CFR 1910.1000.

OSHA. 1999b. Air contaminants. Occupational Safety and Health Administration. Code of Federal Regulations. 29 CFR 1915.1000.

# COBALT 353 9. REFERENCES

- OSHA. 1999c. Gases, vapors, fumes, dusts, and mists. Occupational Safety and Health Administration. Code of Federal Regulations. 29 CFR 1926.55.
- \*OSHA. 2001d. Construction industry, cobalt metal, dust, and fume. Occupational Safety and Health Administration. <a href="http://www.osha.gov/OshStd\_data/1926\_0055.html">http://www.osha.gov/OshStd\_data/1926\_0055.html</a>. June 7, 2001.
- \*OSHA. 2001e. General industry, cobalt metal, dust, and fume. Occupational Safety and Health Administration. <a href="http://www.osha.gov/OshStd\_data/1910\_0000.html">http://www.osha.gov/OshStd\_data/1910\_0000.html</a>. June 7, 2001.
- \*OSHA. 2001a. Ionizing radiation. Occupational Safety and Health Administration, U.S. Department of Labor. Code of Federal Regulations. 29 CFR 1910.1096. <a href="http://www.osha-slc.gov/OshStd"><u>Http://www.osha-slc.gov/OshStd</u></a> data/1910 1096.html. June 7, 2001.
- \*OSHA. 2001b. Safety and health regulations for construction. Ionizing radiation. Occupational Safety and Health Administration, U.S. Department of Labor. Code of Federal Regulations. 29 CFR 1926.53. <a href="http://www.osha-slc.gov/OshStd\_data/1926\_0053.html">http://www.osha-slc.gov/OshStd\_data/1926\_0053.html</a>. June 7, 2001.
- \*OSHA. 2001c. Shipyards, cobalt metal, dust, and fume. Occupational Safety and Health Administration. Http://www.osha.gov/OshStd data/1915 0000.html. June 7, 2001.
- \*Ostapczuk P, Froning M, Stoeppler M, et al. 1985. Square wave voltammetry: A new approach for the sensitive determination of nickel and cobalt in human samples. In: Brown SS, Sunderman FW, ed. Progress in nickel toxicology: Proceedings of the 3<sup>rd</sup> international conference on nickel metabolism and toxicology held in Paris 4-7 September 1984. Palo Alto, CA: Blackwell Scientific Publications, 129-132.
- \*Ostapczuk P, Valenta P, Rutzel H, et al. 1987. Application of differential pulse anodic stripping voltammetry to the determination of heavy metals in environmental samples. Sci Total Environ 60:1-16.
- Osuna Lopez JI, Zazueta-Padilla HM, Rodriguez-Higuera A, et al. 1990. Trace metal concentrations in mangrove oyster (Crassostrea corteziensis) from tropical lagoon environments, Mexico. Mar Pollut Bull 21(10):486-488.
- \*Outridge PM, Noller BN. 1991. Accumulation of toxic trace elements by freshwater vascular plants. Rev Environ Contam Toxicol 121:1-63.
- \*Owen GM, Brozek J. 1966. Influence of age, sex and nutrition on body composition during childhood and adolescence. In: Falkner F, ed. Human development. Philadelphia, PA: WB Saunders, 222-238.
- Owens PN, Walling DE, He Q. 1996. The behavior of bomb-derived caesium-137 fallout in catchment soils. J Environ Radioact 32(3):169-191.
- Paez-Osuna F, Marmolejo-Rivasa C. 1990a. Occurrence and seasonal variation of heavy metals in the oyster saccrostrea iridescens. Bull Environ Contam Toxicol 44:129-134.
- Paez-Osuna F, Marmolejo-Rivasa C. 1990b. Trace metals in tropical coastal lagoon bivalves crassostrea corteziensis. Bull Environ Contam Toxicol 45:538-544.
- \*Page NP, Ainsworth EJ, Leong GF. 1968. The relationship of exposure rate and exposure time to radiation injury in sheep. Radiat Res 33:94-106.

# COBALT 9. REFERENCES

- \*Painter RB, Howard R. 1982. The hela DNA-synthesis inhibition test as a rapid screen for mutagenic carcinogens. Mutat Res 92:427-437.
- Paksy K, Forgacs Z, Gati I. 1999. In vitro comparative effect of Cd<sup>2+</sup>, Ni<sup>2+</sup>, and Co<sup>2+</sup> on mouse postblastocyst development. Environmental Research (Section A) 80:340-347.
- \*Paley KR, Sobel ES, Yalow RS. 1958. Effect of oral and intravenous cobaltous chloride on thyroid function. J Clin Endocrinol Metab 18:850-859.
- \*Palit S, Ghosh AK, Sharma A, et al. 1991a. Modification of the clastogenic effects of cobalt by calcium in bone marrow cells of mice in vivo. Cytologia 56:373-377.
- \*Palit S, Sharma A, Talukder G. 1991b. Chromosomal aberrations induced by cobaltous chloride in mice in vivo. Biol Trace Elem Res 29:139-145.
- \*Palit S, Sharma A, Talukder G. 1991c. Cytotoxic effects of cobalt chloride on mouse bone marrow cells in vivo. Cytobios 65:85-89.
- \*Palit S, Sharma A, Talukder G. 1991d. Protection by chlorophyllin against induction of chromosomal aberrations by cobalt in bone marrow cells of mice in vivo. Fitoterapia 62:(5)425-428.
- \*Palko J, Yli-Halla M. 1988. Solubility of Co, Ni, and Mn in some extractants in a Finnish acid sulphate soil area. Acta Agric Scand 38:153-158.
- \*Palmes ED, Nelson N, Laskin S, et al. 1959. Inhalation toxicity of cobalt hydrocarbonyl. Am Ind Hyg Assoc J 20:453-468.
- Palmiter RD. 1994. Regulation of metallothionein genes by heavy metals appears to be mediated by a zinc-sensitive inhibitor that interacts with a constitutively active transcription factor, MTF-1. Proc Natl Acad Sci USA 91:1219-1223.
- \*Paternain JL, Domingo JL, Corbella J. 1988. Developmental toxicity of cobalt in the rat. J Toxicol Environ Health 24:193-200.
- Pathak SP, Kumar S, Ramteke PW, et al. 1992. Riverine pollution in some northern and northeastern states of India. Environ Monit Assess 22:227-236.
- \*Patrick G, Batchelor AL, Stirling C. 1989. An interspecies comparison of the lung clearance of inhaled monodisperse cobalt oxide particles- part VI: Lung clearance of inhaled cobalt oxide particles in SPF Fischer rats. J Aerosol Sci 20(2):249-255.
- Patrick G, Stirling C, Kreyling WG, et al. 1994. Interspecies comparison of the clearance of ionic cobalt from the lungs. Inhal Toxicol 6:225-240.
- Payan H. 1971. Morphology of cobalt experimental epilepsy in rats. Exp Mol Pathol 15:312-319.
- \*Payan HM, Conard JR. 1974. Cobalt-induced epilepsy in rats: A study in biochemical substances. Arch Pathol 97:170-172.
- Pedigo NG. 1994. Time course of cobalt toxicity in murine preimplantation embryos and dose responsive induction of metallothionein. Biol Reprod 50(Suppl. 1):89.

# COBALT 355 9. REFERENCES

\*Pedigo NG, Vernon MW. 1993. Embryonic losses after 10-week administration of cobalt to male mice. Reprod Toxicol 7:111-116.

\*Pedigo NG, George WJ, Anderson MB. 1988. Effects of acute and chronic exposure to cobalt in male reproduction in mice. Reprod Toxicol 2:45-53.

Peet MJ, Gregersen H, McLennan H. 1986. 2-Amino-5-phosphonovalerate and Co<sup>2+</sup> selectively block depolarization and burst firing of rat hippocampal CA1 pyramidal neurones by N-methyl-D-aspartate. Neuroscience 12(3):635-641.

\*Pehrsson SK, Hatori N, Clyne N, et al. 1991. The effect of chronic cobalt exposure on cardiac function in rats. Trace Elem Med 8(4):195-198.

\*Persson B, Carlenor E, Clyne N, et al. 1992. Binding of dietary cobalt to sarcoplasmic reticulum proteins. Scand J Clin Lab Invest 52:137-140.

Peryakov EA, Berliner LJ. 1994. Co<sup>2+</sup> binding to α-lactalbumin. J Protein Chem 13(3):277-281.

Pery-Man N, Houeto P, Coirault C, et al. 1996. Hydroxocobalamin vs cobalt toxicity on rat cardiac and diaphragmatic muscles. Intensive Care Med 22:108-115.

Pesch G, Reynolds B, Rogerson P. 1978. Trace metals in scallops from within and around two ocean disposal sites. Mar Pollut Bull 8(10):224-228.

\*Pettine M, Camusso M, Martinotti W, et al. 1994. Soluble and particulate metals in the Po River: factors affecting concentrations and partitioning. Sci Total Environ 145:243-265.

\*Philippe JV. 1975. Fertility and irradiation: A preconceptional investigation in teratology. Am J Obstet Gynecol 123(7):714-718.

Pinkerton BW, Brown KW. 1985. Plant accumulation and soil sorption of cobalt from cobalt-amended soils. Agron J 77:634-638.

Pisati G, Zedda S. 1994. Outcome of occupational asthma due to cobalt hypersensitivity. Sci Total Environ 150:167-171.

\*Pitkanen A, Saano V, Hyvonen K, et al. 1987. Decreased GABA, benzodiazepine, and picrotoxinin receptor binding in brains of rats after cobalt-induced epilepsy. Epilepsia 28:11-16.

\*Planinsek F, Newkirk JB. 1979. Cobalt and cobalt alloys. In: Kirk RE, Othmer DF, Grayson M, et al., ed. Kirk-Othmer encyclopedia of chemical technology. New York, NY: John Wiley and Sons, 481-494.

PNL. 2000. Hanford site environmental report for calendar year 1994. Richland, WA: Pacific Northwest National Laboratory. <a href="http://www.pnl.gov/env/toc.html">http://www.pnl.gov/env/toc.html</a>. March 17, 2000.

\*PNNL 1996. Hanford site environmental report for calendar year 1995. Richland, WA: Pacific Northwest National Laboratory. <a href="http://www.hanford.gov/docs/annualrp/1995/index.htm">http://www.hanford.gov/docs/annualrp/1995/index.htm</a>. February 12, 1996.

Polyak K, Bodog I, Hlavay J. 1994. Determination of chemical species if selected trace elements in fly ash. Talanta 41(7):1151-1159.

# COBALT 356 9. REFERENCES

Popov LN. 1977. An experimental study of the effects of low concentrations of metallic cobalt aerosols on the animal organism. Gig Sanit 4:97-98.

\*Potolicchio I, Festucci A, Hausler P, et al. 1999. HLA-DP molecules bind cobalt: a possible explanation for the genetic association with hard metal disease. Eur J Immunol 29:2140-2147.

\*Potolicchio I, Mosconi G, Forni A, et al. 1997. Susceptibility to hard metal lung disease is strongly associated with the presence of glutamate 69 in HLA-Dpβ chain. Eur J Immunol 27:2741-2743.

\*Poulsen OM, Christensen JM, Sabbioni E, et al. 1994. Trace element reference values in tissues from inhabitants of the European community. V. Review of trace elements in blood, serum and urine and critical evaluation of reference values for the Danish Population. Sci Total Environ 141:197-215.

Poulsen OM, Olsen E, Christensen JM, et al. 1995. Geltape method for measurement of work related surface contamination with cobalt containing dust: Correlation between surface contamination and airborne exposure. Occup Environ Med 52:827-833.

\*Prager D, Sembrot JT, Southard M. 1972. Cobalt-60 therapy of Hodgkin's disease and the subsequent development of hypothyroidism. Cancer 29(2):458-460.

Prangere T, Bowden AD, Beauchat V, et al. 1997. A study of the behavior of cobalt chloride, during the labeling of leukocytes with <sup>99</sup>Tc <sup>m</sup>- HMPAO stabilized in vitro by the addition of cobalt chloride solution. Nucl Med Commun 18:258-261.

\*Prescott E, Netterstrom B, Faber J, et al. 1992. Effect of occupational exposure to cobalt blue dyes on the thyroid volume and function of female plate painters. Scand J Work Environ Health 18:101-104.

Probst T, Zeh P, Kim J-I. 1995. Multielement determinations in ground water ultrafiltrates using inductively coupled plasma mass spectrometry and monostandard neutron activation analysis. Fresenius J Anal Chem 351:745-751.

Pruss RM, Akeson RL, Racke MM, et al. 1991. Agonist-activated cobalt uptake identifies divalent cation-permeable kainate receptors on neurons and glial cells. Neuron 7:509-518.

\*Pryce DW, King CM. 1990. Orofacial granulomatosis associated with delayed hypersensitivity to cobalt. Clin Exp Dermatol 15:384-386.

Pyatt FB. 1999. Comparison of foliar and stem bioaccumualtion of heavy metals by corsican pines in the Mount Olympus area of Cyprus. Ecotoxicol Environ Saf 42:57-61.

Que Hee SS, Finelli VN, Fricke FL, et al. 1982. Metal content of stack emissions, coal and fly ash from some eastern and western power plants in the U.S.A. as obtained by ICP-AES. Int J Environ Anal Chem 13:1-18.

Rae T. 1978. The haemolytic action of particulate metals (Cd, Cr, Co, Fe, Mo, Ni, Ta, Ti, Zn, Co-Cr alloy). J Pathol 125:81-89.

\*Raffn E, Mikkelsen S, Altman DG, et al. 1988. Health effects due to occupational exposure to cobalt blue dye among plate painters in a porcelain factory in Denmark. Scand J Work Environ Health 14:378-384.

# COBALT 357 9. REFERENCES

\*Raghavendran KV, Satbhai PD, Unnikrishnan K, et al. 1978. Long-term retention studies of 131I, 137Cs and 60Co in Indian workers. Health Phys 34:185-188.

Rainbow PS, White SL. 1990. Comparative accumulation of cobalt by three crustaceans: A decapod, an amphipod and barnacle. Aquat Toxicol 16:113-126.

Rakusan K, Rajhathy J. 1974. Oxygen affinity of blood in rats during cobalt-induced erythrocytic polycythemia and after its correction. Life Sci 15(1):23-28.

\*Rastogi SK, Gupta BN, Husain T, et al. 1991. A cross-sectional study of pulmonary function among workers exposed to multimetals in the glass bangle industry. Am J Ind Med 20:391-399.

Ratcliffe J, English JSC. 1997. Allergic contact dermatitis from cobalt in animal feed. Contact Dermatitis 39:201-202.

\*Rauscher AH, Bauchinger M. 1983. Chromosome aberrations induced in patients treated with chemotherapeutic drugs and irradiation for acute lymphatic leukemia. Hum Genet 64:73-79.

\*Raven KP, Loeppert RH. 1997. Trace element composition of fertilizers and soil amendments. J Environ Qual 26:551-557.

Ravichandran M, Baskaran M, Santschi PH, et al. 1995. History of trace metal pollution in Sabine-Neches Estuary, Beaumont, Texas. Environ Sci Technol 29:1495-1503.

Reagan EL. 1992a. Acute oral LD[-50] study in rats with cobalt (II) cabonate hydrate. J Am Coll Toxicol 11(6):687.

Reagan EL. 1992b. Acute oral LD[-50] study in rats with cobalt powder. J Am Coll Toxicol 11(6):686.

Reagan EL. 1992c. Acute oral LD[-50] study in rats with cobalt sulfate. J Am Coll Toxicol 11(6):688.

Reagan EL. 1992d. Acute oral toxicity study in rats with cobalt (II) sulfide. J Am Coll Toxicol 11(6):693.

Reddy PRK, Reddy SJ. 1997. Elemental concentrations in medicinally important leafy materials. Chemosphere 34(9/10):2193-2212.

\*Reimann C, DeCaritat P, Halleraker JH, et al. 1997. Rainwater composition in eight arctic catchments in northern Europe (Finland, Norway and Russia). Atmos Environ 31(2):159-170.

Remez VP, Sapozhnikov YA. 1996. The rapid determination of caesium radionuclides in water systems using composite sorbents. Appl Radiat Isot 47:885-886.

Remy Davee Guimarraes J. 1992. Bioaccumulation of <sup>137</sup>Cs and <sup>60</sup>Co by a tropical marine teleost Epinephelus sp. Sci Total Environ 120:205-212.

\*Rengasamy A, Kommineni C, Jones JA, et al. 1999. Effects of hard metal on nitric oxide pathways and airway reactivity to methacholine in rat lungs. Toxicol Appl Pharmacol 157:178-191.

Repetto G, Sanz P, Repetto M. 1995. Effects of cobalt on mouse neuroblastoma cells cultured in vitro. Toxicol in Vitro 9(4):375-379.

# COBALT 358 9. REFERENCES

Ressetar HG, Overman DO. 1987. Neurotoxicity of cobaltous chloride during myelination in the golden hamster brain. Anat Rec 218(1):113A.

\*Reuber S, Kreuzer M, Kirchgessner M. 1994. Interactions of cobalt and iron in absorption and retention. J Trace Elem Electrolytes Health Dis 8:151-158.

\*Reuff J, Bras A, Cristovao L, et al. 1993. DNA strand breaks and chromosomal aberrations induced by H2O2 and 60Co  $\tau$ -radiation. Mutat Res 289:197-204.

\*Reyners H, De Reyners EG, Poortmans F, et al. 1992. Brain atrophy after foetal exposure to very low doses of ionizing radiation. Int J Radiat Biol 62:(5)619-626.

\*Rezvani M, Heryet JC, Hopewell JW. 1989. Effects of single doses of gamma-radiation on pig lung. Radiother Oncol 14:132-142.

Rhoads K, Samders CL. 1985. Lung clearance, translocation, and acute toxicity of arsenic, beryllium, cadmium, cobalt, lead, selenium, vanadium and ytterbium oxides following deposition in rat lung. Environ Res 36:359-378.

\*Richardson HW. 1993. Cobalt compounds. In: Kroschwitz JI, Howe-Grant M, ed. Kirk-Othmer Encyclopedia of chemical technology. New York, NY: John Wiley & Sons, 778-793.

Richter H, Lorenz W, Bahadir M. 1997. Examination of organic and inorganic xenobiotics in equipped printed circuits. Chemosphere 35(1):169-179.

Ridout PS, Rainbow PS, Roe HSJ, et al. 1989. Concentrations of V, Cr, Mn, Fe, Ni, Co, Cu, Zn, As and Cd in mesopelagic crustaceans from the North East Atlantic Ocean. Mar Biol 100:465-471.

Rizzato G, Fraioli P, Sabbioni E, et al. 1994. The differential diagnosis of hard metal lung disease. Sci Total Environ 150:77-83.

\*Robbins MEC, Bywaters T, Rezvani M, et al. 1991a. Residual radiation-induced damage to the kidney of the pig as assayed by retreatment. Int J Radiat Biol 60:(6)917-928.

\*Robbins MEC, Campling D, Rezvani M, et al. 1989a. Nephropathy in the mature pig after the irradiation of a single kidney: A comparison with the mature pig. Int J Radiat Oncol Biol Phys 16:1519-1528.

\*Robbins MEC, Campling D, Rezvani M, et al. 1989b. Radiation nephropathy in mature pigs following the irradiation of both kidneys. Int J Radiat Biol 56:(1)83-98.

\*Robbins MEC, Campling D, Rezvani M, et al. 1989c. The effect of age and the proportion of renal tissue irradiated on the apparent radiosensitivity of the pig kidney. Int J Radiat Biol 6:(1)99-106.

\*Robbins MEC, Wooldridge MJA, Jaenke RS, et al. 1991b. A morphological study of radiation nephropathy in the pig. Radiat Res 126:317-327.

\*Roche M, Layrisse M. 1956. Effect of cobalt on the thyroidal uptake of I131. J Clin Endocrinol Metab 16:831-833.

Rodgers GM, George WJ, Fisher JW. 1972. Increased kidney cyclic AMP levels and erythropoietin production following cobalt administration. Proc Soc Exp Biol Med 140(3):977-981.

# COBALT 359 9. REFERENCES

Roesems G, Hoet PHM, Demedts M, et al. 1997. In vitro toxicity of cobalt and hard metal dust in rat and human type II pneumocytes. Pharmacol Toxicol 81:74-80.

\*Romaguera C, Lecha M, Grimalt F, et al. 1982. Photocontact dermatitis to cobalt salts. Contact Dermatitis 8:383-388.

Ronde P, Nichols RA. 1996. Uptake of cadmium and cobalt in rat brain synaptosomes in the absence of depolarization. J Neurochem 66(Suppl. 1):S51.

Rooney C, Beral V, Maconochie N, et al. 1993. Case-control study of prostatic cancer in employees of the United Kingdom Atomic Energy Authority. Br Med J 307(6916):1391-1397.

\*Roscher AA, Woodard JS. 1969. Fatal gastrointestinal complications following cobalt therapy for carcinoma of the uterine cervix. Int Surg 51(6):526-536.

\*Rosenberg DW. 1993. Pharmacokinetics of cobalt chloride and cobalt-protoporphyrin. Drug Metab Dispos 21(5):846-849.

\*Rossmann R, Barres J. 1988. Trace element concentrations in near-surface waters of the Great Lakes and methods of collection storage, and analysis. J Great Lakes Res 14(2):188-204.

\*Roswit B, White DC. 1977. Severe radiation injuries of the lung. AJR Am J Roentgenol 129:(1)127-136

Roto P. 1980. Asthma, symptoms of chronic bronchitis and ventilatory capacity among cobalt and zinc production workers. Scand J Work Environ Health 6(Suppl. 1):1-49.

\*Roy PE, Bonenfant JT, Turcot L. 1968. Thyroid changes in cases of Quebec beer drinkers myocardosis. Am J Clin Pathol 50:234-239.

Roy WR. 1994. Groundwater contamination from municipal landfills in the USA. In: Adriano DC, ed. Contamination of groundwaters: Case studies. Northwood, UK: Scientific Review, 411-446.

\*Rubin ES. 1999. Toxic releases from power plants. Environ Sci Technol 33:3062-3067.

\*Rueff J, Bras A, Cristovao L, et al. 1993. DNA strand breaks and chromosomal aberrations induces by  $H_2O2$  and  $^{60}Co$   $\gamma$ -radiation. Mutation Research. 289:197-204.

\*Ruokonen E-L, Linnainmaa M, Seuri M, et al. 1996. A fatal case of hard-metal disease. Scand J Work Environ Health 22:62-65.

\*Russell-Jones GJ, Alpers DH. 1999. Vitamin B12 transporters. In: Amidon GL, Sadee W, ed. Pharmaceutical biotechnology. New York, NY: Kluwer Academic/Plenum Publishers, 493-520.

\*Rystedt I, Fischer T. 1983. Relationship between nickel and cobalt sensitization in hard metal workers. Contact Dermatitis 9:195-200.

Saad AY, Abdelazim AA, El-Khashab MM, et al. 1991. Effects of gamma radiation on incisor development of the prenatal albino mouse. J Oral Pathol Med 20:385-388.

Sadiq M, Zaidi TH. 1994. Sediment composition and metal concentrations in mangrove leaves from the Saudi coast of the Arabian Gulf. Sci Total Environ 155:1-8.

# COBALT 360 9. REFERENCES

Sadiq M, Mian AA, Althagafi KM. 1992. Inter-city comparison of metals in scalp hair collected after the Gulf War 1991. J Environ Sci Health Part A 27(6):1415-1431.

\*Saker F, Ybarra J, Leahy P, et al. 1998. Glycemia-lowering effect of cobalt chloride in the diabetic rat: role of decreased gluconeogenesis. Am J Physiol 274:E984-E991.

Sala C, Mosconi G, Bacis M, et al. 1994. Cobalt exposure in 'hard metal' and diamonds grinding tools manufacturing and in grinding processes. Sci Total Environ 150:111-116.

Salmi HA, Lindgren I. 1969. Retention of cobalt in experimentally induced kidney disease. Acta Radiologica Therapy Physics Biology 8(3):208-214.

\*Saltzman BE, Keenan RG. 1957. Microdetermination of cobalt in biological materials. Methods Biochem Anal 5:181-223.

Sanchez JH, Abernethy DJ, Boreiko CJ. 1987. Lack of di-(2-ethyhhexyl) phthalate activity in the C3H/10T1/2 cell transformation system. Toxicol in Vitro 1(1):49-53.

\*Sanudo-Wilhelmy SA, Flegal AR. 1996. Trace metal concentrations in the surf zone and in coastal waters off Baja California, Mexico. Environ Sci Technol 30:1575-1580.

\*Sanyal B, Pant GC, Subrahmaniyam K, et al. 1979. Radiation myelopathy. J Neurol Neurosurg Psychiatry 42:413-418.

\*Sarkar B. 1995. Metal replacement in DNA-binding zinc finger proteins and its relevance to mutagenicity and carcinogenicity through free radical generation. Nutrition 11(5):646-649.

Sasame HA, Boyd MR, Mitchell JR, et al. 1977. Increased tissue levels of reduced glutathione produced by cobaltous chloride. Fed Proc 36:405.

Satoh-Kamachi A, Munakata M, Kusaka Y, et al. 1998. A case of sarcoidosis that developed three years after the onset of hard metal asthma. Am Ind Hyg Assoc J 33:379-383.

Scanes P. 1996. Oyster watch: Monitoring trace metal and organochlorine concentrations in Sydney's coastal waters. Mar Pollut Bull 33(7-12):226-238.

\*Scansetti G, Botta GC, Spinelli P, et al. 1994. Absorption and excretion of cobalt in the hard metal industry. Sci Total Environ 150:141-144.

\*Scansetti G, Lamon S, Talarico S, et al. 1985. Urinary cobalt as a measure of exposure in the hard metal industry. Int Arch Occup Environ Health 57:19-26.

Scansetti G, Maina G, Botta GC, et al. 1998. Exposure to cobalt and nickel in the hard-metal production industry. Int Arch Occup Environ Health 71:60-63.

\*Schade SG, Felsher BF, Bernier GM, et al. 1970. Interrelationship of cobalt and iron absorption. J Lab Clin Med 75:435-441.

Schaeffer J, El-Mahdi AM, Peeples WJ. 1977. Treatment of intraperitoneal implants in mice using <sup>32</sup>P or <sup>60</sup>Co. Int J Nucl Med Biol 4:77-79.

# COBALT 9. REFERENCES 361

Schaller H, Neeb R. 1987. Gas-chromatographic elemental analysis via di(triflouroethyl)dithiocarbamato-chelates: X. Capillary gas chromatography at the pg-level - determination of Co and Cr[VI] besides Cr[III] in river water. Fresenius Z Anal Chem 327:170-174.

Schimmel RJ. 1978. Calcium antagonists and lipolysis in isolated rat epididymal adipocytes: Effects of tetracaine, manganese, cobaltous and lanthium ions and D600. Horm Metab Res 10:128-134.

\*Schmidt SL, Lent R. 1987. Effects of prenatal irradiation on the development of cerebral cortex and corpus callosum of the mouse. J Comp Neurol 264:193-204.

\*Schnitzer M. 1969. Reactions between fulvic acid, a soil humic compound and inorganic soil constituents. Soil Sci Soc Am Proc 33:75-81.

\*Schramel P. 1989. Determination of some additional trace elements in certified standard reference materials (soils, sludges, sediment) by ICP-emission spectrometry. Fresenius J Anal Chem 333:203-210.

\*Schroeder WH, Dobson M, Kane DM, et al. 1987. Toxic trace elements associated with airborne particulate matter: A review. J Air Pollut Control Assoc 37(11):1267-1285.

\*Schull WJ, Otake M, Yoshimaru H. 1988. Effect on intelligence test score of prenatal exposure to ionizing radiation in Hiroshima and Nagasaki: A comparison of the T65DR and DS86 dosimetry systems.

Schulman HM, Ponka P. 1981. The stimulation of globin synthesis by cobalt in reticulocytes with inhibited heme synthesis. Biochim Biophys Acta 654:166-168.

\*Schultz PN, Warren G, Kosso C, et al. 1982. Mutagenicity of a series of hexacoordinate cobalt(III) compounds. Mutat Res 102:393-400.

Schuster SJ, Badiavas EV, Costa-Giomi P, et al. 1989. Stimulation of erythropoietin during hypoxia and cobalt exposure. Blood 73(1):13-16.

Schwartz JL, Giovanazzi SM, Karrison T, et al. 1988. 2-[(Aminopropyl)amino] ethanethiol-mediated reductions in  $^{60}$ Co  $\gamma$ -ray and fission-spectrum neutron-induced chromosome damage in V79 cells. Radiat Res 113:145-154.

Schwartzkroin PA, Shimada Y, Bromley B. 1977. Recordings from cortical epileptogenic foci induced by cobalt iontophoresis. Exper Neurol 55:353-367.

\*Schweitzer DJ, Benjamin SA, Lee AC. 1987. Retinal dysplasia and progressive atrophy in dogs irradiated during ocular development. Radiat Res 111:340-353

\*Searl AG, Beechey CV, Green D, et al. 1976. Cytogenetic effects of protracted exposures to alphaparticles from plutonium-239 and to gamma-rays from cobalt-60 compared in male mice. Mutat Res 41:297-310.

\*Searl AG, Beechey CV, Green D, et al. 1980. Comparative effects of protracted exposures to  $^{60}$ Co $\gamma$ -radiation and  $^{239}$ Pu  $\alpha$ -radiation on breeding performance in female mice. Int J Radiat Biol 37:(2)189-200.

\*Sedlet J, Robinson J, Fairman W. 1958. A cobalt and a tritium incident at Argonne National Laboratory. In: Proceedings of the bio-assay and analytical chemistry annual meeting, 101-106.

# COBALT 362 9. REFERENCES

- \*Seed TM, Carnes BA, Tolle DV, et al. 1989. Blood responses under chronic low daily dose gamma irradiation: Differential preclinical responses of irradiated male dogs in progression to either aplastic anemia or myeloproliferative disease. Leukemia Research 13:(12)1069-1084.
- Seghizzi P, D'Adda F, Borleri D, et al. 1994. Cobalt myocardiopathy. A critical review of literature. Sci Total Environ 150:105-109.
- \*Seidenberg JM, Anderson DG, Becker RA. 1986. Validation of an in vivo developmental toxicity screen in the mouse. Teratogenesis Carcinog Mutagen 6:361-374.
- \*Semenza GL, Roth PH, Fang H-M, et al. 1994. Transcriptional regulation of genes encoding glycolytic enzymes by hypoxia-inducible factor 1. J Biol Chem 269(38):23757-23763.
- \*Sesana G, Cortona G, Baj A, et al. 1994. Cobalt exposure in wet grinding of hard metal tools for wood manufacture. Sci Total Environ 150:117-119.
- \*Setchell BP, Waites GMH. 1975. The blood-testis barrier. In: Creep RO, Astwood EB, Geiger SR, eds. Handbook of physiology: Endocrinology V. Washington, DC: American Physiological Society.
- \*Shabaan AA, Marks V, Lancaster MC, et al. 1977. Fibrosarcomas induced by cobalt chloride (CoCl<sub>2</sub>) in rats. Lab Anim 11:43-46.
- \*Sheets RW. 1998. Release of heavy metals from European and Asian porcelain dinnerware. Sci Total Environ 212:107-113.
- \*Sheline GE, Chaikoff IL, Montgomery ML. 1945. The elimination of administered cobalt in pancreatic juice and bile of the dog, as measured with its radioactive isotopes. Am J Physiol 145:285-290.
- Sheridan PJ, Zoller WH. 1989. Elemental composition of particulate material sampled from the Arctic haze aerosol. J Atmos Chem 9:363-381.
- Shibuya M, Fariello R, Farley IJ, et al. 1978. Cobalt injections into the substantia nigra of the rat: Effects on behavior and dopamine metabolism in the striatum. Exper Neurol 58:486-499.
- \*Shine JP, Ika RV, Ford TE. 1995. Multivariate statistical examination of spatial and temporal patterns of heavy metal equipment in New Bedford Harbor marine sediments. Environ Sci Technol 29:1781-1788.
- Shirakawa T, Morimoto K. 1997. Interplay of cigarette smoking and occupational exposure on specific immunoglobulin E antibodies to cobalt. Arch Env Health 52(2):124-128.
- \*Shirakawa T, Kusaka Y, Fujimura N, et al. 1988. The existence of specific antibodies to cobalt in hard metal asthma. Clin Allergy 18:451-460.
- \*Shirakawa T, Kusaka Y, Fujimura N, et al. 1989. Occupational asthma from cobalt sensitivity in workers exposed to hard metal dust. Chest 95(1):29-37.
- Shirakawa T, Kusaka Y, Fujimura N, et al. 1990. Hard metal asthma: Cross immunological and respiratory reactivity between cobalt and nickel? Thorax 45:267-271.
- Shirakawa T, Kusaka Y, Morimoto K. 1992a. Combined effect of smoking habits and occupational exposure to hard metal on total IgE antibodies. Chest 101(6):1569-1576.

# COBALT 363 9. REFERENCES

- Shirakawa T, Kusaka Y, Morimoto K. 1992b. Specific IgE antibodies to nickel in workers with known reactivity to cobalt. Clin Exp Allergy 22:213-218.
- Shoji S, Watanabe H, Komatsu K. 1998. Teratogenic effects of <sup>60</sup>Co γ-rays irradiation on the embryonic development of the scid mice and CB-17 mice. Teratology 57:230.
- \*Shrivastava VK, David CV, Khare N, et al. 1996. Cobalt chloride induced histopathological changes in thyroid gland of female mice, Mus musculus (P.). Pollut Res 15(3):307-309.
- \*Simesen M. 1939. The fate of cobalt after oral administration of metallic cobalt and subcutaneous injection of carbonatotetraminecobalt chloride, with remarks on the quantitative estimation of cobalt in organic materials. Arch Int Pharmacodyn 62:347-356.
- Sinclair JF, Sinclair PR, Healey JF, et al. 1982. Decrease in hepatic cytochrome P-450 by cobalt. Biochem J 204:103-109.
- \*Sinclair P, Gibbs AH, Sinclair JF, et al. 1979. Formation of cobalt protoporphyrin in the liver of rats. Biochem J 178:529-538.
- \*Singh I. 1983. Induction of reverse mutation and mitotic gene conversion by some metal compounds in Saccharomyces cerevisiae. Mutat Res 117:149-152.
- \*Singh PP, Junnarkar AY. 1991. Behavioral and toxic profile of some essential trace metal salts in mice and rats. Indian J Pharmacol 23:153-159.
- \*Smith IC, Carson BL. 1979. Trace metals in the environment. Ann Arbor, MI: Ann Arbor Science Publishers.
- \*Smith IC, Carson BL. 1981. Trace metals in the environment. Ann Arbor, MI: Ann Arbor Science Publishers.
- \*Smith RJ. 1972. I. The effect of cobalt on hydrolase activity in kidney and plasma and its relationship to erythropoietin production. II. Structure activity relationships of several protein and polypeptide potentiators of bradykinin action on rat uterus. Diss Abstr Int B 32(10):6132.
- Smith RJ, Fisher JW. 1976. Neutral protease activity and erythropoietin production in the rat after cobalt administration. J Pharmacol Exp Ther 197(3):714-722.
- Smith RP. 1969. Cobalt salts: Effects in cyanide and sulfide poisoning and on methemoglobinemia. Toxicol Appl Pharmacol 15:505-516.
- \*Smith T, Edmonds CJ, Barnaby CF. 1972. Absorption and retention of cobalt in man by whole-body counting. Health Phys 22:359-367.
- Sonnhof U, Krupp J, Claus H. 1990. The cobalt-epilepsy, a phenomenon of a modified sodium channel. Pflugers Arch 415(Suppl. 1):R87.
- Soon YK, Bates TE. 1985. Molybdenum, cobalt and boron uptake from sewage-sludge-amended soils. Can J Soil Sci 65:507-517.
- Sora S, Carbone MLA, Pacciarini M, et al. 1986. Disomic and diploid meiotic products induced in Saccharomyces cerevisiae by the salts of 27 elements. Mutagenesis 1(1):21-28.

# COBALT 9. REFERENCES

\*Sorbie J, Olatunbosun D, Corbett WEN, et al. 1971. Cobalt excretion test for the assessment of body iron stores. Can Med Assoc J 104(9):777-782.

\*Speijers GJA, Krajnc EI, Berkvens JM, et al. 1982. Acute oral toxicity of inorganic cobalt compounds in rats. Food Chem Toxicol 20:311-314.

Spiegel SJ, Farmer JK, Garver SR. 1985. Heavy metal concentrations in municipal wastewater treatment plant sludge. Bull Environ Contam Toxicol 35:38-43.

Sprince NL, Oliver LC, Chamberlin RI, et al. 1987. Exposure to cobalt and interstitial lung disease in tungsten carbide production workers. Am Rev Respir Dis 135:A20.

\*Sprince NL, Oliver LC, Eisen EA, et al. 1988. Cobalt exposure and lung disease in tungsten carbide production: A cross-sectional study of current workers. Am Rev Respir Dis 138:1220-1226.

SRI. 1989. 1989 Directory of chemical producers: United States of America. Menlo Park, CA: Stanford Research Institute International, 535-537.

\*SRI. 1999. 1999 Directory of chemical producers: United States of America. Menlo Park, CA: Stanford Research Institute International, 529-531.

Srivastava AK, Gupta BN, Mathur N, et al. 1991. An investigation of metal concentrations in blood of industrial workers. Vet Hum Toxicol 33(3):280-282.

\*Stanley AJ, Hopps HC, Shideler AM. 1947. Cobalt polycythemia. II. Relative effects of oral and subcutaneous administration of cobaltous chloride. Proc Soc Exp Biol Med 66:19-20.

\*Stavem P, Brogger A, Devik F, et al. 1985. Lethal acute gamma radiation accident at Kjeller, Norway. Acta Radiologica Oncology 24:61-80.

\*Stebbins AI, Horstman SW, Daniell WE, et al. 1992. Cobalt exposure in a carbide tip grinding process. Am Ind Hyg Assoc J 53(3):186-192.

Steel LK, Sweedler IK, Catravas GN. 1983. Effects of 60Co radiation on synthesis of prostaglandins  $F2\alpha$ , E, and thromboxane B2 in lung airways of guinea pigs. Radiat Res 94:156-165.

Steinhoff D, Mohr U. 1991. On the question of a carcinogenic action of cobalt-containing compounds. Exp Pathol 41:169-174.

Stephenson T, Lester JN. 1987a. Heavy metal behavior during the activated sludge process I. Extent of soluble and insoluble metal removal. Sci Total Environ 63:199-214.

Stephenson T, Lester JN. 1987b. Heavy metal behavior during the activated sludge process II. Insoluble metal removal mechanisms. Sci Total Environ 63:215-230.

\*Stokinger HE. 1981. The metals. In: Clayton GD, Clayton FE, ed. Patty's industrial hygiene and toxicology. New York, NY: John Wiley and Sons, 1493-1619.

\*Stokinger HE, Wagner WD. 1958. Early metabolic changes following cobalt exposure. Arch Ind Health 17:273-279.

# COBALT 365 9. REFERENCES

\*Stutz DR, Janusz SJ. 1988. Hazardous materials injuries: A handbook for pre-hospital care. 2nd ed. Beltsville, MD: Bradford Communications Corp.

Suardi R, Belotti L, Ferrari MT, et al. 1994. Health survey of workers occupationally exposed to cobalt. Sci Total Environ 150:197-200.

\*Sugaya E, Ishige A, Sediguchi K, et al. 1988. Damage of hippocampal neurons caused by cobalt focus in the cerebral cortex of rats. Brain Res 459:196-199.

Sugimoto T, Itoh K, Yasui Y, et al. 1985. Coexistence of neuropeptides in projection neurons of the thalamus in the cat. Brain Res 347:381-384.

\*Sun LC, Clinton JH, Kaplan E, et al. 1997. <sup>137</sup>Cs exposure in the Marshallese populations: An assessment based on whole-body counting measurements (1989-1994). Health Phys 73:86-99.

\*Sunderman WF. 1987. Metal induction of heme oxygenase. Ann N Y Acad Sci 514:65-80.

\*Sunderman FW, Zaharia O. 1988. Hepatic lipid peroxidation in CoCl<sub>2</sub>-treated rats, evidenced by elevated concentrations of thiobarbituric acid chromogens. Res Commun Chem Pathol Pharmacol 59(1):69-78.

Sunderman FW, Hopfer SM, Swift T, et al. 1989. Cobalt, chromium, and nickel concentrations in body fluids of patients with porous-coated knee or hip prostheses. J Orthop Res 7(3):307-315.

\*Suzuki K, Takahashi M, Ishii-Ohba H, et al. 1990. Steroidogenesis in the testes and the adrenals of adult male rats after γ-irradiation in utero at late pregnancy. J Steroid Biochem 35(2):301-305.

\*Suzuki Y, Shimizu H, Nagae Y, et al. 1993. Micronucleus test and erythropoiesis: Effect of cobalt on the induction of micronuclei by mutagens. Environ Mol Mutagen 22:101-106.

\*Swanson JL. 1984. Mobility of organic complexes of nickel and cobalt in soils. Department of Energy, Washington, DC. NTIS/DE830178997.

\*Sweeney WT, Elzay RP, Levitt SH. 1977. Histologic effect of fractionated doses of selectively applied <sup>60</sup>Co irradiation on the teeth of albino rats. J Dent Res 56(11):1403-1407.

\*Sweet CW, Vermette SJ, Landsberger S. 1993. Sources of toxic trace elements in urban air in Illinois. Environ Sci Technol 27:2502-2510.

\*Swennen B, Buchet J-P, Stanescu D, et al. 1993. Epidemiological survey of workers exposed to cobalt oxides, cobalt salts, and cobalt metal. Br J Ind Med 50:835-842.

Sypert GW, Bidgood WD. 1977. Effect of intracellular cobalt ions in postsynaptic inhibition in cat spinal motoneurons. Brain Res 134:372-376.

Szakmary E, Morvai V, Naray N, et al. 1992. The pre- and perinatal offspring damaging effect of cobalt. Reprod Toxicol 6:188-189.

Szakmary E, Ungvary G, Naray M, et al. 1989. Harmful effects of heavy metals (chromium, nickel, cobalt) on offspring. Teratology 40(3):298-299.

# COBALT 366 9. REFERENCES

\*Szebeni J, Garcia R, Eskelson CD, et al. 1989. The organ distribution of liposome-encapsulated and free cobalt in rats. Liposomes decrease the cardiac uptake of the metal. Life Sci 45:729-736.

Szefer P, Ikuta K, Kushiyama S, et al. 1997. Distribution of trace metals in the Pacific oyster, Crassostrea gigas, and crabs from the East Coast of Kyushu Island, Japan. Bull Environ Contam Toxicol 58:108-114.

\*Szefer P, Penpkowiak J, Skwarzec B, et al. 1993. Concentration of selected metals in penguins and other representative fauna of the Antarctica. Sci Total Environ 138:281-288.

\*Szefer P, Szefer K, Glasby GP, et al. 1996. Heavy-metal pollution in surficial sediments from the southern Baltic Sea off Poland. J Environ Sci Health Part A 31(10):2723-2754.

Szefer P, Szefer K, Skwarzec B. 1990. Distribution of trace metals in some representative fauna of the Southern Baltic. Mar Pollut Bull 21(2):60-62.

Szliska C, Raskoski J. 1990. Sensitization to nickel, cobalt and chromium in surgical patients. Contact Dermatitis 23:378-379.

\*Tabatowski K, Roggli VL, Fulkerson WJ, et al. 1988. Giant cell interstitial pneumonia in a hard-metal worker: Cytologic, histologic and analytical electron microscopic investigation. Acta Cytol 32(2):240-246

\*Takagi Y, Matsuda S, Imai S, et al. 1986. Trace elements in human hair: An international comparison. Bull Environ Contam Toxicol 36:793-800.

\*Takagi Y, Matsuda S, Imai S, et al. 1988. Survey of trace elements in human nails: An international comparison. Bull Environ Contam Toxicol 41:690-695.

\*Talbot RJ, Morgan A. 1989. An interspecies comparison of the lung clearance of inhaled monodisperse cobalt oxide particles- part VIII: Lung clearance of inhaled cobalt oxide particles in mice. J Aerosol Sci 20(2):261-265.

Talbot V. 1983. Lead and other trace metals in the sediments and selected biota of Princess Royal Harbour, Albany, Western Australia. Environ Pollut Ser B 5:35-49.

Tandon L, Iyengar GV, Parr RM. 1998. A review of radiologically important trace elements in human bones. Appl Radiat Isot 8:903-910.

Tanizaki Y, Shimokawa T, Yamazaki M. 1992. Physico-chemical speciation of trace elements in urban streams by size fractionation. Water Res 26(1):55-63.

Taubman SB, MAlnick JW. 1975. Inability of Ni<sup>++</sup> and Co<sup>++</sup> to release histamine from rat peritoneal mast cells. Res Commun Chem Pathol Pharmacol 10(2):383-386.

Taylor A, Marks V. 1978. Cobalt: a review. J Hum Nutr 32:165-177.

\*Taylor A, Marks V, Shabaan AA, et al. 1977. Cobalt induced lipaemia and erthropoiesis. Dev Toxicol Environ Sci 1:105-108.

\*Taylor DM. 1962. The absorption of cobalt from the gastro-intestinal tract of the rat. Phys Med Biol 6:445-451.

# COBALT 9. REFERENCES 367

\*Taylor JJ. 1996. Nuclear reactors. Safety in nuclear power facilities. In: Kroschwitz JI, Howe-Grant M, ed. Kirk-Othmer Encyclopedia of chemical technology. New York, NY: John Wiley & Sons, Vol. 17 473-507.

Tephly TR, Hibbeln P. 1971. The effect of cobalt chloride administration on the synthesis of hepatic microsomal cytochrome P-450. Biochem Biophys Res Commun 42(4):589-595.

\*Teraoka H. 1981. Distribution of 24 elements in the internal organs of normal males and the metallic workers in Japan. Arch Env Health 36(4):155-165.

Thaw CN, Raaka EG, Gershengorn MC. 1984. Evidence that cobalt ion inhibition of prolactin secretion occurs at an intracellular locus. Am J Physiol 247(3):C150-C155.

Theis TL, Young TC, Huang M, et al. 1994. Leachate characteristics and composition of cyanide-bearing wastes from manufactured gas plants. Environ Sci Technol 28:99-106.

\*Thibadoux GM, Pereira WV, Hodges JM, et al. 1980. Effects of cranial radiation n hearing in children with acute lymphocytic leukemia. J Pediatr 96(3):403-406.

Thomas RAP, Lawlor K, Bailey M, et al. 1998. Biodegradation of metal-EDTA complexes by an enriched microbial population. Appl Environ Microbiol 64(4):1319-1322.

\*Thomas RG, Furchner JE, London JE, et al. 1976. Comparative metabolism of radionuclides in mammals-x. Retention of tracer-level cobalt in the mouse, rat, monkey, and dog. Health Phys 31:323-333.

Thompson LJ, Ebel JG, Manzell KL, et al. 1995. Analytical survey of elements in veterinary college incinerator ashes. Chemosphere 30(4):807-811.

Thomson ABR, Valberg LS, Sinclair DG. 1971. Competitive nature of the intestinal transport mechanism for cobalt and iron in the rat. J Clin Invest 50:2384-2394.

Tian L, Lawrence DA. 1996. Metal-induced modulation of oxide production in vitro by murine macrophages: Lead, nickel, and cobalt utilize different mechanisms. Toxicol Appl Pharmacol 141:540-547.

Tilsley DA, Rostein H. 1980. Sensitivity caused by internal exposure to nickel, chrome and cobalt. Contact Dermatitis 6:175-178.

\*Tinsley DA, Baron AR, Critchley R, et al. 1983. Extraction procedures for atomic absorption spectrometric analysis of toxic metals in urban dust. Int J Environ Anal Chem 14:285-298.

\*Tipping E, Lofts S, Lawlor AJ. 1998. Modelling the chemical speciation of trace metals in the surface waters of the Humber system. Sci Total Environ 210/211:63-77.

\*Tolle DA, Arthur MF, Van Voris P. 1983. Microcosm/field comparison of trace element uptake in crops grown in fly ash-amended soil. Sci Total Environ 31:243-261.

Tolle DV, Fritz TE, Norris WP. 1977. Radiation-induced erythroleukemia in the beagle dog. Am J Pathol 87(3):499-510.

# COBALT 368 9. REFERENCES

Tom DJ, Rodgers PA, Shokoohi V, et al. 1996. Hepatic heme oxygenase is inducible in neonatal rats during the early postnatal period. Pediatr Res 40(2):288-293.

Tonna EA, Pavelec M. 1970. Changes in the proliferative activity of young and old mouse skeletal tissues following Co60 whole-body irradiation. J Gerontol 25(1):9-16.

Toran L. 1994. Radionuclide contamination in groundwater: Is there a problem? In: Environmental science and pollution control. Groundwater contamination and control. New York, NY: Dekker, M, 437-455.

Torre FD, Cassani M, Segale M, et al. 1990. Trace metal lung diseases: A new fatal case of hard metal pneumoconiosis. Respiration 57:248-253.

Tossavainen A, Jaakkola J. 1994. Occupational exposure to chemical agents in Finland. Appl Occup Environ Hyg 9(1):28-31.

\*Toste AP, Kirby LJ, Pahl TR. 1984. Role of organics in the subsurface migration of radionuclides in groundwater. In: Barney GS, Navratil JD, Schulz WW, ed. Geochemical behavior of disposed radioactive waste. Washington, DC: American Chemical Society, 251-270.

\*TRI98. 2000. National Library of Medicine, National Toxicology Information Program, Bethesda, MD. Http://www.epa.gov/triexplorer/chemical.htm. June 12, 2000.

\*TRI99. 2001. TRI explorer: Providing access to EPA's toxics release inventory data. Washington, DC: Office of Information Analysis and Access, Offices of Environmental Information, U.S. Environmental Protection Agency. Toxic Release Inventory. <a href="http://www.epa.gov/triexplorer/">http://www.epa.gov/triexplorer/</a>. June 7, 2001.

\*Trocine RP, Trefry JH. 1996. Metal concentrations in sediment, water and clams from the Indian River Lagoon, Florida. Mar Pollut Bull 32(10):754-759.

\*Tso W-W, Fung W-P. 1981. Mutagenicity of metallic cations. Toxicol Lett 8:195-200.

\*Tuchsen F, Jensen MV, Villadsen E, et al. 1996. Incidence of lung cancer among cobalt-exposed women. Scand J Work Environ Health 22:444-450.

Uchiyama M, Shiraishi Y, Akiba S. 1980. Kinetics of inhaled <sup>54</sup>Mn and <sup>60</sup>Co after an accidental human exposure. DOE Symp Ser 53:162-176.

Ueda T, Nakahara M, Nakamura R, et al. 1985. Accumulation of <sup>60</sup>Co by marine organisms under resuction of radioactivity in sea water. Bull Jpn Soc Sci Fish 51(11):1811-1816.

\*USAEC. 1973. Environmental levels of radioactivity Atomic Energy Commission installations. 1. National reactor testing stations, January-December 1970. Radiation Data and Reports 14:762-774.

\*USAEC. 1974a. Environmental levels of radioactivity Atomic Energy Commission installations. 1. Argonne National Laboratory, January-December 1972. Radiation Data and Reports 15:518-537.

\*USAEC. 1974b. Environmental levels of radioactivity Atomic Energy Commission installations. 1. Hanford atomic products operations, January-December 1971. Radiation Data and Reports 15:356-373.

\*USC. 1999. Hazardous air pollutants. United States Code. 42 USC 7412.

# COBALT 369 9. REFERENCES

- \*USC. 2001a. Hazardous air pollutants, cobalt compounds. United States Code. 42USC7412. <a href="http://www.4.law.cornell.edu.">http://www.4.law.cornell.edu.</a> June 18, 2001.
- \*USC. 2001b. Exemption of tax imposed on recycled cobalt. United States Code. 26USC4662. <a href="http://www.4.law.cornell.edu">http://www.4.law.cornell.edu</a>. June 18, 2001.
- \*USC. 2001c. Superfund, imposition of taxes. United States Code. 26USC4661. <a href="http://www.4.law.cornell.edu"><u>Http://www.4.law.cornell.edu</u></a>. June 18, 2001.
- \*USGS. 1998. Cobalt. U.S. Geological Survey Mineral Information 1998 by Kim B. Shedd. Http://minerals.usgs.gov/minerals/pubs/commodity/cobalt/210498.pdf. March 7, 1998.
- \*USGS. 1999. Cobalt. U.S. Geological Survey Mineral Information 1999 by Kim B. Shedd. Http://minerals.usgs.gov/minerals/pubs/commodity/cobalt/210499.pdf. April 13, 1999.
- \*USGS. 2000. Mineral Commodity Summaries 1999. Cobalt. U.S. Geological Survey. <u>Http://minerals.usgs.gov/minerals/pubs/commodity/cobalt/210300.pdf.0</u> February 3, 2000.
- \*USGS. 2001. Mineral Commodity Summaries 2000. Cobalt. U.S. Geological Survey. Http://minerals.usgs.gov/minerals/pubs/commodity/cobalt/210301.pdf. June 7, 2001.
- \*Valberg LS, Ludwig J, Olatunbosun D. 1969. Alteration in cobalt absorption in patients with disorders of iron metabolism. Gastroenterology 56(2):241-251.
- Valchev G, Tzvetkova A, Dimitrov L, et al. 1998. Assessment of <sup>60</sup>Co and <sup>54</sup>Mn intakes from whole-body measurements. Radiat Prot Dosim 78(2):151-155.
- \*Valer M, Somogyi Z, Racz I. 1967. Studies concerning the sensitizing effect of cobalt. Dermatologica 134:36-50.
- Van Bastelaere PBM, Callens M, Vangrysperre AE, et al. 1992. Binding characteristics of Mn<sup>2+</sup>, Co<sup>2+</sup> and Mg<sup>2+</sup> ions with several D-xylose isomerases. Biochem J 286:729-735.
- \*Van Bruwaene R, Gerber GB, Kirchmann R, et al. 1984. Metabolism of <sup>51</sup>Cr, <sup>54</sup>Mn, <sup>59</sup>Fe and <sup>60</sup>Co in lactating dairy cows. Health Phys 46(5):1069-1082.
- \*Van Campenhout E. 1955. The cytotoxic effect of cobalt salts on the alpha cells of the Islands of Langerhans. J Exp Zool 124:535-559.
- \*Van Cutsem EJ, Ceuppens JL, Lacquet LM, et al. 1987. Combined asthma and alveolitis induced by cobalt in a diamond polisher. Eur J Respir Dis 70:54-61.
- Van Den Broeke LT, Graslund A, Nilsson JLG, et al. 1998. Free radicals as potential mediators of metal-allergy: Ni<sup>2+</sup>- and Co<sup>2+</sup>-mediated free radical generation. Egypt J Pharm Sci 6:279-286.
- Van Goethem F, Lison D, Kirsch-Volders M. 1997. Comparative evaluation of the in vitro micronucleus test and the alkaline single cell gel electrophoresis assay for the detection of DNA damaging agents: Genotoxic effects of cobalt powder, tungsten carbide and cobalt-tungsten carbide. Mutat Res 392:31-43.
- \*Van Oort RP, Veremy J, Bosch JJT. 1984. Skin response to cobalt 60 irradiation and the consequences for matching the color of facial prostheses. J Prosthet Dent 52:704-710.

# COBALT 370 9. REFERENCES

Van Ostrand G, Cooper RM. 1994. [<sup>14</sup>C]2-deoxyglucose autoradiographic technique provides a metabolic signature of cobalt-induced focal epileptogenesis. Epilepsia 35(5):939-949.

\*Vassilev PP, Venkova K, Pencheva N, et al. 1993. Changes in the contractile responses to carbachol and in the inhibitory effects of verapamil and nitrendipine on isolated smooth muscle preparations from rats subchronically exposed to Co<sup>2+</sup> and Ni<sup>2+</sup>. Arch Toxicol 67:330-337.

Vazquez FG, Aguilera LJ, Sharma VK. 1994. Metals in sediments of San Andres Lagoon, Tamaulipas, Mexico. Bull Environ Contam Toxicol 52:382-387.

Veien NK, Svejgaard E. 1978. Lymphocyte transformation in patients with cobalt dermatitis. Br J Dermatol 99:191-196.

\*Veien NK, Hattel T, Justesen O, et al. 1987. Oral challenge with nickel and cobalt in patients with positive patch tests to nickel and/or cobalt. Acta Derm Venereol (Stockh) 67:321-325.

Veien NK, Hattel T, Laurberg G. 1995. Placebo-controlled oral challenge with cobalt in patients with positive patch tests to cobalt. Contact Dermatitis 33:54-55.

Venkataramani ES, Ahlert RC, Corbo P. 1984. Biological treatment of landfill leachates. CRC Crit Rev Environ Control 14(4):333-376.

Verhamme EN. 1973. Contribution to the evaluation of the toxicity of cobalt. Cobalt 2:29-32.

Verrengia Guerrero NR, Kesten EM. 1994. Levels of heavy metals in waters from the La Plata River, Argentina: An approach to assess bioavailability. Bull Environ Contam Toxicol 52:254-260.

Vertacnik A, Prohic E, Juracic M, et al. 1997. Selected element concentrations in alluvial sediments under garbage disposal site (Zagreb, Croatia). Water Res 31(6):1421-1429.

\*Vieira I, Sonnier M, Cresteil T. 1996. Developmental expression of *CYP2E1* in the human liver: Hypermethylation control of gene expression during the neonatal period. Eur J Biochem 238:476-483.

\*Vienna A, Capucci E, Wolfsperger M, et al. 1995. Heavy metal concentration in hair of students in Rome. Anthropol Anz 53(1):27-32.

\*Vilaplana J, Grimalt F, Romaguera C, et al. 1987. Cobalt content of household cleaning products. Contact Dermatitis 16:139-141.

\*Villanueva S, Botello AV. 1998. Metal pollution in coastal areas of Mexico. Rev Environ Contam Toxicol 157:53-94.

Vitagliano S, Berrino L, Pizzirusso A, et al. 1994. Cobalt blocks L-Glutamate-induced apnea and arterial hypotension in the nucleus tractus solitarii of anaesthetized rats. Neuropharmacology 33(1):145-146.

Volkert WA, Goeckeler WF, Ehrhardt GJ, et al. 1991. Therapeutic radionuclides: Production and decay property considerations. J Nucl Med 32(1):174-185.

Von Gunten HR, Kull TP. 1986. Infiltration of inorganic compounds from the Glatt River, Switzerland, into a groundwater aquifer. Water Air Soil Pollut 29:333-346.

# COBALT 9. REFERENCES 371

Von Zallinger C, Tempel K. 1998. Transplacental transfer of radionuclides. A review. Vet Med (Prague) A45:581-590.

Vos CM, Westera G, Van der Jagt PJ, et al. 1979. The effect of dose loading and of double labeling with <sup>57</sup>Co and <sup>125</sup>I on the tissue distribution in animals. Eur J Nucl Med 4:393-396.

\*Vouk VB. 1986. General chemistry of metals. In: Friberg L, Nordberg GF, Vouk VB, eds. Handbook on the toxicology of metals. 2<sup>nd</sup> ed. New York, NY: Elsevier Science Publishers, 33-34.

\*Voutsinou-Taliadour F, Varnavas SP, Nakopoulou C, et al. 1997. Dissolved trace elements in South Agean seawater. Mar Pollut Bull 34(10):840-843.

WA Dept of Ecology. 2000. Controls for new sources of toxic air pollutants. Washington Department of Ecology. <a href="http://www.wa.gov/ecology/leg/ecywac.html"><u>Http://www.wa.gov/ecology/leg/ecywac.html</u></a>. March 13, 2000.

Walker PR, LeBlanc J, Sikorska M. 1989. Effects of aluminum and other cations on the structure of brain and liver chromatin. Biochem 28:3911-3915.

Wallmann K. 1992. Solubility of cadmium and cobalt in a post-oxic sediment suspension. Hydrobiologia 235/236:611-622.

\*Walter JF. 1980. Cobalt radiation-induced comedones. Arch Dermatol 116:1073-1074.

\*Wang H, Chen D, Gao C, et al. 1993. Effects of low level prenatal <sup>60</sup>Co gamma-irradiation on postnatal growth and behavior in mice. Teratology 48:451-457.

\*Wang JY, Tsukayama DT, Wicklund BH, et al. 1996. Inhibition of T and B cell proliferation by titanium, cobalt, and chromium: Role of IL-2 and IL-6. J Biomed Mater Res 32:655-661.

\*Wang X, Yokoi I, Liu J, et al. 1993. Cobalt(II) and nickel(II) ions as promoters of free radicals in vivo: Detected directly using electron spin resonance spectrometry in circulating blood in rats. Arch Biochem Biophys 306(2):402-406.

\*Watabe T, Uchida S, Kamada H. 1984. Transfer of radionuclides through soil-plant pathway. J Radiat Res 25:274-282.

Watkins S, BAron J, Tephly TR. 1980. Identification of cobalt protoporphyrin IX formation in vivo following cobalt administration to rats. Biochem Pharmacol 29:2319-2323.

\*Weakly JN. 1973. The action of cobalt ions on neuromuscular transmission in the frog. J Physiol 234:597-612.

\*Weast RC. 1985. CRC handbook of chemistry and physics. 66th ed. Boca Raton, Florida: CRC Press.

Webb M. 1962 The biological action of caoble and other metals. III. Chelation of cations by dihydrolipoic acid. Biochim. Biophys. Acta 65:47-65.

\*WEB Research Co. 1999. Mössbauer gamma sources: New lower prices for AEA technology Co57 sources. <a href="http://www.webres.com/gamma\_price.html"><u>Http://www.webres.com/gamma\_price.html.</u></a> April 4, 1999.

\*Wehner AP, Craig DK. 1972. Toxicology of inhaled NiO and CoO in Syrian golden hamsters. Am Ind Hyg Assoc J 33:146-155.

# COBALT 9. REFERENCES

\*Wehner AP, Busch RH, Olson RJ, et al. 1977. Chronic inhalation of cobalt oxide and cigarette smoke by hamsters. Am Ind Hyg Assoc J 38:338-346.

Weinberg SR. 1983. Effects of prenatal irradiation on fetal, neonate, and young adult murine hemopoiesis. Int J Radiat Oncol Biol Phys 9:1825-1831.

\*Wellman PJ, Watkins PA, Nation JR, et al. 1984. Conditioned taste aversion in the adult rat induced by dietary ingestion of cadmium or cobalt. Neurotoxicology 5(2):81-90.

\*West JR, Smith HW, Chasis H. 1948. Glomerular filtration rate, effective renal blood flow, and maximal tubular excretory capacity in infancy. J Pediatr 32:10-18.

Whanger PD, Weswig PH, Schmitz JA, et al. 1976. Effects of selenium, cadmium, mercury, tellurium, arsenic, silver and cobalt on white muscle disease in lambs and effect of dietary forms of arsenic on its accumulation in tissues. Nutr Rep Int 14(1):63-72.

White MA, Dyne D. 1994. Biological monitoring of occupational cobalt exposure in the United Kingdom. Sci Total Environ 150:209-213.

WHO. 2000. Drinking water quality. World Health Organization. Http://www.who.int/. June 5, 2000.

\*Wiberg GS. 1968. The effect of cobalt ions on energy metabolism in the rat. Can J Biochem 46:549-554.

\*Widdowson EM, Dickerson JWT. 1964. Chemical composition of the body. In: Comar CL, Bronner F, eds. Mineral metabolism: An advanced treatise. Volume II: The elements Part A. New York: Academic Press.

\*WI Dept of Natural Resources. 2000. Air pollution control. Wisconsin Department of Natural Resources. Http://www.legis.state.wi.us/rsb/code/nr/nr400.html. March 13, 2000.

Wiegand H, Uhlig S, Gotzsch U, et al. 1990. The action of cobalt, cadmium and thallium on presynaptic currents in mouse motor nerve endings. Neurotoxicol Teratol 12:313-318.

\*Wiersema JM, Wright L, Rogers B, et al. 1984. Human exposure to potentially toxic elements through ambient air in Texas. In: Proceedings of the Air Pollution Control Association 77th Annual Meeting, Austin, TX.

\*Wild P, Perdrix A, Romazini S, et al. 2000. Lung cancer mortality in a site producing hard metals. Occup Environ Med 57:568-573.

\*Wilde M. 1984. Effect of short-term exposure to five industrial metals on the embryonic and fetal development of the mouse. Environ Res 33:47-53.

\*Williams DE, Vlamis J, Pukite AH, et al. 1985. Metal movement in sludge-treated soils after six years of sludge addition: 2. Nickel, cobalt, iron, manganese, chromium, and mercury. Soil Sci 140(2):120-125

Williams LR, Pregenzer JF, Oostveen JA. 1992. Induction of cobalt accumulation by excitatory amino acids within neurons of the hippocampal slice. Brain Res 581:181-189.

# COBALT 9. REFERENCES

Williams SJ, Sabransky M, Menzel DB. 1979. Pulmonary absorption of cobalt salts. Fed Proc 38:394.

Windham ST, Phillips CR. 1973. Radiological survey of New London harbor, Thames River, Conn., and environs. Radiation Data and Reports 14:659-666.

Windholz M. 1983. The Merck index. 10th ed. Rahway, NJ: Merck and Co.

\*Windom HL, Schropp SJ, Calder FD, et al. 1989. Natural trace metal concentrations in estuarine and coastal marine sediments of the southeastern United States. Environ Sci Technol 23(3):314-320.

\*Winger PV, Schultz DP, Johnson WW. 1990. Environmental contamination concentrations in biota from the lower Savannah River, Georgia and South Carolina. Arch Environ Contam Toxicol 19:101-117.

Wojcicki J, Rozewicka L, Kadykow M. 1973. Experimental studies on cobalt cardiopathy. Arch Immunol Ther Exp 21:287-296.

Wolf W. 1993. Radionuclides. In: Elvers B, Hawkins S, Russey W, et al., ed. Ullman's encyclopedia of industrial chemistry. New York, NY: VCH, Vol. A22, 500-543.

\*Wollenberg A, Peter RU, Przybilla B. 1995. Multiple superficial basal cell carcinoma (basalomatosis) following cobalt irradiation. Br J Dermatol 133:644-646.

Wollheim CB, Janjic D. 1984. Cobalt inhibition of insulin release: Evidence for an action not related to Ca<sup>2+</sup> uptake. Am J Physiol 246:C57-C62.

Woltering DM, Larson RJ, Hopping WD, et al. 1987. The environmental fate and effects of detergents. Tens Surfactants Deterg 24(5):286-296.

Woods JS, Carver GT. 1977. Action of cobalt chloride on the biosynthesis, degradation, and utilization of heme in fetal rat liver. Drug Metab Dispos 5(5):487-492.

Yalcintas MG, Jones TD, Meyer HR, et al. 1980. Estimation of dose due to accidental exposure to a <sup>60</sup>Co therapy source. Health Phys 38:187-191.

Yamada H, Koizumi S. 1991. Metallothionein induction in human peripheral blood lymphocytes by heavy metals. Chem Biol Interact 78:347-354.

\*Yamagata N, Murata S, Torii T. 1962. The cobalt content of human body. J Radiat Res 5:4-8.

\*Yamatani K, Saito K, Ikezawa Y, et al. 1998. Relative contribution of Ca<sup>2+</sup>-dependent mechanism in glucagon-induced glucose output from the liver. Arch Biochem Biophys 355(2):175-180.

Yang EYT, Umezawa M, Nahrwold DL. 1991. A relationship between insulin and enterooxyntin. Surg Forum 42:177-179.

\*Yasuda H, Uchida S, Muramatsu Y, et al. 1995. Sorption of manganese, cobalt, zinc, strontium, and cesium onto agricultural soils: Statistical analysis on effects of soil properties. Water Air Soil Pollut 83:85-96.

\*Yasukochi Y, Nakamura M, Minakami S. 1974. Effect of cobalt on the synthesis and degradation of hepatic catalase in vivo. Biochem J 144:455-464.

# COBALT 9. REFERENCES

- \*Ybarra J, Behrooz A, Gabriel A, et al. 1997. Glycemia-lowering effect of cobalt chloride in the diabetic rat: increased GLUT1 mRNA expression. Mol Cell Endocrinol 133:151-160.
- Yifen G, Lianping H, Dechang W. 1992. Effect of <sup>60</sup>Co γ-irradiation n the nonspecific cytotoxicity of alveolar macrophages in vitro. Environ Health Perspect 97:167-170.
- Yoshida T, Numazawa S, Kuroiwa Y. 1986. Induction of hepatic and renal ornithine decarboxylase by cobalt and other metal ions in rats. Biochem J 233:577-581.
- \*Young RS. 1979. Cobalt in biology and biochemistry. London: Academic Press.
- \*Yukawa M, Suzuki-Yasumoto M, Amano K, et al. 1980. Distribution of trace elements in the human body determined by neutron activation analysis. Arch Env Health 35:36-44.
- \*Zanelli R, Barbic F, Migliori M, et al. 1994. Uncommon evolution of fibrosing alveolitis in a hard metal grinder exposed to cobalt dusts. Sci Total Environ 150:225-229.
- \*Zanetti G, Fubini B. 1997. Surface interaction between metallic cobalt and tungsten carbide particles as a primary cause of hard metal lung disease. J Mater Chem 7(8):1647-1654.
- Zenorola P, Bisceglia M, Lomuto M. 1994. Ashy dermatosis associated with cobalt allergy. Contact Dermatitis 31:53-54.
- \*Zhang C, Cai W, Li Y, et al. 1998a. Quantitative analysis of calcitonin gene-related peptide- and neuropeptide Y-immunoreactive nerve fibers in mesenteric blood vessels of rats irradiated with cobalt-60 gamma rays. Radiat Res 149:19-26.
- \*Zhang H, Van Den Berg CMG, Wollast R. 1990. The determination of interactions of cobalt (II) with organic compounds in seawater using cathodic stripping voltammetry. Mar Chem 28:285-300.
- Zhang Q, Kusaka Y, Sato K, et al. 1998b. Differences in the extent of inflammation caused by intratracheal exposure to three ultrafine metals: Role of free radicals. J Toxicol Environ Health, Part A 53:423-438.
- Zhang Q, Kusaka Y, Sato K, et al. 1999. Tumor necrosis factor-alpha release from rat pulmonary leukocytes exposed to ultrafine cobalt: in vivo and in vitro studies. Environ Health Prev Med 4:87-91.
- \*Zhao D, Feng G, Wu X, et al. 1985. Seizures induced by intraventricular microinjection of ionized cobalt in the rat a new experimental model of epilepsy. Brain Res 342:323-329.
- \*Zhong DZ, Pei C, Xiu-Qin L. 1996. Neurobehavioral study of prenatal exposure to hyperthermia combined with irradiation in mice. Neurotoxicol Teratol 18:(6)703-709.
- \*Zhou X-Y, Dong J-C, Geng X-S, et al. 1986. Tritium beta-ray and  $^{60}$ Co gamma-ray caused dominant lethal mutation in mice. Chin Med J 99(5):420-423.
- \*Ziegler EE, Edwards BB, Jensen RL, et al. 1978. Absorption and retention of lead by infants. Pediatr Res 12:29-34.
- \*Zyball A. 1993. Radionuclides. In: Elvers B, Hawkins S, Russey W, et al., ed. Ullman's encyclopedia of industrial chemistry. New York, NY: VCH, Vol. A22, 553-560.

## COBALT 9. REFERENCES

\*Zylicz E, Zablotna R, Geisler J, et al. 1975. Effects of DTPA on the deposition of <sup>65</sup>Zn, <sup>60</sup>Co and <sup>144</sup>Ce in pregnant rat and in fetoplacental unit. Int J Radiat Biol 28(2):125-136.

\*Zylicz E, Zablotna R, Szot Z. 1976. Placental transfer of <sup>60</sup>Co as a function of gestation age. Nukleonika 12:1204-1210.

COBALT 377

#### 10. GLOSSARY

**Absorbed Dose, Chemical**—The amount of a substance that is either absorbed into the body or placed in contact with the skin. For oral or inhalation routes, this is normally the product of the intake quantity and the uptake fraction divided by the body weight and, if appropriate, the time, expressed as mg/kg for a single intake or mg/kg/day for multiple intakes. For dermal exposure, this is the amount of material applied to the skin, and is normally divided by the body mass and expressed as mg/kg.

**Absorbed Dose, Radiation**—The mean energy imparted to the irradiated medium, per unit mass, by ionizing radiation. Units: rad (rad), gray (Gy).

**Absorbed Fraction**—A term used in internal dosimetry. It is that fraction of the photon energy (emitted within a specified volume of material) which is absorbed by the volume. The absorbed fraction depends on the source distribution, the photon energy, and the size, shape and composition of the volume.

**Absorption**—The process by which a chemical penetrates the exchange boundaries of an organism after contact, or the process by which radiation imparts some or all of its energy to any material through which it passes.

**Absorption Coefficient**—Fractional absorption of the energy of an unscattered beam of x- or gamma-radiation per unit thickness (linear absorption coefficient), per unit mass (mass absorption coefficient), or per atom (atomic absorption coefficient) of absorber, due to transfer of energy to the absorber. The total absorption coefficient is the sum of individual energy absorption processes (see Compton Effect, Photoelectric Effect, and Pair Production).

**Absorption Coefficient, Linear**—A factor expressing the fraction of a beam of x- or gamma radiation absorbed in a unit thickness of material. In the expression  $I=I_oe^{-\mu x}$ ,  $I_o$  is the initial intensity, I the intensity of the beam after passage through a thickness of the material x, and  $\mu$  is the linear absorption coefficient.

**Absorption Coefficient, Mass**—The linear absorption coefficient per cm divided by the density of the absorber in grams per cubic centimeter. It is frequently expressed as  $\mu/\rho$ , where  $\mu$  is the linear absorption coefficient and  $\rho$  the absorber density.

**Absorption Ratio, Differential**—Ratio of concentration of a nuclide in a given organ or tissue to the concentration that would be obtained if the same administered quantity of this nuclide were uniformly distributed throughout the body.

**Activation**—The process of making a material radioactive by bombardment with neutrons or protons.

**Activity**—The number of radioactive nuclear transformations occurring in a material per unit time (see Curie, Becquerel). The term for activity per unit mass is specific activity.

Activity Median Aerodynamic Diameter (AMAD)—The diameter of a unit-density sphere with the same terminal settling velocity in air as that of the aerosol particle whose activity is the median for the entire size distribution of the aerosol.

**Acute Exposure, Chemical**—Exposure to a chemical for a duration of 14 days or less, as specified in the Toxicological Profiles.

**Acute Exposure, Radiation**—The absorption of a relatively large amount of radiation (or intake of a radioactive material) over a short period of time.

**Acute Radiation Syndrome**—The symptoms which taken together characterize a person suffering from the effects of intense radiation. The effects occur within hours or days.

Ad libitum—Available in excess and freely accessible.

Adsorption Coefficient  $(K_{oc})$ —The ratio of the amount of a chemical adsorbed per unit surface area or per unit weight of organic carbon of a specific particle size in the soil or sediment to the concentration of the chemical in solution at equilibrium.

Adsorption Ratio  $(K_d)$ —The amount of a chemical adsorbed by a sediment or soil (i.e., the solid phase) divided by the amount of chemical in the solution phase, which is in equilibrium with the solid phase, at a fixed solid/solution ratio. It is generally expressed in micrograms of chemical sorbed per gram of soil or sediment.

**Alpha Particle**—A positively charged particle ejected spontaneously from the nuclei of some radioactive elements. It is identical to a helium nucleus, i.e., 2 neutrons and two protons, with a mass number of 4 and an electrostatic charge of +2.

**Alpha Track**—The track of ionized atoms (pattern of ionization) left in a medium by an alpha particle that has traveled through the medium.

**Annihilation (Positron-Electron)**—An interaction between a positive and a negative electron in which they both disappear; their rest mass, being converted into electromagnetic radiation (called annihilation radiation) with two 0.51 MeV gamma photons emitted at an angle of 180E to each other.

**Atom**—The smallest particle of an element that cannot be divided or broken up by chemical means. It consists of a central core called the *nucleus*, which contains *protons* and *neutrons* and an outer shell of *electrons*.

**Atomic Mass (u)**—The mass of a neutral atom of a nuclide, usually expressed in terms of "atomic mass units." The "atomic mass unit" is one-twelfth the mass of one neutral atom of carbon-12; equivalent to  $1.6604 \times 10^{-24}$  g.

**Atomic Number**—The number of protons in the nucleus of an atom. The "effective atomic number" is calculated from the composition and atomic numbers of a compound or mixture. An element of this atomic number would interact with photons in the same way as the compound or mixture. (Symbol: Z).

Atomic Mass Number—See Mass Number.

**Atomic Weight**—The weighted mean of the masses of the neutral isotopes of an element expressed in atomic mass units.

**Attenuation**—A process by which a beam from a source of radiation is reduced in intensity by absorption and scattering when passing through some material.

**Attenuation Coefficient**—The fractional reduction in the intensity of a beam of radiation as it passes through an absorbing medium. It may be expressed as reduction per unit distance, per unit mass thickness, or per atom, and is called the linear, mass, or atomic attenuation coefficient, respectively.

**Background Radiation**—The amount of radiation to which a member of the general population is exposed from natural sources, such as terrestrial radiation from naturally occurring radionuclides in the soil, cosmic radiation originating from outer space, and naturally occurring radionuclides deposited in the human body.

**Becquerel (Bq)**—International System of Units unit of activity and equals that quantity of radioactive material in which one transformation (disintegration) occurs per second (see Units).

Beta Particle—An electron that is emitted from the nucleus of an atom during one type of radioactive transformation. A beta particle has a mass and charge equal in magnitude to that of the electron. The charge may be either +1 or -1. Beta particles with +1 charges are called positrons (symbolized β<sup>+</sup>), and beta particles with -1 charges are called negatrons (symbolized β-).

**Biological Half-time**—The time required for a biological system, such as that of a human, to eliminate by natural process half of the amount of a substance (such as a chemical substance, either stable or radioactive) that has entered it.

**Bioconcentration Factor (BCF)**—The quotient of the concentration of a chemical in aquatic organisms at a specific time or during a discrete time period of exposure divided by the concentration in the surrounding water at the same time or during the same period.

Biologic Effectiveness of Radiation—See Relative Biological Effectiveness.

**Biomarkers**—Broadly defined as indicators signaling events in biologic systems or samples. They have been classified as markers of exposure, markers of effect, and markers of susceptibility.

Body Burden, Chemical—The total amount of a chemical found in an animal or human body.

Body Burden, Radioactivity—The amount of radioactive material found in an animal or human body.

Bone Seeker—Any compound or ion which migrates in the body and preferentially deposits into bone.

**Buildup Factor**—The ratio of the radiation intensity, including both primary and scattered radiation, to the intensity of the primary (unscattered) radiation.

Cancer Effect Level (CEL)—The lowest dose of chemical or radiation in a study, or group of studies, that produces significant increases in the incidence of cancer (or tumors) between the exposed population and its appropriate control.

Carcinogen—A chemical or radiation that is capable of inducing cancer.

Carcinoma—Malignant neoplasm composed of epithelial cells, regardless of their derivation.

Case-Control Study—A type of epidemiological study which examines the relationship between a particular outcome (disease or condition) and a variety of potential causative agents (such as toxic chemicals). In a case-controlled study, a group of people with a specified and well-defined outcome is identified and compared to a similar group of people without outcome.

**Case Report**—Describes a single individual with a particular disease or exposure. These may suggest some potential topics for scientific research but are not actual research studies.

Cataract—A clouding of the crystalline lens of the eye which obstructs the passage of light.

Ceiling Value—A concentration of a substance that should not be exceeded, even temporarily.

**Charged Particle**—A nuclear particle, atom, or molecule carrying a positive or negative charge.

**Chronic Exposure**—Exposure to a chemical for 365 days or more, as specified in the Toxicological Profiles.

**Cohort Study**—A type of epidemiological study of a specific group or groups of people who have had a common insult (e.g., exposure to an agent suspected of causing disease or a common disease) and are followed forward from exposure to outcome. At least one exposed group is compared to one unexposed group.

**Collective dose**—The sum of the individual doses received in a given period of time by a specified population from exposure to a specified source of radiation. Collective dose is expressed in units such as man-rem and person-sievert.

**Containment**—The confinement of a chemical or radioactive substance in such a way that it is prevented from being dispersed from its container or into the environment, or is released only at a specified rate.

**Contamination**—Deposition of a stable or radioactive substance in any place where it is not desired.

**Cosmic Rays**—High-energy particulate and electromagnetic radiations which originate outside the earth's atmosphere.

**Count (Radiation Measurements)**—The external indication of a radiation-measuring device designed to enumerate ionizing events. It refers to a single detected event. The term "count rate" refers to the total number registered in a given period of time. The term is sometimes erroneously used to designate a disintegration, ionizing event, or voltage pulse.

Counter, Gas-flow Proportional (GPC)— $\beta$ -particles are detected by ionization of the counter gas which results in an electrical impulse at an anode wire.

**Counter, Geiger-Mueller (GM counter)**—Highly sensitive, gas-filled radiation-measuring device to detect (count) individual photons or particulate radiation.

Counter, Scintillation—The combination of phosphor, photomultiplier tube, and associated circuits for counting light emissions produced in the phosphors by ionizing radiation. Employs certain organic compounds (scintillators) which fluoresce when exposed to ionizing radiation. Each fluorescence event is proportional to a radioactive decay event. The frequency of these events is directly proportional to the number of  $\beta$ -emitting atoms present in the sample.

Counting, Cerenkov — Relatively energetic  $\beta$ -particles pass through a transparent medium of high refractive index and a highly-directional, bluish-white light ("Cerenkov" light) is emitted. This light is detected using liquid scintillation counting equipment.

**Cross-sectional Study**—A type of epidemiological study of a group or groups which examines the relationship between exposure and outcome to a chemical or to chemicals at one point in time.

Curie (Ci)—A unit of radioactivity. One curie equals that quantity of radioactive material in which there are  $3.7 \times 10^{10}$  nuclear transformations per second. The activity of 1 gram of radium is approximately 1 Ci.

**Attocurie (aCi)**—One-thousanth of a femtocurie (3.7x10<sup>-8</sup> disintegrations per second).

**Femtocurie** (fCi)—One-billionth of a microcurie (3.7x10<sup>-5</sup> disintegrations per second).

**Megacurie (MCi)**—One million curies (3.7x10<sup>16</sup> disintegrations per sec).

Microcurie ( $\mu$ Ci)—One-millionth of a curie (3.7x10<sup>4</sup> disintegrations per sec).

Millicurie (mCi)—One-thousandth of a curie (3.7x10<sup>7</sup> disintegrations per sec).

Nanocurie (nCi)—One-billionth of a curie (3.7x10<sup>1</sup> disintegrations per sec).

**Picocurie (pCi)**—One-millionth of a microcurie (3.7x10<sup>-2</sup> disintegrations per second.

**Data Needs**—Substance-specific informational needs that if met would reduce the uncertainties of human health assessment.

Daughter Products—See Progeny and Decay Product

**Decay, Radioactive**—Transformation of the nucleus of an unstable nuclide by spontaneous emission of charged particles and/or photons (see Disintegration).

Decay Chain or Decay Series—A sequence of radioactive decays (transformations) beginning with one nucleus. The initial nucleus, the parent, decays into a daughter or progeny nucleus that differs from the first by whatever particles were emitted during the decay. If further decays take place, the subsequent nuclei are also usually called daughters or progeny. Sometimes, to distinguish the sequence, the daughter of the first daughter is called the granddaughter, etc.

**Decay Constant (\lambda)**—The fraction of the number of atoms of a radioactive nuclide which decay in unit time (see Disintegration Constant).

**Decay Product, Daughter Product, Progeny**—A new nuclide formed as a result of radioactive decay. A nuclide resulting from the radioactive transformation of a radionuclide, formed either directly or as the result of successive transformations in a radioactive series. A decay product (daughter product or progeny) may be either radioactive or stable.

**Developmental Toxicity**—The occurrence of adverse effects on the developing organism that may result from exposure to a chemical or radiation prior to conception (either parent), during prenatal development, or postnatally to the time of sexual maturation. Adverse developmental effects may be detected at any point in the life span of the organism.

**Disintegration Constant**—Synonymous with decay constant. The fraction of the number of atoms of a radioactive material that decays per unit time (see Decay Constant.)

**Disintegration, Nuclear**—A spontaneous nuclear transformation (radioactivity) characterized by the emission of energy and/or mass from the nucleus. When large numbers of nuclei are involved, the process is characterized by a definite half-life (see Transformation, Nuclear).

**Distribution coefficient** ( $K_d$ )—Describes the distribution of cobalt between the solid and aqueous phase at thermodynamic equilibrium, is given as follows:

$$K_d = \frac{[Co]_s}{[Co]_{e0}}$$
, Units = (L solution)/(kg solid),

where  $[Co]_s$  is the concentration of cobalt associated with the mineral phase in units of (mg Co)/(kg solid), and  $[Co]_{aq}$  is the concentration of cobalt in the aqueous phase in units of (mg Co)/(L solution). As the magnitude of  $K_ds$  decreases, the potential mobility of cobalt in groundwater systems increases and vice versa .

**Dose**—A general term denoting the quantity of a substance, radiation, or energy absorbed. For special purposes it must be appropriately qualified. If unqualified, it refers to radiation absorbed dose.

**Absorbed Dose**—The energy imparted to matter by ionizing radiation per unit mass of irradiated material at the place of interest. The unit of absorbed dose is the rad. One rad equals 100 ergs per gram. In SI units, the absorbed dose is the gray which is 1 J/kg (see Rad).

**Cumulative Dose (Radiation)**—The total dose resulting from repeated or continuous exposures to radiation.

**Dose Assessment**—An estimate of the radiation dose to an individual or a population group usually by means of predictive modeling techniques, sometimes supplemented by the results of measurement.

**Dose Equivalent (DE)**—A quantity used in radiation safety practice to account for the relative biological effectiveness of the several types of radiation. It expresses all radiations on a common scale for calculating the effective absorbed dose. It is defined as the product of the absorbed dose in rad and certain modifying factors. (The unit of dose equivalent is the rem. In SI units, the dose equivalent is the sievert, which equals 100 rem.)

**Dose, Radiation**—The amount of energy imparted to matter by ionizing radiation per unit mass of the matter, usually expressed as the unit rad, or in SI units, the gray. 100 rad' 1 gray (Gy) (see Absorbed Dose).

Maximum Permissible Dose Equivalent (MPD)—The greatest dose equivalent that a person or specified part thereof shall be allowed to receive in a given period of time.

**Median Lethal Dose (MLD)**—Dose of radiation required to kill, within a specified period (usually 30 days), 50% of the individuals in a large group of animals or organisms. Also called the  $LD_{50}$ , or  $LD_{50/30}$  if for 30 days..

**Threshold Dose**—The minimum absorbed dose that will produce a detectable degree of any given effect.

**Tissue Dose**—Absorbed dose received by tissue in the region of interest, expressed in rad (see Dose, Gray, and Rad).

**Dose, Fractionation**—A method of administering therapeutic radiation in which relatively small doses are given daily or at longer intervals.

**Dose, Protraction**—A method of administering therapeutic radiation by delivering it continuously over a relatively long period at a low dose rate.

**Dose Rate**—Absorbed dose delivered per unit time.

**Dose-Response Relationship**—The quantitative relationship between the amount of exposure to a toxicant and the incidence of the adverse effects.

**Dosimetry**—Quantification of radiation doses to cells, tissues, organs, individuals or populations resulting from specified exposures.

Early Effects (of radiation exposure)—Effects that appear within 60 days of an acute exposure.

**Electron**—A stable elementary particle having an electric charge equal to  $\pm 1.60210 \times 10^{-19}$  C (Coulombs) and a rest mass equal to  $9.1091 \times 10^{-31}$  kg. A positron is a positively charged "electron" (see Positron).

**Electron Volt**—A unit of energy equivalent to the energy gained by an electron in passing through a potential difference of one volt. Larger multiple units of the electron volt are frequently used: keV for thousand or kilo electron volts; MeV for million or mega electron volts (eV).  $1 \text{ eV}=1.6 \text{x} 10^{-12} \text{ erg}$ .

**Embryotoxicity and Fetotoxicity**—Any toxic effect on the conceptus as a result of prenatal exposure to a chemical; the distinguishing feature between the two terms is the stage of development during which the insult occurred. The terms, as used here, include malformations and variations, altered growth, and *in utero* death.

**Energy**—Capacity for doing work. "Potential energy" is the energy inherent in a mass because of its spatial relation to other masses. "Kinetic energy" is the energy possessed by a mass because of its motion (SI unit: joules):

**Binding Energy (Electron)**—The amount of energy that must be expended to remove an electron from an atom.

**Binding Energy (Nuclear)**—The energy represented by the difference in mass between the sum of the component parts and the actual mass of the nucleus. It represents the amount of energy that must be expended to break a nucleus into its component neutrons and protons.

**Excitation Energy**—The energy required to change a system from its ground state to an excited state. Each different excited state has a different excitation energy.

**Ionizing Energy**—The energy required to knock an electron out of an atom. The average energy lost by electrons or beta particles in producing an ion pair in air or in soft tissue is about 34 eV.

**Radiant Energy**—The energy of electromagnetic radiation, such as radio waves, visible light, x and gamma rays.

**EPA Health Advisory**—An estimate of acceptable drinking water levels for a chemical substance based on health effects information. A health advisory is not a legally enforceable federal standard, but serves as technical guidance to assist federal, state, and local officials.

**Epidemiology**—Refers to the investigation of factors that determine the frequency and distribution of disease or other health-related conditions within a defined human population during a specified period.

**Equilibrium, Radioactive**—In a radioactive series, the state which prevails when the ratios between the activities of two or more successive members of the series remains constant.

**Secular Equilibrium**—If a parent element has a very much longer half-life than the daughters (so there is not appreciable change in its amount in the time interval required for later products to attain equilibrium) then, after equilibrium is reached, equal numbers of atoms of all members of the series disintegrate in unit time. This condition is never exactly attained, but is essentially established in such a case as <sup>226</sup>Ra and its transformation series to stable <sup>206</sup>Pb. The half-life of <sup>226</sup>Ra is about 1,600 years; of <sup>222</sup>Rn, approximately 3.82 days, and of each of the subsequent members, a few minutes. After about a month, essentially the equilibrium amount of radon is present; then (and for a long time) all members of the series disintegrate the same number of atoms per unit time. At this time, the activity of the daughter is equal to the activity of the parent.

**Transient Equilibrium**—If the half-life of the parent is short enough so the quantity present decreases appreciably during the period under consideration, but is still longer than that of successive members of the series, a stage of equilibrium will be reached after which all members of the series decrease in activity exponentially with the period of the parent. At this time, the ratio of the parent activity to the daughter activity is constant.

**Equilibrium, Electron**—The condition in a radiation field where the energy of the electrons entering a volume equals the energy of the electrons leaving that volume.

**Equilibrium Fraction (F)**—In radon-radon daughter equilibrium, the parents and daughters have equal radioactivity, that is, as many decay into a specific nuclide as decay out. However, if fresh radon is continually entering a volume of air or if daughters are lost by processes other than radioactive decay, e.g., plate out or migration out of the volume, a disequilibrium develops. The equilibrium fraction is a measure of the degree of equilibrium/disequilibrium. The equilibrium fraction is used to estimate working levels based on measurement of radon only. For radon, 1 working-level concentration is defined at 100 pCi of radon in equilibrium with its 4 successive progeny in 1 liter of air. Thus, 100 pCi/L radon at 50% equilibrium is 0.5 WL.

**Excitation**—The addition of energy to a system, thereby transferring it from its ground state to an excited state. Excitation of a nucleus, an atom, or a molecule can result from absorption of photons or from inelastic collisions with other particles. The excited state of an atom is an unstable or metastable state and will return to ground state by radiation of the excess energy.

**Exposure (Chemical)**—Contact of an organism with a chemical or physical agent. Exposure is quantified as the amount of the agent available at the exchange boundaries of the organism (e.g., skin, lungs, gut) and available for absorption.

**Exposure (Radiation)**—Being exposed to ionizing radiation or to a radioactive material. A measure of the ionization produced in air by x or gamma radiation; the sum of the electric charges on all ions of one sign produced in air when all electrons liberated by photons in a volume of air are completely stopped in air (dQ), divided by the mass of the air in the volume (dm). The unit of exposure in air is the roentgen, or coulomb per kilogram (SI units). One roentgen is equal to  $2.58 \times 10^{-4}$  coulomb per kilogram (C/kg).

**Fission, Nuclear**—A nuclear transformation characterized by the splitting of a nucleus into at least two other nuclei and several neutrons, and is accompanied by the release of a relatively large amount of energy.

Gamma Ray, Penetrating—Short wavelength electromagnetic radiation of nuclear origin.

Genetic Effect of Radiation—Inheritable change, chiefly mutations, produced by the absorption of ionizing radiation by germ cells. Genetic effects have not been observed in any human population exposed at any dose level.

**Genotoxicity**—A specific adverse effect on the genome of living cells that, upon the duplication of affected cells, can be expressed as a mutagenic, clastogenic or carcinogenic event because of specific alteration of the molecular structure of the genome.

**Gray (Gy)**—SI unit of absorbed dose, 1 J/kg. One gray equals 100 rad (see Units).

**Half-life, Radioactive**—Time required for a radioactive substance to lose 50% of its activity by decay. Each radio-nuclide has a unique physical half-life. Known also as physical half-time and symbolized as  $T_r$  or  $T_{rad}$ .

Half-life, Effective—See Half-Time, Effective.

**Half-time, Biological**—Time required for an organ, tissue, or the whole body to eliminate one-half of any absorbed substance by regular processes of elimination. This is the same for both stable and radioactive isotopes of a particular element, and is sometimes referred to as half-time, symbolized as t<sub>biol</sub> or T<sub>b</sub>.

**Half-time, Effective**—Time required for a radioactive element in an organ, tissue, or the whole body to be diminished 50% as a result of the combined action of radioactive decay and biological elimination, symbolized as  $T_e$  or  $T_{eff}$ .

Effective Half&time | Biological half&time x Radioactive half&life | Biological half&time % Radioactive half&life

Immediately Dangerous to Life or Health (IDLH)—The maximum environmental concentration of a contaminant from which one could escape within 30 minutes without any escape-impairing symptoms or irreversible health effects.

**Immunologic Toxicity**—The occurrence of adverse effects on the immune system that may result from exposure to environmental agents such as chemicals.

**Immunological Effects**—Functional changes in the immune response.

*In Vitro*—Isolated from the living organism and artificially maintained, as in a test tube. Literally, "in glass."

In Vivo—Occurring within the living organism. Literally, "in life."

**Intensity**—Amount of energy per unit time passing through a unit area perpendicular to the line of propagation at the point in question.

**Intermediate Exposure**—Exposure to a chemical for a duration of 15–364 days, as specified in the Toxicological Profiles.

Internal Conversion—Process in which a gamma ray knocks an electron out of the same atom from which the gamma ray was emitted. The ratio of the number of internal conversion electrons to the number of gamma quanta emitted in the de-excitation of the nucleus is called the "conversion ratio."

**Ion**—Atomic particle, atom or chemical radical bearing a net electrical charge, either negative or positive.

**Ion Pair**—Two particles of opposite charge, usually referring to the electron and positive atomic or molecular residue resulting after the interaction of ionizing radiation with the orbital electrons of atoms.

**Ionization**—The process by which a neutral atom or molecule acquires a positive or negative charge.

**Primary Ionization**—(1) In collision theory: the ionization produced by the primary particles as contrasted to the "total ionization" which includes the "secondary ionization" produced by delta rays. (2) In counter tubes: the total ionization produced by incident radiation without gas amplification.

**Specific Ionization**—Number of ion pairs per unit length of path of ionizing radiation in a medium; e.g., per centimeter of air or per micrometer of tissue.

**Total Ionization**—The total electric charge of one sign on the ions produced by radiation in the process of losing its kinetic energy. For a given gas, the total ionization is closely proportional to the initial ionization and is nearly independent of the nature of the ionizing radiation. It is frequently used as a measure of absorption of radiation energy.

**Ionization Density**—Number of ion pairs per unit volume.

**Ionization Path (Track)**—The trail of ion pairs produced by an ionizing particle in its passage through matter.

**Ionizing Radiation**—Any radiation capable of knocking electrons out of atoms and producing ions. Examples: alpha, beta, gamma and x rays, and neutrons.

**Isotopes**—Nuclides having the same number of protons in their nuclei, and hence the same atomic number, but differing in the number of neutrons, and therefore in the mass number. Identical chemical properties exist in isotopes of a particular element. The term should not be used as a synonym for nuclide because isotopes refer specifically to different nuclei of the same element.

**Stable Isotope**—A nonradioactive isotope of an element.

**Kerma (k)**—A measure of the kinetic energy transferred from gamma rays or neutrons to a unit mass of absorbing medium in the initial collision between the radiation and the absorber atoms. The SI unit is J/kg. The special name of this unit is the rad (traditional system of units) or Gray (SI).

**Joule**—The S.I. unit for work and energy. It is equal to the work done by raising a mass of one newton through a distance of one meter (J = Nm), which corresponds to about 0.7 ft-pound.

**Labeled Compound**—A compound containing one or more radioactive atoms intentionally added to its structure. By observations of radioactivity or isotopic composition, this compound or its fragments may be followed through physical, chemical, or biological processes.

Late Effects (of radiation exposure)—Effects which appear 60 days or more following an acute exposure.

 $LD_{50/30}$ —The dose of a chemical or radiation expected to cause 50% mortality in those exposed within 30 days. For radiation, this is about 350 rad (3.5 gray) received by humans over a short period of time.

**Lethal Concentration**<sub>(LO)</sub> (LC<sub>LO</sub>)—The lowest concentration of a chemical in air that has been reported to have caused death in humans or animals.

**Lethal Concentration**<sub>(50)</sub> ( $LC_{50}$ )—A calculated concentration of a chemical in air to which exposure for a specific length of time is expected to cause death in 50% of a defined experimental animal population within a specified time, usually 30 days.

**Lethal Dose**<sub>(L0)</sub> ( $LD_{L0}$ )—The lowest dose of a chemical introduced by a route other than inhalation that is expected to have caused death in humans or animals within a specified time, usually 30 days.

**Lethal Dose**<sub>(50)</sub> ( $LD_{50}$ )—The dose of a chemical which has been calculated to cause death in 50% of a defined experimental animal population.

**Lethal Time** $_{(50)}$  (LT $_{50}$ )—A calculated period of time within which a specific concentration of a chemical is expected to cause death in 50% of a defined experimental animal population.

**Linear Energy Transfer (LET)**—A measure of the energy that a charged particle transfers to a material per unit path length.

**Low-LET**—Energy transfer characteristic of light charged particles such as electrons produced by x and gamma rays where the distance between ionizing events is large on the scale of a cellular nucleus.

**High-LET**—Energy transfer characteristic of heavy charged particles such as protons and alpha particles where the distance between ionizing events is small on the scale of a cellular nucleus.

**Average LET**—The energy of a charged particle divided by the length of the path over which it deposits all its energy in a material.

Lowest-Observed-Adverse-Effect Level (LOAEL)—The lowest dose of chemical in a study, or group of studies, that produces statistically or biologically significant increases in frequency or severity of adverse effects between the exposed population and its appropriate control.

Lung Clearance Class (fast, F; medium, M; slow, S)—A classification scheme for inhaled material according to its rate of clearance from the pulmonary region of the lungs to the blood and the gastrointestinal tract.

**Lymphoreticular Effects**—Represent morphological effects involving lymphatic tissues such as the lymph nodes, spleen, and thymus.

**Malformations**—Permanent structural changes that may adversely affect survival, development, or function.

Mass Numbers (A)—The number of nucleons (protons and neutrons) in the nucleus of an atom.

**Minimal Risk Level**—An estimate of daily human exposure to a substance that is likely to be without an appreciable risk of adverse noncancerous effects over a specified duration of exposure.

**Morbidity**—State of being diseased; morbidity rate is the incidence or prevalence of disease in a specific population.

**Mutagen**—A substance that causes changes (mutations) in the genetic material in a cell. Mutations can lead to birth defects, miscarriages, or cancer.

**Necropsy**—The gross examination of the organs and tissues of a dead body to determine the cause of death or pathological conditions.

**Neurotoxicity**—The occurrence of adverse effects on the nervous system following exposure to a substance.

**Neutrino** (v)—A neutral particle of infinitesimally small rest mass emitted during beta plus or beta minus decay. This particle accounts for conservation of energy in beta plus and beta minus decays. It plays no role in damage from radiation.

**No-Observed-Adverse-Effect Level (NOAEL)**—The dose of a substance at which there were no statistically or biologically significant increases in frequency or severity of adverse effects seen between the exposed population and its appropriate control. Effects may be produced at this dose, but they are not considered to be adverse.

**Nuclear reactor**—A power plant that heats water by using nuclear reactions instead of burning coal, oil, or natural gas. All of these sources of energy simply heat water and use the steam which is produced to turn turbines that make electricity or propel a ship.

**Nucleon**—Common name for a constituent particle of the nucleus. Applied to a proton or neutron.

**Nuclide**—A species of atom characterized by the constitution of its nucleus. The nuclear constitution is specified by the number of protons (Z), number of neutrons (N), and energy content; or, alternatively, by the atomic number (Z), mass number A' (N+Z), and atomic mass. To be regarded as a distinct nuclide, the atom must be capable of existing for a measurable time. Thus, nuclear isomers are separate nuclides, whereas promptly decaying excited nuclear states and unstable intermediates in nuclear reactions are not so considered.

Octanol-Water Partition Coefficient (K<sub>ow</sub>)—The equilibrium ratio of the concentrations of a chemical in n-octanol and water, in dilute solution.

**Odds Ratio (OR)**—A means of measuring the association between an exposure (such as toxic substances and a disease or condition) which represents the best estimate of relative risk (risk as a ratio of the incidence among subjects exposed to a particular risk factor divided by the incidence among subjects who were not exposed to the risk factor). An odds ratio of greater than 1 is considered to indicate greater risk of disease in the exposed group compared to the unexposed.

**Pair Production**—An absorption process for x- and gamma radiation in which the incident photon is absorbed in the vicinity of the nucleus of the absorbing atom, with subsequent production of an electron and positron pair (see annihilation). This reaction can only occur for incident photon energies exceeding 1.02 MeV.

**Parent**—A radionuclide which, upon disintegration, yields a new nuclide, either directly or as a later member of a radioactive series.

**Permissible Exposure Limit (PEL)**—A maximum allowable atmospheric level of a substance in workplace air averaged over an 8-hour shift.

**Pharmacokinetics**—The science of quantitatively predicting the fate (disposition) of an exogenous substance in an organism. Utilizing computational techniques, it provides the means of studying the absorption, distribution, metabolism and excretion of chemicals by the body.

**Pharmacokinetic Model**—A set of equations that can be used to describe the time course of a parent chemical or metabolite in an animal system. There are two types of pharmacokinetic models: data-based and physiologically-based. A data-based model divides the animal system into a series of compartments which, in general, do not represent real, identifiable anatomic regions of the body whereas the physiologically-based model compartments represent real anatomic regions of the body.

Physiologically Based Pharmacodynamic (PBPD) Model—A type of physiologically-based dose-response model which quantitatively describes the relationship between target tissue dose and toxic end points. These models advance the importance of physiologically based models in that they clearly describe the biological effect (response) produced by the system following exposure to an exogenous substance

Physiologically Based Pharmacokinetic (PBPK) Model—A model comprising a series of compartments representing organs or tissue groups with realistic weights and blood flows. These models require a variety of physiological information: tissue volumes, blood flow rates to tissues, cardiac output, alveolar ventilation rates and, possibly membrane permeabilities. The models also utilize biochemical information such as air/blood partition coefficients, and metabolic parameters. PBPK models are also called biologically based tissue dosimetry models.

**Photon**—A quantum of electromagnetic energy (E) whose value is the product of its frequency (v) in hertz and Planck's constant (h). The equation is: E = hv.

**Photoelectric Effect**—An attenuation process observed for x and gamma radiation in which an incident photon interacts with a tightly bound inner orbital electron of an atom delivering all of its energy to knock the electron out of the atom. The incident photon disappears in the process.

**Positron**—A positively charged electron.

**Potential, Ionization**—The energy expressed as electron volts (eV) necessary to separate one electron from an atom, resulting in the formation of an ion pair.

**Power, Stopping**—A measure of the ability of a material to absorb energy from an ionizing particle passing through it; the greater the stopping power, the greater the energy absorbing ability (see Linear Energy Transfer).

**Progeny**—The decay product or products resulting after a radioactive decay or a series of radioactive decays. The progeny can also be radioactive, and the chain continues until a stable nuclide is formed.

**Proton**—Elementary nuclear particle with a positive electric charge equal numerically to the charge of the electron and a rest mass of 1.007 mass units.

**Quality**—A term describing the distribution of the energy deposited by a particle along its track; radiations that produce different densities of ionization per unit intensity are said to have different "qualities."

**Quality Factor (Q)**—The linear-energy-transfer-dependent factor by which absorbed doses are multiplied to obtain (for radiation protection purposes) a quantity that expresses - on a common scale for all ionizing radiation - the approximate biological effectiveness of the absorbed dose.

**Rad**—The unit of absorbed dose equal to 100 ergs per gram, or 0.01 joule per kilogram (0.01 Gy) in any medium (see Absorbed Dose).

**Radiation**—The emission and propagation of energy through space or through a material medium in the form of waves (e.g., the emission and propagation of electromagnetic waves, or of sound and elastic waves). The term radiation or radiant energy, when unqualified, usually refers to electromagnetic radiation. Such radiation commonly is classified according to frequency, as microwaves, infrared, visible (light), ultraviolet, and x and gamma rays (see Photon.) and, by extension, corpuscular emission, such as alpha and beta radiation, neutrons, or rays of mixed or unknown type, as cosmic radiation.

**Radiation, Annihilation**—Photons produced when an electron and a positron unite and cease to exist. The annihilation of a positron-electron pair results in the production of two photons, each of 0.51 MeV energy.

Radiation, Background—See Background Radiation.

**Radiation, Characteristic (Discrete)**—Radiation originating from an excited atom after removal of an electron from an atom. The wavelength of the emitted radiation is specific, depending only on the element and particular energy levels involved.

**Radiation**, **External**—Radiation from a source outside the body.

**Radiation, Internal**—Radiation from a source within the body (as a result of deposition of radionuclides in body tissues).

**Radiation, Ionizing**—Any electromagnetic or particulate radiation capable of producing ions, directly or indirectly, in its passage through matter (see Radiation).

**Radiation, Monoenergetic**—Radiation of a given type in which all particles or photons originate with and have the same energy.

**Radiation, Scattered**—Radiation which during its passage through a substance, has been deviated in direction. It may also have been modified by a decrease in energy.

**Radiation, Secondary**—A particle or ray that is produced when the primary radiation interacts with a material, and which has sufficient energy to produce its own ionization, such as bremsstrahlung or electrons knocked from atomic orbitals with enough energy to then produce ionization (see Delta Rays).

Radioactive Material—Material containing radioactive atoms.

**Radioactivity**—Spontaneous nuclear transformations that result in the formation of new elements. These transformations are accomplished by emission of alpha or beta particles from the nucleus or by the capture of an orbital electron. Each of these reactions may or may not be accompanied by a gamma photon.

**Radioactivity, Artificial**—Man-made radioactivity produced by particle bombardment or nuclear fission, as opposed to naturally occurring radioactivity.

**Radioactivity, Induced**—Radioactivity produced in a substance after bombardment with neutrons or other particles. The resulting activity is "natural radioactivity" if formed by nuclear reactions occurring in nature and "artificial radioactivity" if the reactions are caused by man.

**Radioactivity, Natural**—The property of radioactivity exhibited by more than 50 naturally occurring radionuclides.

**Radioisotope**—An unstable or radioactive isotope of an element that decays or disintegrates spontaneously, emitting radiation. Approximately 5,000 natural and artificial radioisotopes have been identified.

Radionuclide—Any radioactive isotope of any element.

**Radiosensitivity**—Relative susceptibility of cells, tissues, organs, organisms, or any living substance to the injurious action of radiation. Radiosensitivity and its antonym, radioresistance, are currently used in a comparative sense, rather than in an absolute one.

**Reference Dose (RfD)**—An estimate (with uncertainty spanning perhaps an order of magnitude) of the daily exposure of the human population to a potential hazard that is likely to be without risk of deleterious effects during a lifetime. The RfD is operationally derived from the NOAEL (from animal and human studies) by a consistent application of uncertainty factors that reflect various types of data used to estimate RfDs and an additional modifying factor, which is based on a professional judgment of the entire database on the chemical. The RfDs are not applicable to non-threshold effects such as cancer.

**Relative Biological Effectiveness (RBE)**—The RBE is a factor used to compare the biological effectiveness of absorbed radiation doses (i.e., rad) due to different types of ionizing radiation. More specifically, it is the experimentally determined ratio of an absorbed dose of a radiation in question to the absorbed dose of a reference radiation (typically <sup>60</sup>Co gamma rays or 200 keV x rays) required to produce an identical biological effect in a particular experimental organism or tissue (see Quality Factor).

**Rem**—A unit of dose equivalent that is used in the regulatory, administrative, and engineering design aspects of radiation safety practice. The dose equivalent in rem is numerically equal to the absorbed dose in rad multiplied by the quality factor (1 rem is equal to 0.01 sievert).

**Reportable Quantity (RQ)**—The quantity of a hazardous substance that is considered reportable under CERCLA. Reportable quantities are (1) 1 pound or greater or (2) for selected substances, an amount established by regulation either under CERCLA or under Sect. 311 of the Clean Water Act. Quantities are measured over a 24-hour period.

**Reproductive Toxicity**—The occurrence of adverse effects on the reproductive system that may result from exposure to a chemical. The toxicity may be directed to the reproductive organs and/or the related endocrine system. The manifestation of such toxicity may be noted as alterations in sexual behavior, fertility, pregnancy outcomes, or modifications in other functions that are dependent on the integrity of this system.

**Roentgen (R)**—A unit of exposure (in air) to ionizing radiation. It is the amount of x or gamma rays required to produce ions carrying 1 electrostatic unit of electrical charge in 1 cubic centimeter of dry air under standard conditions. Named after William Roentgen, a German scientist who discovered x rays in 1895.

**Retrospective Study**—A type of cohort study based on a group of persons known to have been exposed at some time in the past. Data are collected from routinely recorded events, up to the time the study is undertaken. Retrospective studies are limited to causal factors that can be ascertained from existing records and/or examining survivors of the cohort.

**Self-Absorption**—Absorption of radiation (emitted by radioactive atoms) by the material in which the atoms are located; in particular, the absorption of radiation within a sample being assayed.

**Short-Term Exposure Limit (STEL)**—The maximum concentration to which workers can be exposed for up to 15 min continually. No more than four excursions are allowed per day, and there must be at least 60 min between exposure periods. The daily TLV-TWA may not be exceeded.

**SI Units**—The International System of Units as defined by the General Conference of Weights and Measures in 1960. These units are generally based on the meter/kilogram/second units, with special quantities for radiation including the becquerel, gray, and sievert.

Sickness, Acute Radiation (Syndrome)—The complex symptoms and signs characterizing the condition resulting from excessive exposure of the whole body (or large part) to ionizing radiation. The earliest of these symptoms are nausea, fatigue, vomiting, and diarrhea, and may be followed by loss of hair (epilation), hemorrhage, inflammation of the mouth and throat, and general loss of energy. In severe cases, where the radiation dose is relatively high (over several hundred rad or several gray), death may occur within two to four weeks. Those who survive six weeks after exposure of a single high dose of radiation may generally be expected to recover.

**Sievert (Sv)**—The SI unit of any of the quantities expressed as dose equivalent. The dose equivalent in sieverts is equal to the absorbed dose, in gray, multiplied by the quality factor (1 sievert equals 100 rem).

**Specific-activity**—Radioactivity per unit mass of material containing a radionuclide, expressed, for example, as Ci/gram or Bq/gram.

**Specific Energy**—The actual energy per unit mass deposited per unit volume in a small target, such as the cell or cell nucleus, as the result of one or more energy-depositing events. This is a stochastic quantity as opposed to the average value over a large number of instance (i.e., the absorbed dose).

**Standard Mortality Ratio (SMR)**—A ratio of the observed number of deaths and the expected number of deaths in a specific standard population.

**Stopping Power**—The average rate of energy loss of a charged particle per unit thickness of a material or per unit mass of material traversed.

**Surface-seeking Radionuclide**—A bone-seeking internal emitter that is deposited and remains on the bone surface for a long period of time, although it may eventually diffuse into the bone mineral. This contrasts with a volume seeker, which deposits more uniformly throughout the bone volume.

**Target Organ Toxicity**—This term covers a broad range of adverse effects on target organs or physiological systems (e.g., renal, cardiovascular) extending from those arising through a single limited exposure to those assumed over a lifetime of exposure to a chemical.

**Target Theory (Hit Theory)**—A theory explaining some biological effects of radiation on the basis that ionization, occurring in a discrete volume (the target) within the cell, directly causes a lesion which subsequently results in a physiological response to the damage at that location. One, two, or more "hits" (ionizing events within the target) may be necessary to elicit the response.

**Teratogen**—A chemical that causes birth defects.

Threshold Limit Value (TLV)—The maximum concentration of a substance to which most workers can be exposed without adverse effect. TLV is a term used exclusively by the ACGIH. Other terms used to express the same concept are the MAC (Maximum Allowable Concentration) and PEL (Permissible Exposure Limits).

**Time-Weighted Average (TWA)**—An allowable exposure concentration averaged over a normal 8-hour workday or 40-hour workweek.

**Toxic Dose (TD**<sub>50</sub>)—A calculated dose of a chemical, introduced by a route other than inhalation, which is expected to cause a specific toxic effect in 50% of a defined experimental animal population.

**Toxicokinetic**—The study of the absorption, distribution and elimination of toxic compounds in the living organism.

**Toxicosis** —A diseased condition resulting from poisoning.

**Transformation, Nuclear**—The process by which a nuclide is transformed into a different nuclide by absorbing or emitting particulate or electromagnetic radiation.

**Transition, Isomeric**—The process by which a nuclide decays to an isomeric nuclide (i.e., one of the same mass number and atomic number) of lower quantum energy. Isomeric transitions (often abbreviated I.T.) proceed by gamma ray and/or internal conversion electron emission.

**Tritium**—The hydrogen isotope with one proton and two neutrons in the nucleus (Symbol: <sup>3</sup>H). It is radioactive and has a physical half-life of 12.3 years.

**Unattached Fraction**—That fraction of the radon daughters, usually <sup>218</sup>Po and <sup>214</sup>Po, which has not yet attached to a dust particle or to water vapor. As a free atom, it has a high probability of being exhaled and not retained within the lung. It is the attached fraction which is primarily retained.

Uncertainty Factor (UF)—A factor used in operationally deriving the RfD from experimental data. UFs are intended to account for (1) the variation in sensitivity among the members of the human population, (2) the uncertainty in extrapolating animal data to the case of human, (3) the uncertainty in extrapolating from data obtained in a study that is of less than lifetime exposure, and (4) the uncertainty in using LOAEL data rather than NOAEL data. Usually each of these factors is set equal to 10.

#### Units, Radiological—

Units	Equivalents	
Becquerel* (Bq)	1 disintegration per second = 2.7x10 <sup>-11</sup> Ci	
Curie (Ci)	$3.7 \times 10^{10}$ disintegrations per second = $3.7 \times 10^{10}$ Bq	
Gray* (Gy)	1  J/kg = 100  rad	
Rad (rad)	100  erg/g = 0.01  Gy	
Rem (rem)	0.01 sievert	
Sievert* (Sv)	100 rem	

<sup>\*</sup>International Units, designated (SI)

Working Level (WL)—Any combination of short-lived radon daughters in 1 liter of air that will result in the ultimate emission of  $1.3 \times 10^5$  MeV of potential alpha energy.

**Working Level Month (WLM)**—A unit of exposure to radon daughters corresponding to the product of the radon daughter concentration in Working Level (WL) and the exposure time in nominal months (1 nominal month = 170 hours). Inhalation of air with a concentration of 1 WL of radon daughters for 170 working hours results in an exposure of 1 WLM.

**X rays**—Penetrating electromagnetic radiations whose wave lengths are very much shorter than those of visible light. They are usually produced by bombarding a metallic target with fast electrons in a high vacuum. X rays (called characteristic x rays) are also produced when an orbital electron falls from a high energy level to a low energy level.

**Zero-Threshold Linear Hypothesis**—The assumption that a dose-response curve derived from data in the high dose and high dose-rate ranges may be extrapolated through the low dose and low dose range to zero, implying that, theoretically, any amount of radiation will cause some damage.

COBALT A-1

# APPENDIX A ATSDR MINIMAL RISK LEVEL AND WORKSHEETS

The Comprehensive Environmental Response, Compensation, and Liability Act (CERCLA) [42 U.S.C. 9601 et seq.], as amended by the Superfund Amendments and Reauthorization Act (SARA) [Pub. L. 99–499], requires that the Agency for Toxic Substances and Disease Registry (ATSDR) develop jointly with the U.S. Environmental Protection Agency (EPA), in order of priority, a list of hazardous substances most commonly found at facilities on the CERCLA National Priorities List (NPL); prepare toxicological profiles for each substance included on the priority list of hazardous substances; and assure the initiation of a research program to fill identified data needs associated with the substances.

The toxicological profiles include an examination, summary, and interpretation of available toxicological information and epidemiologic evaluations of a hazardous substance. During the development of toxicological profiles, Minimal Risk Levels (MRLs) are derived when reliable and sufficient data exist to identify the target organ(s) of effect or the most sensitive health effect(s) for a specific duration for a given route of exposure. An MRL is an estimate of the daily human exposure to a hazardous substance that is likely to be without appreciable risk of adverse noncancer health effects over a specified duration of exposure. MRLs are based on noncancer health effects only and are not based on a consideration of cancer effects. These substance-specific estimates, which are intended to serve as screening levels, are used by ATSDR health assessors to identify contaminants and potential health effects that may be of concern at hazardous waste sites. It is important to note that MRLs are not intended to define clean-up or action levels.

MRLs are derived for hazardous substances using the no-observed-adverse-effect level/uncertainty factor approach. They are below levels that might cause adverse health effects in the people most sensitive to such chemical-induced effects. MRLs are derived for acute (1–14 days), intermediate (15–364 days), and chronic (365 days and longer) durations and for the oral and inhalation routes of exposure. Currently, MRLs for the dermal route of exposure are not derived because ATSDR has not yet identified a method suitable for this route of exposure. MRLs are generally based on the most sensitive chemical-induced end point considered to be of relevance to humans. Serious health effects (such as irreparable damage to the liver or kidneys, or birth defects) are not used as a basis for establishing MRLs. Exposure to a level above the MRL does not mean that adverse health effects will occur.

MRLs are intended only to serve as a screening tool to help public health professionals decide where to look more closely. They may also be viewed as a mechanism to identify those hazardous waste sites that

are not expected to cause adverse health effects. Most MRLs contain a degree of uncertainty because of the lack of precise toxicological information on the people who might be most sensitive (e.g., infants, elderly, nutritionally or immunologically compromised) to the effects of hazardous substances. ATSDR uses a conservative (i.e., protective) approach to address this uncertainty consistent with the public health principle of prevention. Although human data are preferred, MRLs often must be based on animal studies because relevant human studies are lacking. In the absence of evidence to the contrary, ATSDR assumes that humans are more sensitive to the effects of hazardous substance than animals and that certain persons may be particularly sensitive. Thus, the resulting MRL may be as much as a hundredfold below levels that have been shown to be nontoxic in laboratory animals.

Proposed MRLs undergo a rigorous review process: Health Effects/MRL Workgroup reviews within the Division of Toxicology, expert panel peer reviews, and agencywide MRL Workgroup reviews, with participation from other federal agencies and comments from the public. They are subject to change as new information becomes available concomitant with updating the toxicological profiles. Thus, MRLs in the most recent toxicological profiles supersede previously published levels. For additional information regarding MRLs, please contact the Division of Toxicology, Agency for Toxic Substances and Disease Registry, 1600 Clifton Road, Mailstop E-29, Atlanta, Georgia 30333.

#### MINIMAL RISK LEVEL (MRL) WORKSHEET

Chemical name: Cobalt
CAS number: 10026-24-1
Date: June 2001
Profile status: Draft 3

Route: [x] Inhalation [ ] Oral

Duration: [ ] Acute [ ] Intermediate [x] Chronic

Key to figure: 26 Species: human

 $\widehat{MRL}$ :  $1x10^{-4} [] mg/kg/day [] ppm [x] mg/m<sup>3</sup>$ 

#### Reference:

Nemery B, Casier P, Roosels D, et al. 1992. Survey of cobalt exposure and respiratory health in diamond polishers. Am Rev Respir Dis 145:610-616.

#### Experimental design:

Nemery et al. (1992) conducted a cross-sectional study of cobalt exposure and respiratory effects in diamond polishers. The study group was composed of 194 polishers working in 10 different workshops. In two of these workshops (#1, 2), the workers used cast iron polishing disks almost exclusively, and in the others, they used cobalt-containing disks primarily. The number of subjects from each workshop varied from 6 to 28 and the participation rate varied from 56 to 100%. The low participation in some workshops reflects the fact that only workers who used cobalt disks were initially asked to be in the study, rather than a high refusal rate (only eight refusals were documented). More than a year after the polishing workshops were studied, an additional three workshops with workers engaged in sawing diamonds, cleaving diamonds, or drawing jewelry were studied as an unexposed control group (n=59 workers). Subjects were asked to fill out a questionnaire regarding employment history, working conditions, medical history, respiratory symptoms, and smoking habits, to give a urine sample for cobalt determination, and to undergo a clinical examination and lung function tests. Both area air samples and personal air samples were collected (always on a Thursday). Sampling for area air determinations started 2 hours after work began and continued until 1 hour before the end of the work day. Personal air samples were collected from the breathing zone of a few workers per workshop for four successive 1-hour periods. Air samples were analyzed for cobalt and iron. In addition, personal air samplers were used to sample the air 1 cm above the polishing disks. These samples were analyzed for the entire spectrum of mineral and metallic compounds. Air samples were not obtained at one of the polishing workshops (#4), but this workshop was reported to be almost identical to an adjoining workshop (#3) for which samples were obtained. Urinary cobalt levels were similar between workers in these two workshops, so exposure was considered to be similar as well.

There was a good correlation (R=0.92) between the results of area and personal air sampling, with area air sampling reporting lower concentrations than personal air samples in all workshops except one (#9) (Nemery et al. 1992). In this workshop, personal air samples appeared to be artificially low in comparison to area air samples and urinary cobalt levels of the workers. When this workshop was excluded, there was a good correlation (R=0.85–0.88) between urinary cobalt and cobalt in the air. Based on urinary cobalt levels, the concentration of cobalt expected in personal air samples from workshop #9 was about 45  $\mu$ g/m³ (the mean value actually reported was 6  $\mu$ g/m³). The polishing workshops were divided into two groups: those with low exposure to cobalt (#1–5, n=102) and those with high exposure to cobalt (#6–10, n=91). Mean cobalt exposure concentrations were 0.4, 1.6, and 10.2  $\mu$ g/m³ by area air sampling and 0.4, 5.3, and 15.1  $\mu$ g/m³ by personal air sampling in the control, low-exposure, and high-

A-4

exposure groups, respectively. The inclusion of the apparently biased personal air samples from workshop #9 means that the reported mean cobalt exposure in the high-exposure group obtained by personal air sampling (15.1  $\mu$ g/m³) may be lower than the true value. Air concentrations of iron were highest in the two polishing workshops that used iron disks and the sawing workshop (highest value =  $62 \mu$ g/m³), and were not correlated with cobalt levels. Analysis of samples taken near the disks showed the presence of cobalt, with occasional traces of copper, zinc, titanium, manganese, chromium, silicates, and silicon dioxide. No tungsten was detected. There is a possibility that some workers had previously been exposed to asbestos, since pastes containing asbestos had been used in the past to glue the diamonds onto holders. However, the degree of asbestos exposure had apparently been insufficient to produce functional impairment. The researchers considered cobalt to be the only relevant exposure. Smoking habits were similar in workers from the high-exposure, low-exposure, and control groups. Duration of exposure was not discussed.

#### Effects noted in study and corresponding doses:

Workers in the high-exposure group were more likely than those in the other groups to complain about respiratory symptoms; the prevalences of eye, nose, and throat irritation and cough, and the fraction of these symptoms related to work, were significantly increased in the high-exposure group (Nemery et al. 1992). Workers in the high-exposure group also had significantly reduced lung function compared to controls and low-exposure group workers, as assessed by FVC (forced vital capacity), FEV<sub>1</sub> (forced expiratory volume in 1 second), MMEF (forced expiratory flow between 25 and 75% of the FVC) and mean PEF (peak expiratory flow rate), although the prevalence of abnormal values did not differ significantly between exposure categories. Results in the low-exposure group did not differ from controls. Two-way analysis of variance was used to show that the effect on spirometric parameters in the high exposure group was present in both men and women. Women seemed to be affected more than men, but the interaction between exposure and sex was not significant. Smoking was found to exert a strong effect on lung function, but lung function level remained negatively correlated with exposure to cobalt, independently of smoking.

#### Dose end point used for MRL derivation:

#### [x] NOAEL [ ] LOAEL

Nemery et al. (1992) established a NOAEL of 0.0053 mg cobalt/m<sup>3</sup> for effects on pulmonary function (decreased values upon spirometric examination).

#### <u>Uncertainty factors used in MRL derivation:</u>

[X] I	L	] 3	[ ] 10	(for use of a NOAEL)
[x] 1	[	] 3	[ ] 10	(for extrapolation from animals to humans)
[]1	[	] 3	[x] 10	(for human variability)

The chronic inhalation MRL for cobalt is derived as follows:

```
\begin{aligned} MRL &= NOAEL_{[ADJ]} \div UF \\ MRL &= 0.0013 \text{ mg cobalt/m}^3 \div 10 \\ MRL &= 1x10^{-4} \text{ mg cobalt/m}^3 \end{aligned}
```

Was a conversion factor used from ppm in food or water to a mg/body weight dose? No.

Was a conversion used from intermittent to continuous exposure? If so, explain:

 $0.0053 \text{ mg cobalt/m}^3 * (8 \text{ hours/24 hours}) * (5 \text{ days/7 days}) = 0.0013 \text{ mg cobalt/m}^3 \text{ continuous exposure.}$ 

If an inhalation study in animals, list conversion factors used in determining human equivalent dose: NA

Other additional studies or pertinent information that lend support to this MRL:

Necrosis and inflammation of the respiratory tract epithelium (larynx, trachea, bronchioles, nasal turbinates) were reported in rats exposed to 19 mg cobalt/m³ and mice exposed to 1.9 mg cobalt/m³ (and above) as cobalt sulfate over 16 days (NTP 1991). Exposure of rats and mice to cobalt as cobalt sulfate for 13 weeks resulted in adverse effects on all parts of the respiratory tract, with the larynx being the most sensitive part (NTP 1991). At concentrations of 0.11 mg cobalt/m³ and above, rats and mice had squamous metaplasia of the larynx. Histiocytic infiltrates in the lung were also reported at similar levels in both the rats and mice. In rats, chronic inflammation of the larynx was found at 0.38 mg cobalt/m³ and above, and more severe effects on the larynx, nose, and lung were reported at higher exposures. In mice, acute inflammation of the nose was found at 1.14 mg cobalt/m³ and above, and more severe effects on the larynx, nose, and lung were reported at higher exposures.

Exposure of rats and mice to aerosols of cobalt (as cobalt sulfate) at concentrations from 0.11 to 1.14 mg cobalt/m³ for 2 years resulted in a spectrum of inflammatory, fibrotic, and proliferative lesions in the respiratory tract of male and female rats and mice (NTP 1998). Squamous metaplasia of the larynx occurred in rats and mice at exposure concentrations of 0.11 mg cobalt/m³ and above, with severity of the lesion increasing with increased exposure concentration. Hyperplastic lesions of the nasal epithelium occurred in rats at concentrations of 0.11 mg cobalt/m³ and above, and in mice at concentrations of 0.38 mg cobalt/m³ and above. Both sexes of rats had greatly increased incidences (>90% incidence) of alveolar lesions at all exposure levels, including inflammatory changes, fibrosis, and metaplasia. Similar changes were seen in mice at all exposure levels, though the changes in mice were less severe.

Both studies by NTP (1991, 1998) failed to define a NOAEL, with the lowest concentration examined (0.11 mg/m³) a LOAEL for a variety of respiratory effects. If an MRL were to be calculated based upon these studies, it would be as follows:

Duration adjustment:  $0.11 \text{ mg cobalt/m}^3 * (6 \text{ h/24 h}) * (5 \text{ d/7 d}) = 0.020 \text{ mg cobalt/m}^3 \text{ continuous exposure.}$ 

Calculation of human equivalent concentration:

If fractional depositions in humans and animals are assumed to be equal, then:

 $RDDR = V_E(animal)/S_{ET}(animal) \div V_E(human)/S_{ET}(human) = 0.24 \text{m}^3/\text{day} / 15 \text{cm}^2 \div 20 \text{m}^3/\text{day} / 200 \text{cm}^2 \\ RDDR = 0.16$ 

 $LOAEL_{IHECI} = LOAEL[ADJ] * RDDR$ 

=  $0.020 \text{ mg cobalt/m}^3 * 0.16 = 0.0032 \text{ mg cobalt/m}^3$ 

To the LOAEL<sub>[HEC]</sub>, an uncertainty factor of 300 (10 for use of a LOAEL, 3 for animal to human extrapolation, and 10 for human variability) to derive an MRL of 1x10<sup>-5</sup> mg/m³. This number is an order of magnitude lower than the number derived from the Nemery et al. (1992) data, reflecting the fact that it is derived from animal data, not from a human study, and is based on a LOAEL, not a NOAEL. As the Nemery et al. (1992) data was a well-performed study in humans that defined a NOAEL and LOAEL, it was selected as the basis for derivation of the MRL.

Agency Contact (Chemical Manager): Obaid Faroon D.V.M., Ph.D.

## MINIMAL RISK LEVEL (MRL) WORKSHEET

Chemical name:	Cobalt
CAS number:	10026-24-1
Date:	August 28, 2001
Profile status:	Final Draft Pre-Public
Route:	[ ] Inhalation [x] Oral
Duration:	Acute [x] Intermediate [ ] Chronic
Key to figure:	30
Species:	human
	ng/kg/day [] ppm [] mg/m³
	E. and Fields, J.P. 1958. Experimental production of polycythemia in humans by balt chloride. Proc Soc Exp Biol Med 99:493-495.
chloride, administer received 150 mg col mg/day and later ind punctures of fingert	2: Six apparently normal men, ages 20-47, were administered a daily dose of cobalt ed as a 2% solution diluted in either water or milk, for up to 22 days. Five of the six balt chloride per day for the entire exposure period, while the sixth was started on 120 creased to 150 mg/day. Blood samples were obtained daily from free-flowing ips at least 2 hours after eating, and at least 15 hours after the last dosage of cobalt. I for red blood cell counts, hemoglobin percentage, leukocyte counts, reticulocyte combocyte counts.
polycythemia in all (~16-20% increase a days after cessation though to a lesser ex In five of the six sub	dy and corresponding doses: Exposure to cobalt resulted in the development of six subjects, with increases in red blood cell numbers ranging from 0.5 to 1.19 million above pre-treatment levels). Polycythemic erythrocyte counts returned to normal 9-15 of cobalt administration. Hemoglobin levels were also increased by cobalt treatment, stent than the erythrocyte values, with increases of 6-11% over pretreatment values. Ojects, reticulocyte levels were elevated, reaching at least twice the pre-experiment te and total leukocyte counts did not deviate significantly from pretreatment values.
Dose end point used	I for MRL derivation:
[ ] NOAEL [x] LOA	AEL
erythrocytes in hum	958) identified a LOAEL of 150 mg cobalt chloride per day for increased levels of an volunteers. 150 mg cobalt chloride/day corresponds to ~1 mg Co/kg/day, e body weight of 70 kg.
Uncertainty factors	used in MRL derivation:
[x] 1 [ ] 3 [ ] 10 (	for use of a LOAEL) for extrapolation from animals to humans) for human variability)
MRL = LO $MRL = 1 m$	al MRL for cobalt is derived as follows:  AEL ÷ UF g cobalt/kg-day ÷ 100 0-2 mg cobalt/kg-day

Was a conversion factor used from ppm in food or water to a mg/body weight dose? No.

Was a conversion used from intermittent to continuous exposure? If so, explain: No.

If an inhalation study in animals, list conversion factors used in determining human equivalent dose: Not applicable.

Other additional studies or pertinent information that lend support to this MRL:

Stanley et al. (1947) exposed groups (n=4, 6 for controls) of 6 Sprague-Dawley rats to 0, 0.62, 2.5, or 10 mg cobalt/kg/day (0, 2.5, 10, or 40 mg/kg-day of CoCl<sub>2</sub>@H<sub>2</sub>O) in gelatin capsules for 8 weeks. Blood counts and hemoglobin levels were examined at the beginning of the experiment and at 2-week intervals. Rats exposed to 0.62 mg cobalt/kg-day showed no change in erythrocyte number. At 2.5 mg cobalt/kg-day, a progressive increase in erythrocyte number was seen, increasing up to a maximum of 17% above pretreatment values on week 6. At the highest exposure level, a progressive increase in erythrocyte numbers was seen, reaching 29% above pretreatment values at 8 weeks of exposure. Statistical analyses of the group means were not provided, and the study provided only mean values of the measurements, precluding statistical analysis. However, if a 10% change is assumed to be an effect level, exposure to 2.5 mg cobalt/kg-day was the LOAEL for this study, with a NOAEL of 0.62 mg cobalt/kg-day.

Whether or not polycythemia, a condition wherein an excess of erythrocytes is produced, constitutes an adverse effect is open to interpretation. At the levels seen in the available studies, and in particular in the Davis and Fields (1958) study, the subjects would be expected to be asymptomatic. At higher erythrocyte levels, the condition may progress to a diagnosis of polycythemia vera, which could result in increases in clot formation and blockage of small blood vessels, with a possible risk of stroke. It is also conceivable that a prolonged excess in erythrocyte production, such as is seen in polycythemia vera, may result in an eventual failure of the bone marrow and a resulting anemia; this "spent phase" occurs in approximately 30% of patients with polycythemia vera. While the increases seen in the available human and animal data do not yet approach those levels, they were considered to be precursor events as a health-protective assumption. Therefore, polycythemia was utilized as a critical endpoint for MRL derivation.

Agency Contact (Chemical Manager): Obaid Faroon D.V.M., Ph.D.

#### MINIMAL RISK LEVEL (MRL) WORKSHEET

Chemical name: Radioactive Cobalt

CAS number: Multiple Date: June 2001

Profile status: Draft 3

Route: [ ] Inhalation [ ] Oral [x] External
Duration: [x] Acute [ ] Intermediate [ ] Chronic

Species: Human

MRL:  $4 \lceil \frac{\text{mg/kg/day}}{\text{mg}} \lceil \frac{\text{mg/m}^3}{\text{mg}} \lceil \frac{\text{mSv}}{\text{mg}} \rceil \rceil$ 

#### References:

Schull WJ, Otake M, and Yoshimaru H. 1988. Effect on intelligence test score of prenatal exposure to ionizing radiation in Hiroshima and Nagasaki: A comparison of the T65DR and DS86 dosimetry systems.

Burt C. 1966. The genetic determination of differences in intelligence: A study of monozygotic twins reared together and apart. Brit J Psychol 57(1 & 2): 137-153.

#### Experimental design:

Schull et al. (1988) study: Schull et al. (1988) evaluated the quantitative effect of exposure to ionizing radiation on the developing fetal and embryonic human brain. The end point measured was changes in intelligence test scores. The effects on individuals exposed *in utero* to the atomic bombing of Hiroshima and Nagasaki were based on the original PE86 samples (n=1,759; data on available intelligence testing) and a clinical sample (n=1,598). The original PE86 sample included virtually all prenatally exposed individuals who received tissue-absorbed doses of 0.50 Gy or more. There were many more individuals in the dose range 0–0.49 Gy in the PE86 sample than in the clinical sample. The clinical sample does not include children prenatally exposed at distances between 2,000 and 2,999 m in Hiroshima and Nagasaki. Children exposed at greater distances or not present in the city were selected as controls. In 1955–1956, Tanaka-B (emphasis on word-sense, arithmetic abilities, and the like, which were associated with the more subtle processing of visual clues than their simple recognition and depended more on connectedness) and the Koga (emphasis on perception of spatial relationships) intelligence tests were conducted in Nagasaki and the Koga test in Hiroshima.

Burt (1966) study: This study determined differences in intelligence in monozygotic twins reared together (n=95) and apart (n=53). All tests conducted in school consisted of (1) a group test of intelligence containing both non-verbal and verbal items, (2) an individual test (the London Revision of the Terman-Binet Scale) used primarily for standardization and for doubtful cases, and (3) a set of performance tests, based on the Pitner-Paterson tests and standardization. The methods and standard remained much the same throughout the study. Some of the reasons for separation of the twins were given as follows: death of the mother (n=9), unable to bring them up properly, mother's poor health (n=12), unmarried (n=6), and economic difficulties. The children were brought up by parents or foster parents (occupation ranged from unskilled to professional). IQ scores in the study group ranged from 66 to 137. The standard deviation of the group of separated monozygotic twins was reported at 15.3 as compared to 15.0 of ordinary siblings. Twins brought up in different environments were compared with those brought up in similar circumstances.

#### Effects noted in study and corresponding doses:

Schull et al. (1986) study: No evidence of radiation-related effect on intelligence was observed among individuals exposed within 0–7 weeks after fertilization or in the 26th or subsequent weeks. The highest risk of radiation damage to the embryonic and fetal brain occurs 8–15-weeks after fertilization under both dosimetric systems. The regression of intelligence score on estimated DS86 uterine absorbed dose is linear with dose, and the diminution in intelligence score is 21–29 points per Gy for the 8–15-week group and 10–26 points per Gy for the 16–25-week group. The results for 8–15 weeks applies regardless whether or not the mentally retarded individuals were included. The cumulative distribution of test scores suggested a progressive shift downwards in individual scores with increasing exposure. The mean IQ scores decrease significantly and systematically with uterine or fetal tissue dose within the 8–15- and 16–25-week groups.

In summary, analysis of intelligence test scores at 10–11 years of age of individuals exposed prenatally showed that:

- There is no evidence of a radiation-related effect on intelligence scores among those individuals exposed within 0–7 weeks of fertilization or in the 26<sup>th</sup> week of gestation and beyond;
- The cumulative distribution of test scores suggests a progressive shift downwards in intelligence scores with increasing exposure to ionizing radiation (dose-response relationship).
- The most sensitive group was the 8–15 weeks exposure group. The regression in intelligence scores was found to be linear, with 1 Gy dose resulting in a 21–29 point decline in intelligence scores.
- There was no indication of groups of individuals with differing sensitivities to radiation.

**Burt (1966) study:** The average intelligence of the twins measured on a conventional IQ scale (SD=15) was 97.8 for the separated monozygotes, 98.1 for monozygotes brought up together, 99.3 for the dizygotes as compared with 100.2 for the siblings, and 100.0 for the population as a whole. The difference of 0.3 IQ point between the separated and unseparated identical twins is considered a NOAEL for this study.

Dose endpoint used for MRL derivation:

[x] NOAEL [ ] LOAEL	0.3 IQ point reduction in twins, between those raised together and those raised apart.
Uncertainty factors (UF)	used in MRL derivation:

[x] 1 [ ] 3	[] 10 (for use of a NOAEL)
[x] 1 [ ] 3	[] 10 (for extrapolation from animals to humans)
[ ] 1 [x] 3	[ ] 10 (for human variability/sensitive population

Was a conversion factor used from ppm in food or water to a mg/body weight dose? If so, explain: No.

If an inhalation study in animals, list conversion factors used in determining human equivalent dose: NA

Was a conversion used from intermittent to continuous exposure? No.

#### Other additional studies or pertinent information that lend support to this MRL:

Husen (1959) reported a study involving 269 pairs of Swedish monozygotic (identical) twins where the intrapair IQ difference was 4 IQ points for a combination of twins raised together and apart. This is somewhat lower than the value of 7 IQ points for identical twins raised apart, and just larger than the range of IQ scores for Washington, DC children repetitively tested (Jacobi and Glauberman 1995).

Supporting evidence for the acute MRL is provided by Jacobi and Glauberman (1995). Children in the 1<sup>st</sup>, 3<sup>rd</sup>, and 5<sup>th</sup> grades born in Washington, DC were tested, and average IQ levels of 94.2, 97.6, and 94.6, respectively, were reported. The range of 3.4 IQ points is considered to be a LOAEL for this study, which, if used for MRL derivation, would yield an MRL of 0.004 Sv (3.4 IQ points x 1 Sv/25 IQ points ÷ 30 [10 for use of a LOAEL and 3 for a sensitive population]).

Additional supporting evidence for the acute MRL is provided by Berger et al. 1997, in a case study of accidental radiation injury to the hand. A Mexican engineer suffered an accidental injury to the hand while repairing an x-ray spectrometer. The day after the accident, his symptoms included a tingling sensation and itching in the index and middle fingers. On days 4 and 7, a "pinching" sensation, swelling, and slight erythema were observed. By day 7, the tip of his index fingers was erythematous and a large blister developed with swelling on other fingers. On day 10, examination by a physician showed that the lesions had worsened and the fingers and palms were discolored. On day 10, he was admitted to the hospital where hyperbaric oxygen therapy was administered without success. One month after the accident, the patient entered the hospital again with pain, discoloration, and desquamation of his hand. Clinical examination showed decreased circulation in the entire hand, most notably in the index and middle finger. Total white blood count decreased to  $3,000/\mu L$  (normal range  $4,300-10,800/\mu L$ ). Cytogenic studies of peripheral blood lymphocytes revealed four dicentrics, two rings, and eight chromosomal fragments in the 300 metaphases studied. The estimated whole body dose was reported to be 0.382 Gy (38.2 rad). This dose is a potential LOAEL for acute ionizing radiation and would yield an MRL of 0.004 Sy (0.38 Sy  $\div 100$  [10 for use of LOAEL and 10 for sensitive human population]).

The NRC set a radiation exposure limit of 0.5 rem (50 mSv) for pregnant working women over the full gestational period (NRC 1991). For the critical gestational period of 8–15 weeks ATSDR believes that the conservative acute MRL of 4 mSv is consistent with the NRC limit and could be applied to either acute (0–14-day) or intermediate (15–365-day) exposure periods.

#### Calculations

**Given**: 0.3 IQ point is a NOAEL. A 1 Sv dose results in a 25 IQ point reduction (range=21–29 points; mean=25) and provides a conversion factor from IQ prediction to radiation dose. Assume that the radiation dose and the subsequent reduction in IQ is a linear relationship.

 $MRL = NOAEL \times CF \div UF$ 

 $MRL = 0.3 \times 1/25 \div 3$ 

MRL = 0.004 Sv = 4 mSv (400 mrem)

Agency Contact (Chemical Manager): Obaid Faroon D.V.M., Ph.D.

## MINIMAL RISK LEVEL (MRL) WORKSHEET

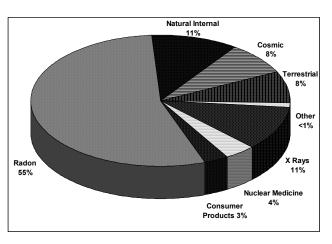
Chemical name: Radio	pactive Cobalt
CAS number:	Multiple
Date:	June 2001
Profile status:	Draft 3
Route:	[] Inhalation [] Oral [x] External
Duration:	[] Acute [] Intermediate [x] Chronic
Species:	Human
MRL:	1 [] mg/kg/day [] ppm [] mg/m <sup>3</sup> [x] mSv/yr (100 mrem/yr)
	1990. Health effects of exposure to low levels of ionizing radiation. Committee on s of Ionizing Radiations, National Research Council. National Academy Press.
Experimental design:	NA
to base a chronic-dura However, two sources have not been reporte estimates of backgrou implicated in produci average annual effect equivalent of 3.6 mSv naturally occurring ra- consumer products.	and corresponding doses: No individual studies were identified that could be used ation external exposure MRL that did not result in a cancer-producing end point. It is of information were identified that did provide doses of ionizing radiation that and to be associated with detrimental effects (NOAELs). These sources provide and levels of primarily natural sources of ionizing radiation that have not been an granderous or non-cancerous toxicological endpoints. BEIR V states that the live dose to the U.S. population is 3.6 mSv/year. A total annual effective dose of (360 mrem)/year to members of the U.S. population is obtained mainly by adiation from external sources, medical uses of radiation, and radiation from The largest contribution (82%) is from natural sources, two-thirds of which is from adon and its decay products. Specific sources of this radiation are demonstrated in
	6 mSv per year has not been associated with adverse health effects or increases in type of cancers in humans or other animals.
Dose endpoint used for	or MRL derivation:
3.6 mSv/year	
[x] NOAEL [ ] LOAF	EL 3.6 mSv/year
Uncertainty factors (U	UF) used in MRL derivation:
[x] 1 [ ] 3 [ ] 10 (for [x] 1 [ ] 3 [ ] 10 (for [x] 1 [ ] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 [ x] 3 [ ] 10 (for [x] 1 [ x] 3 ( x	r extrapolation from animals to humans)

Was a conversion factor used from ppm in food or water to a mg/body weight dose? If so, explain: No.

#### APPENDIX A

Table A-1. Average Annual Effective Dose Equivalent of Ionizing Radiation to a Member of the U.S. Population<sup>a</sup>

	Effective Dose Equivalent	
Source	mSv	Percent of Total Dose
Natural		
Radon <sup>b</sup>	2.0	55
Cosmic	0.27	8.0
Terrestrial	0.28	8.0
Internal	0.39	11
Total Natural	3.0	82
Artificial		
Medical	_	
X-ray	0.39	11
Nuclear	0.14	4.0
Consumer Products	0.10	3.0
Other		
Occupational	< 0.01	< 0.3
Nuclear Fuel Cycle	< 0.01	< 0.03
Fallout	< 0.01	< 0.03
Miscellaneous <sup>c</sup>	< 0.01	< 0.03
Total Artificial	0.63	18
Total Natural and Artificial	3.6	100



If an inhalation study in animals, list conversion factors used in determining human equivalent dose: NA

Was a conversion used from intermittent to continuous exposure? No.

 $<sup>^{\</sup>text{a}}\text{adapted}$  from BEIR V, Table 1-3 , page 18.

<sup>&</sup>lt;sup>b</sup>Dose equivalent to bronci from radon daughter products

<sup>&</sup>lt;sup>c</sup>DOE facilities, smelter, transportation, etc.

#### Other additional studies or pertinent information that lend support to this MRL:

ICRP has developed recommended dose limits for occupational and public exposure to ionizing radiation sources and the US has adopted these values (10 CFR 20). The ICRP recommends limiting public exposure to 1 mSv/year (100 mrem/year), but does note that values at high altitudes above sea level and in some geological areas can sometimes be twice that value (\$2 mSv). In Annex C of ICRP 60, the commission provides projections of risk from the high dose, high dose rate Japanese atomic bomb survivor data and a relatively low dose, low dose rate Swedish population (considering a dose and dose rate effectiveness factor of 2) that predict a potentially small increase in age-specific human mortality rate with increasing radiation dose from 1 mSv to 5 mSv. This increase may not be able to be detected in a typical population. ICRP states that the value of 1 mSv/year was chosen over the 5 mSv value because 5 mSv/year (500 mrem/year) could result in a small increase in age specific mortality rate, and 1 mSv/year (100 mrem/year) is typical of the annual effective dose from background, less radon (ICRP 1991). The 1 mSv estimate may underestimate the annual exposure to external sources of ionizing radiation to the U.S. population, as it does not include radiation from radon. Conversely, the 5 mSv estimate may be high, in that this larger dose could potentially result in a small mortality increase. The most useful estimate appears to be the BEIR V estimate of 3.6 mSv as a nationwide average that accounts for an annual exposure to radon, is specific to the U.S. population, has not been associated with increases mortality, and it falls short of the 5 mSv value potentially associated with small increases in human mortality.

#### Calculations

$$\begin{split} MRL &= NOAEL_{(ADJ)} \div UF \\ MRL &= 3.6 \text{ mSv/year} \div 3 \\ MRL &= 1.20 \text{ mSv/year} \\ MRL &= 1.0 \text{ mSv/year} = 100 \text{ mrem/yr above background} \end{split}$$

Agency Contact (Chemical Manager): Obaid Faroon D.V.M., Ph.D.

COBALT B-1

# APPENDIX B USER'S GUIDE

#### Chapter 1

#### Public Health Statement

This chapter of the profile is a health effects summary written in non-technical language. Its intended audience is the general public especially people living in the vicinity of a hazardous waste site or chemical release. If the Public Health Statement were removed from the rest of the document, it would still communicate to the lay public essential information about the chemical.

The major headings in the Public Health Statement are useful to find specific topics of concern. The topics are written in a question and answer format. The answer to each question includes a sentence that will direct the reader to chapters in the profile that will provide more information on the given topic.

#### Chapter 2

#### Relevance to Public Health

This chapter provides an overview of the nature, manufacture, uses, general population exposures, and health effects of the substance under review. This overview is followed by a discussion of the most critical health effects. The discussion of health effects is based on evaluations of existing toxicologic, epidemiologic, and toxicokinetic information. This summary is designed to present interpretive, weight-of-evidence discussions for human health end points by addressing the following questions.

- 1. What effects are known to occur in humans?
- 2. What effects observed in animals are likely to be of concern to humans?
- 3. What exposure conditions are likely to be of concern to humans, especially around hazardous waste sites?

The chapter covers end points in the same order they appear within the Discussion of Health Effects by Route of Exposure section, by route (inhalation, oral, dermal) and within route by effect. Human data are presented first, then animal data. Both are organized by duration (acute, intermediate, chronic). *In vitro* data and data from parenteral routes (intramuscular, intravenous, subcutaneous, etc.) are also considered in this chapter. If data are located in the scientific literature, a table of genotoxicity information is included.

The carcinogenic potential of the profiled substance is qualitatively evaluated, when appropriate, using existing toxicokinetic, genotoxic, and carcinogenic data. ATSDR does not currently assess cancer potency or perform cancer risk assessments. Minimal risk levels (MRLs) for noncancer end points (if derived) and the end points from which they were derived are indicated and discussed.

Limitations to existing scientific literature that prevent a satisfactory evaluation of the relevance to public health are identified in the Chapter 3 Data Needs section.

#### Interpretation of Minimal Risk Levels

Where sufficient toxicologic information is available, we have derived minimal risk levels (MRLs) for inhalation and oral routes of entry at each duration of exposure (acute, intermediate, and chronic). These MRLs are not meant to support regulatory action; but to acquaint health professionals with exposure levels at which adverse health effects are not expected to occur in humans. They should help physicians and public health officials determine the safety of a community living near a chemical emission, given the concentration of a contaminant in air or the estimated daily dose in water. MRLs are based largely on toxicological studies in animals and on reports of human occupational exposure.

MRL users should be familiar with the toxicologic information on which the number is based. Chapter 2, "Relevance to Public Health," contains basic information known about the substance. Other sections such as Chapter 3 Section 3.9, "Interactions with Other Substances," and Section 3.10, "Populations that are Unusually Susceptible" provide important supplemental information.

MRL users should also understand the MRL derivation methodology. MRLs are derived using a modified version of the risk assessment methodology the Environmental Protection Agency (EPA) provides (Barnes and Dourson 1988) to determine reference doses for lifetime exposure (RfDs).

To derive an MRL, ATSDR generally selects the most sensitive end point which, in its best judgement, represents the most sensitive human health effect for a given exposure route and duration. ATSDR cannot make this judgement or derive an MRL unless information (quantitative or qualitative) is available for all potential systemic, neurological, and developmental effects. If this information and reliable quantitative data on the chosen end point are available, ATSDR derives an MRL using the most sensitive species (when information from multiple species is available) with the highest NOAEL that does not exceed any adverse effect levels. When a NOAEL is not available, a lowest-observed-adverse-effect level (LOAEL) can be used to derive an MRL, and an uncertainty factor (UF) of 10 must be employed. Additional uncertainty factors of 10 must be used both for human variability to protect sensitive subpopulations (people who are most susceptible to the health effects caused by the substance) and for interspecies variability (extrapolation from animals to humans). In deriving an MRL, these individual uncertainty factors are multiplied together. The product is then divided into the inhalation concentration or oral dosage selected from the study. Uncertainty factors used in developing a substance-specific MRL are provided in the footnotes of the LSE Tables.

## Chapter 3

#### **Health Effects**

#### Tables and Figures for Levels of Significant Exposure (LSE)

Tables (3-1, 3-2, and 3-3) and figures (3-1 and 3-2) are used to summarize health effects and illustrate graphically levels of exposure associated with those effects. These levels cover health effects observed at increasing dose concentrations and durations, differences in response by species, minimal risk levels (MRLs) to humans for noncancer end points, and EPA's estimated range associated with an upper-bound individual lifetime cancer risk of 1 in 10,000 to 1 in 10,000,000. Use the LSE tables and figures for a quick review of the health effects and to locate data for a specific exposure scenario. The LSE tables and figures should always be used in conjunction with the text. All entries in these tables and figures represent studies that provide reliable, quantitative estimates of No-Observed-Adverse-Effect Levels (NOAELs), Lowest-Observed-Adverse-Effect Levels (LOAELs), or Cancer Effect Levels (CELs).

The legends presented below demonstrate the application of these tables and figures. Representative examples of LSE Table 3-1 and Figure 3-1 are shown. The numbers in the left column of the legends correspond to the numbers in the example table and figure.

#### LEGEND

#### See LSE Table 3-1

- (1) Route of Exposure One of the first considerations when reviewing the toxicity of a substance using these tables and figures should be the relevant and appropriate route of exposure. When sufficient data exists, three LSE tables and two LSE figures are presented in the document. The three LSE tables present data on the three principal routes of exposure, i.e., inhalation, oral, and dermal (LSE Table 3-1, 3-2, and 3-3, respectively). LSE figures are limited to the inhalation (LSE Figure 3-1) and oral (LSE Figure 3-2) routes. Not all substances will have data on each route of exposure and will not therefore have all five of the tables and figures. In the case of radionuclide profiles, there are separate LSE tables, by route of exposure, for toxicity of stable isotopes and radioactive isotopes; for radioactive isotopes, there may be an LSE table for a fourth route, external exposure.
- (2) Exposure Period Three exposure periods acute (less than 15 days), intermediate (15–364 days), and chronic (365 days or more) are presented within each relevant route of exposure. In this example, an inhalation study of intermediate exposure duration is reported. For quick reference to health effects occurring from a known length of exposure, locate the applicable exposure period within the LSE table and figure.
- (3) <u>Health Effect</u> The major categories of health effects included in LSE tables and figures are death, systemic, immunological, neurological, developmental, reproductive, and cancer. NOAELs and LOAELs can be reported in the tables and figures for all effects but cancer. Systemic effects are further defined in the "System" column of the LSE table (see key number 18).
- (4) <u>Key to Figure</u> Each key number in the LSE table links study information to one or more data points using the same key number in the corresponding LSE figure. In this example, the study represented by key number 18 has been used to derive a NOAEL and a Less Serious LOAEL (also see the 2 "18r" data points in Figure 3-1).
- (5) Species The test species, whether animal or human, are identified in this column. Chapter 2, "Relevance to Public Health," covers the relevance of animal data to human toxicity and Section 3.4, "Toxicokinetics," contains any available information on comparative toxicokinetics. Although NOAELs and LOAELs are species specific, the levels are extrapolated to equivalent human doses to derive an MRL.
- (6) Exposure Frequency/Duration The duration of the study and the weekly and daily exposure regimen are provided in this column. This permits comparison of NOAELs and LOAELs from different studies. In this case (key number 18), rats were exposed to 1,1,2,2-tetrachloroethane via inhalation for 6 hours per day, 5 days per week, for 3 weeks. For a more complete review of the dosing regimen refer to the appropriate sections of the text or the original reference paper, i.e., Nitschke et al. 1981.
- (7) <u>System</u> This column further defines the systemic effects. These systems include: respiratory, cardiovascular, gastrointestinal, hematological, musculoskeletal, hepatic, renal, and dermal/ocular. "Other" refers to any systemic effect (e.g., a decrease in body weight) not covered in these systems. In the example of key number 18, 1 systemic effect (respiratory) was investigated.

- (8) <u>NOAEL</u> A No-Observed-Adverse-Effect Level (NOAEL) is the highest exposure level at which no harmful effects were seen in the organ system studied. Key number 18 reports a NOAEL of 3 ppm for the respiratory system which was used to derive an intermediate exposure, inhalation MRL of 0.005 ppm (see footnote "b").
- (9) <u>LOAEL</u> A Lowest-Observed-Adverse-Effect Level (LOAEL) is the lowest dose used in the study that caused a harmful health effect. LOAELs have been classified into "Less Serious" and "Serious" effects. These distinctions help readers identify the levels of exposure at which adverse health effects first appear and the gradation of effects with increasing dose. A brief description of the specific end point used to quantify the adverse effect accompanies the LOAEL. The respiratory effect reported in key number 18 (hyperplasia) is a Less serious LOAEL of 10 ppm. MRLs are not derived from Serious LOAELs.
- (10) <u>Reference</u> The complete reference citation is given in Chapter 9 of the profile.
- (11) <u>CEL</u> A Cancer Effect Level (CEL) is the lowest exposure level associated with the onset of carcinogenesis in experimental or epidemiologic studies. CELs are always considered serious effects. The LSE tables and figures do not contain NOAELs for cancer, but the text may report doses not causing measurable cancer increases.
- (12) <u>Footnotes</u> Explanations of abbreviations or reference notes for data in the LSE tables are found in the footnotes. Footnote "b" indicates the NOAEL of 3 ppm in key number 18 was used to derive an MRL of 0.005 ppm.

#### LEGEND

#### See Figure 3-1

LSE figures graphically illustrate the data presented in the corresponding LSE tables. Figures help the reader quickly compare health effects according to exposure concentrations for particular exposure periods.

- (13) Exposure Period The same exposure periods appear as in the LSE table. In this example, health effects observed within the intermediate and chronic exposure periods are illustrated.
- (14) <u>Health Effect</u> These are the categories of health effects for which reliable quantitative data exists. The same health effects appear in the LSE table.
- (15) <u>Levels of Exposure</u> concentrations or doses for each health effect in the LSE tables are graphically displayed in the LSE figures. Exposure concentration or dose is measured on the log scale "y" axis. Inhalation exposure is reported in mg/m³ or ppm and oral exposure is reported in mg/kg/day.
- (16) <u>NOAEL</u> In this example, 18r NOAEL is the critical end point for which an intermediate inhalation exposure MRL is based. As you can see from the LSE figure key, the open-circle symbol indicates to a NOAEL for the test species-rat. The key number 18 corresponds to the entry in the LSE table. The dashed descending arrow indicates the extrapolation from the exposure level of 3 ppm (see entry 18 in the Table) to the MRL of 0.005 ppm (see footnote "b" in the LSE table).
- (17) <u>CEL</u> Key number 38r is 1 of 3 studies for which Cancer Effect Levels were derived. The diamond symbol refers to a Cancer Effect Level for the test species-mouse. The number 38 corresponds to the entry in the LSE table.

- (18) Estimated Upper-Bound Human Cancer Risk Levels This is the range associated with the upper-bound for lifetime cancer risk of 1 in 10,000 to 1 in 10,000,000. These risk levels are derived from the EPA's Human Health Assessment Group's upper-bound estimates of the slope of the cancer dose response curve at low dose levels (q<sub>1</sub>\*).
- (19) <u>Key to LSE Figure</u> The Key explains the abbreviations and symbols used in the figure.

12

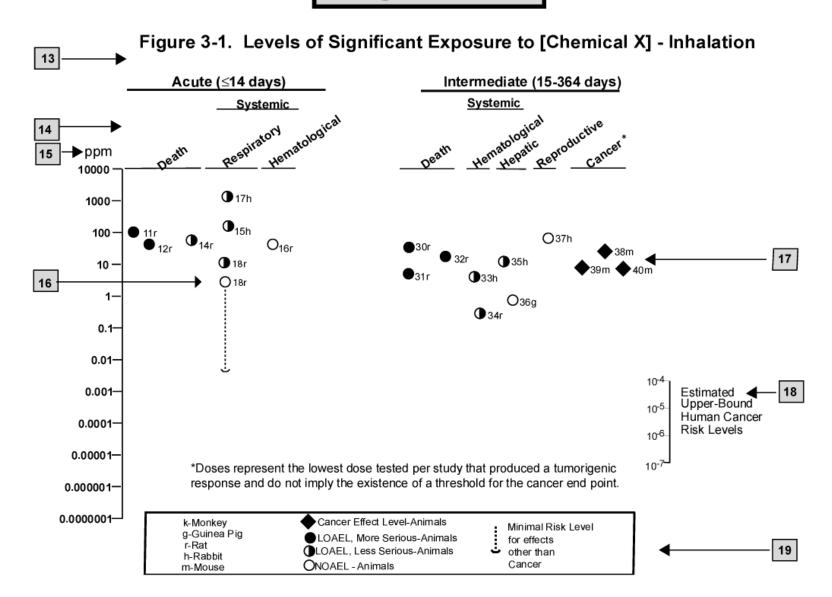
# **SAMPLE**

		Exposure			LO	AEL (effect	<b>:</b> )	
Key to figure <sup>a</sup>	Species	frequency/ duration	System	NOAEL (ppm)	Less serious (ppm)		Serious (ppm)	Reference
INTERMED	JATE EXPO	OSURE 6	7	8	9			10
Systemic	9	9	9	9	9			9
18	Rat	13 wk 5 d/wk 6 hr/d	Resp	3 <sup>b</sup>	10 (hyperplasia)			Nitschke et al. 1981
CHRONIC	EXPOSUR	E				11	 ]	
Cancer						9		
38	Rat	18 mo 5 d/wk 7 hr/d				20	(CEL, multiple organs)	Wong et al. 198
39	Rat	89–104 wk 5 d/wk 6 hr/d				10	(CEL, lung tumors, nasal tumors)	NTP 1982
40	Mouse	79–103 wk 5 d/wk 6 hr/d				10	(CEL, lung tumors, hemangiosarcomas)	NTP 1982

<sup>&</sup>lt;sup>a</sup> The number corresponds to entries in Figure 3-1.

<sup>&</sup>lt;sup>b</sup> Used to derive an intermediate inhalation Minimal Risk Level (MRL) of 5 x 10<sup>-3</sup> ppm; dose adjusted for intermittent exposure and divided by an uncertainty factor of 100 (10 for extrapolation from animal to humans, 10 for human variability).

# SAMPLE



COBALT C-1

#### APPENDIX C

# ACRONYMS, ABBREVIATIONS, AND SYMBOLS

ACGIH American Conference of Governmental Industrial Hygienists

ADI Acceptable Daily Intake

ADME Absorption, Distribution, Metabolism, and Excretion

AFID alkali flame ionization detector

AFOSH Air Force Office of Safety and Health

AML acute myeloid leukemia

AOAC Association of Official Analytical Chemists

atm atmosphere

ATSDR Agency for Toxic Substances and Disease Registry

AWQC Ambient Water Quality Criteria
BAT Best Available Technology
BCF bioconcentration factor
BEI Biological Exposure Index
BSC Board of Scientific Counselors

C Centigrade CAA Clean Air Act

CAG Cancer Assessment Group of the U.S. Environmental Protection Agency

CAS Chemical Abstract Services

CDC Centers for Disease Control and Prevention

CEL Cancer Effect Level

CELDS Computer-Environmental Legislative Data System

CERCLA Comprehensive Environmental Response, Compensation, and Liability Act

CFR Code of Federal Regulations

Ci curie

CL ceiling limit value

CLP Contract Laboratory Program

cm centimeter

CML chronic myeloid leukemia CNS central nervous system

CPSC Consumer Products Safety Commission

CWA Clean Water Act

d day Derm dermal

DHEW Department of Health, Education, and Welfare DHHS Department of Health and Human Services

DNA deoxyribonucleic acid DOD Department of Defense DOE Department of Energy DOL Department of Labor

DOT Department of Transportation

DOT/UN/ Department of Transportation/United Nations/

NA/IMCO North America/International Maritime Dangerous Goods Code

DWEL Drinking Water Exposure Level ECD electron capture detection

ECG/EKG electrocardiogram EEG electroencephalogram

APPENDIX C

EEGL Emergency Exposure Guidance Level EPA Environmental Protection Agency

F Fahrenheit

F<sub>1</sub> first-filial generation

FAO Food and Agricultural Organization of the United Nations

FDA Food and Drug Administration

FEMA Federal Emergency Management Agency

FIFRA Federal Insecticide, Fungicide, and Rodenticide Act

FPD flame photometric detection

fpm feet per minute

ft foot

FR Federal Register

g gram

GC gas chromatography Gd gestational day gen generation

GLC gas liquid chromatography
GPC gel permeation chromatography

HPLC high-performance liquid chromatography

hr hour

HRGC high resolution gas chromatography HSDB Hazardous Substance Data Bank

IDLH Immediately Dangerous to Life and Health IARC International Agency for Research on Cancer

ILO International Labor Organization

in inch

IRIS Integrated Risk Information System

Kd adsorption ratio kg kilogram kkg metric ton

 $K_{oc}$  organic carbon partition coefficient  $K_{ow}$  octanol-water partition coefficient

L liter

 $\begin{array}{ll} LC & liquid chromatography \\ LC_{Lo} & lethal concentration, low \\ LC_{50} & lethal concentration, 50\% kill \\ \end{array}$ 

 $\begin{array}{ccc} \text{LD}_{\text{Lo}} & \text{lethal dose, low} \\ \text{LD}_{50} & \text{lethal dose, 50\% kill} \\ \text{LT}_{50} & \text{lethal time, 50\% kill} \end{array}$ 

LOAEL lowest-observed-adverse-effect level LSE Levels of Significant Exposure

m meter

MA trans, trans-muconic acid MAL Maximum Allowable Level

mCi millicurie

MCL Maximum Contaminant Level
MCLG Maximum Contaminant Level Goal

mg milligram
min minute
mL milliliter
mm millimeter

mm Hg millimeters of mercury

mmol millimole mo month

mppcf millions of particles per cubic foot

MRL Minimal Risk Level MS mass spectrometry

NAAQS National Ambient Air Quality Standard

NAS National Academy of Science

NATICH National Air Toxics Information Clearinghouse

NATO North Atlantic Treaty Organization NCE normochromatic erythrocytes NCI National Cancer Institute

NIEHS National Institute of Environmental Health Sciences
NIOSH National Institute for Occupational Safety and Health
NIOSHTIC NIOSH's Computerized Information Retrieval System

NFPA National Fire Protection Association

ng nanogram

NLM National Library of Medicine

nm nanometer

NHANES National Health and Nutrition Examination Survey

nmol nanomole

NOAELno-observed-adverse-effect levelNOESNational Occupational Exposure SurveyNOHSNational Occupational Hazard Survey

NPD nitrogen phosphorus detection

NPDES National Pollutant Discharge Elimination System

NPL National Priorities List

NR not reported

NRC National Research Council

NS not specified

NSPS New Source Performance Standards
NTIS National Technical Information Service

NTP National Toxicology Program ODW Office of Drinking Water, EPA

OERR Office of Emergency and Remedial Response, EPA

OHM/TADS Oil and Hazardous Materials/Technical Assistance Data System

OPP Office of Pesticide Programs, EPA

OPPTS Office of Prevention, Pesticides and Toxic Substances, EPA

OPPT Office of Pollution Prevention and Toxics, EPA OSHA Occupational Safety and Health Administration

OSW Office of Solid Waste, EPA OTS Office of Toxic Substances

OW Office of Water

OWRS Office of Water Regulations and Standards, EPA

PAH Polycyclic Aromatic Hydrocarbon

PBPD Physiologically Based Pharmacodynamic PBPK Physiologically Based Pharmacokinetic

PCE polychromatic erythrocytes PEL permissible exposure limit PID photo ionization detector

pg picogram

pmol picomole

PHS Public Health Service PMR proportionate mortality ratio

ppb parts per billion ppm parts per million ppt parts per trillion

PSNS Pretreatment Standards for New Sources
REL recommended exposure level/limit

RfC Reference Concentration

RfD Reference Dose RNA ribonucleic acid

RTECS Registry of Toxic Effects of Chemical Substances

RQ Reportable Quantity

SARA Superfund Amendments and Reauthorization Act

SCE sister chromatid exchange

sec second

SIC Standard Industrial Classification

SIM selected ion monitoring

SMCL Secondary Maximum Contaminant Level

SMR standard mortality ratio

SNARL Suggested No Adverse Response Level

SPEGL Short-Term Public Emergency Guidance Level

STEL short term exposure limit STORET Storage and Retrieval

TD<sub>50</sub> toxic dose, 50% specific toxic effect

TLV threshold limit value
TOC Total Organic Compound
TPQ Threshold Planning Quantity
TRI Toxics Release Inventory
TSCA Toxic Substances Control Act
TRI Toxics Release Inventory
TWA time-weighted average

U.S. United States
UF uncertainty factor

VOC Volatile Organic Compound

yr year

WHO World Health Organization

wk week

> greater than

> greater than or equal to

= equal to < less than

 $\leq$  less than or equal to

APPENDIX C

$q_1^*$	cancer slope factor
_	negative
+	positive
(+)	weakly positive result
(-)	weakly negative result

COBALT D-1

#### APPENDIX D

# OVERVIEW OF BASIC RADIATION PHYSICS, CHEMISTRY, AND BIOLOGY

Understanding the basic concepts in radiation physics, chemistry, and biology is important to the evaluation and interpretation of radiation-induced adverse health effects and to the derivation of radiation protection principles. This appendix presents a brief overview of the areas of radiation physics, chemistry, and biology and is based to a large extent on the reviews of Mettler and Moseley (1985), Hobbs and McClellan (1986), Eichholz (1982), Hendee (1973), Cember (1996), and Early et al. (1979).

#### D.1 RADIONUCLIDES AND RADIOACTIVITY

The substances we call elements are composed of atoms. Atoms in turn are made up of neutrons, protons and electrons: neutrons and protons in the nucleus and electrons in a cloud of orbits around the nucleus. Nuclide is the general term referring to any nucleus along with its orbital electrons. The nuclide is characterized by the composition of its nucleus and hence by the number of protons and neutrons in the nucleus. All atoms of an element have the same number of protons (this is given by the atomic number) but may have different numbers of neutrons (this is reflected by the atomic mass numbers or atomic weight of the element). Atoms with different atomic mass but the same atomic numbers are referred to as isotopes of an element.

The numerical combination of protons and neutrons in most nuclides is such that the nucleus is quantum mechanically stable and the atom is said to be stable, i.e., not radioactive; however, if there are too few or too many neutrons, the nucleus is unstable and the atom is said to be radioactive. Unstable nuclides undergo radioactive transformation, a process in which a neutron or proton converts into the other and a beta particle is emitted, or else an alpha particle is emitted. Each type of decay is typically accompanied by the emission of gamma rays. These unstable atoms are called radionuclides; their emissions are called ionizing radiation; and the whole property is called radioactivity. Transformation or decay results in the formation of new nuclides some of which may themselves be radionuclides, while others are stable nuclides. This series of transformations is called the decay chain of the radionuclide. The first radionuclide in the chain is called the parent; the subsequent products of the transformation are called progeny, daughters, or decay products.

In general there are two classifications of radioactivity and radionuclides: natural and artificial (manmade). Naturally-occurring radioactive material (NORM) exists in nature and no additional energy is necessary to place them in an unstable state. Natural radioactivity is the property of some naturally occurring, usually heavy elements, that are heavier than lead. Radionuclides, such as radium and uranium, primarily emit alpha particles. Some lighter elements such as carbon-14 and tritium (hydrogen-3) primarily emit beta particles as they transform to a more stable atom. Natural radioactive atoms heavier than lead cannot attain a stable nucleus heavier than lead. Everyone is exposed to background radiation from naturally-occurring radionuclides throughout life. This background radiation is the major source of radiation exposure to man and arises from several sources. The natural background exposures are frequently used as a standard of comparison for exposures to various artificial sources of ionizing radiation.

Artificial radioactive atoms are produced either as a by-product of fission of uranium or plutonium atoms in a nuclear reactor or by bombarding stable atoms with particles, such as neutrons or protons, directed at the stable atoms with high velocity. These artificially produced radioactive elements usually decay by emission of particles, such as positive or negative beta particles and one or more high energy photons (gamma rays). Unstable (radioactive) atoms of any element can be produced.

Both naturally occurring and artificial radioisotopes find application in medicine, industrial products, and consumer products. Some specific radioisotopes, called fall-out, are still found in the environment as a result of nuclear weapons use or testing.

#### **D.2 RADIOACTIVE DECAY**

# **D.2.1 Principles of Radioactive Decay**

The stability of an atom is the result of the balance of the forces of the various components of the nucleus. An atom that is unstable (radionuclide) will release energy (decay) in various ways and transform to stable atoms or to other radioactive species called daughters, often with the release of ionizing radiation. If there are either too many or too few neutrons for a given number of protons, the resulting nucleus may undergo transformation. For some elements, a chain of daughter decay products may be produced until stable atoms are formed. Radionuclides can be characterized by the type and energy of the radiation emitted, the rate of decay, and the mode of decay. The mode of decay indicates how a parent compound undergoes transformation. Radiations considered here are primarily of nuclear origin, i.e., they arise from nuclear excitation, usually caused by the capture of charged or uncharged nucleons by a nucleus, or by the radioactive decay or transformation of an unstable nuclide. The type of radiation may be categorized as charged or uncharged particles, protons, and fission products) or electromagnetic radiation (gamma rays and x rays). Table D-1 summarizes the basic characteristics of the more common types of radiation encountered.

# D.2.2 Half-Life and Activity

For any given radionuclide, the rate of decay is a first-order process that is constant, regardless of the radioactive atoms present and is characteristic for each radionuclide. The process of decay is a series of random events; temperature, pressure, or chemical combinations do not effect the rate of decay. While it may not be possible to predict exactly which atom is going to undergo transformation at any given time, it is possible to predict, on average, the fraction of the radioactive atoms that will transform during any interval of time.

The *activity* is a measure of the quantity of radioactive material. For these radioactive materials it is customary to describe the activity as the number of disintegrations (transformations) per unit time. The unit of activity is the curie (Ci), which was originally related to the activity of one gram of radium, but is now defined as that quantity of radioactive material in which there are:

1 curie (Ci) =  $3.7x10^{10}$  disintegrations (transformations)/second (dps) or  $2.22x10^{12}$  disintegrations (transformations)/minute (dpm).

The SI unit of activity is the becquerel (Bq); 1 Bq = that quantity of radioactive material in which there is 1 transformation/second. Since activity is proportional to the number of atoms of the radioactive material, the quantity of any radioactive material is usually expressed in curies, regardless of its purity or concentration. The transformation of radioactive nuclei is a random process, and the number of transformations is directly proportional to the number of radioactive atoms present. For any pure radioactive substance, the rate of decay is usually described by its radiological half-life,  $T_R$ , i.e., the time it takes for a specified source material to decay to half its initial activity. The specific activity is an indirect measure of the rate of decay, and is defined as the activity per unit mass or per unit volume. The higher the specific activity of a radioisotope, the faster it is decaying.

The activity of a radionuclide at time t may be calculated by:

$$A = A_o e^{-0.693t/Trad}$$

where A is the activity in dps or curies or becquerels,  $A_o$  is the activity at time zero, t is the time at which measured, and  $T_{rad}$  is the radiological half-life of the radionuclide ( $T_{rad}$  and t must be in the same units of time). The time when the activity of a sample of radioactivity becomes one-half its original value is the radioactive half-life and is expressed in any suitable unit of time.

Table D-1. Characteristics of Nuclear Radiatio	Table D-1.	Characteristics	of Nuclear	Radiation
------------------------------------------------	------------	-----------------	------------	-----------

			Typical	Path	length <sup>b</sup>	
Radiation	Rest mass <sup>a</sup>	Charge	energy range	Air	Solid	Comments
Alpha (α)	4.00 amu	+2	4–10 MeV	5–10 cm	25–80 μm	Identical to ionized He nucleus
Negatron (β <sup>-</sup> )	5.48x10 <sup>-4</sup> amu; 0.51 MeV	-1	0–4 Mev	0–10 m	0–1 cm	Identical to electron
Positron $(\beta^+)$	5.48x10 <sup>-4</sup> amu; 0.51 Mev	+1	0-4 Mev	0–10 m	0–1 cm	Identical to electron except for sign of charge
Neutron	1.0086 amu; 939.55 MeV	0	0–15 MeV	b	b	Free half-life: 16 min
$X \; ray_{\;(e.m.\;\;photon)}$	_	0	5 keV-100 keV	b	b	Photon from transition of an electron between atomic orbits
Gamma (p) (e.m. photon)	_	0	10 keV–3 MeV	b	b	Photon from nuclear transformation

<sup>&</sup>lt;sup>a</sup> The rest mass (in amu) has an energy equivalent in MeV that is obtained using the equation E=mc<sup>2</sup>, where 1 amu = 932 MeV.

amu = atomic mass unit; e.m. = electromagnetic; MeV = MegaElectron Volts

The specific activity is a measure of activity, and is fined as the activity per unit mass or per unit volume. This activity is usually expressed in curies per gram and may be calculated by

curies/gram = 
$$1.3 \times 10^8 / (T_{rad})$$
 (atomic weight) or 
$$[3.577 \times 10^5 \times mass(g)] / [T_{rad} \times atomic weight]$$

where  $T_{rad}$  is the radiological half-life in days.

In the case of radioactive materials contained in living organisms, an additional consideration is made for the reduction in observed activity due to regular processes of elimination of the respective chemical or biochemical substance from the organism. This introduces a rate constant called the biological half-life

<sup>&</sup>lt;sup>b</sup> Path lengths are not applicable to x- and gamma rays since their intensities decrease exponentially; path lengths in solid tissue are variable, depending on particle energy, electron density of material, and other factors.

 $(T_{biol})$  which is the time required for biological processes to eliminate one-half of the activity. This time is virtually the same for both stable and radioactive isotopes of any given element.

Under such conditions the time required for a radioactive element to be halved as a result of the combined action of radioactive decay and biological elimination is the effective clearance half-time:

$$T_{eff} = (T_{biol} \times T_{rad}) / (T_{biol} + T_{rad}).$$

Table D-2 presents representative effective half-lives of particular interest.

Table D-2. Half-Lives of Some Radionuclides in Adult Body Organs

			Half-life <sup>a</sup>	
Radionuclide	Critical organ	Physical	Biological	Effective
Uranium-238	Kidney	4,460,000,000 y	4 d	4 d
Hydrogen-3 <sup>b</sup> (Tritium)	Whole body	12.3 y	10 d	10 d
Iodine-131	Thyroid	8 d	80 d	7.3 d
Strontium-90	Bone	28 y	50 y	18 y
Plutonium-239	Bone surface	24,400 y	50 y	50 y
	Lung	24,400 y	500 d	500 d
Cobalt-60	Whole body	5.3 y	99.5 d	95 d
Iron-55	Spleen	2.7 y	600 d	388 d
Iron-59	Spleen	45.1 d	600 d	42 d
Manganese-54	Liver	303 d	25 d	23 d
Cesium-137	Whole body	30 y	70 d	70 d

 $<sup>^{</sup>a}d = days, y = years$ 

#### D.2.3 Interaction of Radiation with Matter

Both ionizing and nonionizing radiation will interact with materials; that is, radiation will lose kinetic energy to any solid, liquid or gas through which it passes by a variety of mechanisms. The transfer of energy to a medium by either electromagnetic or particulate radiation may be sufficient to cause formation of ions. This process is called ionization. Compared to other types of radiation that may be absorbed, such as ultraviolet radiation, ionizing radiation deposits a relatively large amount of energy into a small volume

The method by which incident radiation interacts with the medium to cause ionization may be direct or indirect. Electromagnetic radiations (x rays and gamma photons) are indirectly ionizing; that is, they give up their energy in various interactions with cellular molecules, and the energy is then utilized to produce a fast-moving charged particle such as an electron. It is the electron that then may react with a target molecule. This particle is called a "primary ionizing particle. Charged particles, in contrast, strike the tissue or medium and directly react with target molecules, such as oxygen or water. These particulate

<sup>&</sup>lt;sup>b</sup>Mixed in body water as tritiated water

radiations are directly ionizing radiations. Examples of directly ionizing particles include alpha and beta particles. Indirectly ionizing radiations are always more penetrating than directly ionizing particulate radiations.

Mass, charge, and velocity of a particle all affect the rate at which ionization occurs. The higher the charge of the particle and the lower the velocity, the greater the propensity to cause ionization. Heavy, highly charged particles, such as alpha particles, lose energy rapidly with distance and, therefore, do not penetrate deeply. The result of these interaction processes is a gradual slowing down of any incident particle until it is brought to rest or "stopped" at the end of its range.

#### D.2.4 Characteristics of Emitted Radiation

**D.2.4.1 Alpha Emission.** In alpha emission, an alpha particle consisting of two protons and two neutrons is emitted with a resulting decrease in the atomic mass number by four and reduction of the atomic number of two, thereby changing the parent to a different element. The alpha particle is identical to a helium nucleus consisting of two neutrons and two protons. It results from the radioactive decay of some heavy elements such as uranium, plutonium, radium, thorium, and radon. All alpha particles emitted by a given radioisotope have the same energy. Most of the alpha particles that are likely to be found have energies in the range of about 4 to 8 MeV, depending on the isotope from which they came.

The alpha particle has an electrical charge of +2. Because of this double positive charge and their size, alpha particles have great ionizing power and, thus, lose their kinetic energy quickly. This results in very little penetrating power. In fact, an alpha particle cannot penetrate a sheet of paper. The range of an alpha particle (the distance the charged particle travels from the point of origin to its resting point) is about 4 cm in air, which decreases considerably to a few micrometers in tissue. These properties cause alpha emitters to be hazardous only if there is internal contamination (i.e., if the radionuclide is inside the body).

**D.2.4.2 Beta Emission.** A beta particle (&) is a high-velocity electron ejected from a disintegrating nucleus. The particle may be either a negatively charged electron, termed a negatron (&-) or a positively charged electron, termed a positron (&+). Although the precise definition of "beta emission" refers to both &- and &+, common usage of the term generally applies only to the negative particle, as distinguished from the positron emission, which refers to the &+ particle.

**D.2.4.2.1 Beta Negative Emission.** Beta particle (&) emission is another process by which a radionuclide, with a neutron excess achieves stability. Beta particle emission decreases the number of neutrons by one and increases the number of protons by one, while the atomic mass number remains unchanged. This transformation results in the formation of a different element. The energy spectrum of beta particle emission ranges from a certain maximum down to zero with the mean energy of the spectrum being about one-third of the maximum. The range in tissue is much less. Beta negative emitting radionuclides can cause injury to the skin and superficial body tissues, but mostly present an internal contamination hazard.

**D.2.4.2.2 Positron Emission.** In cases in which there are too many protons in the nucleus, positron emission may occur. In this case a proton may be thought of as being converted into a neutron, and a positron (&+) is emitted. This increases the number of neutrons by one, decreases the number of protons by one, and again leaves the atomic mass number unchanged. The gamma radiation resulting from the annihilation (see glossary) of the positron makes all positron emitting isotopes more of an external radiation hazard than pure & emitters of equal energy.

<sup>&</sup>lt;sup>1</sup>Neutrinos also accompany negative beta particles and positron emissions.

**D.2.4.2.3 Gamma Emission.** Radioactive decay by alpha, beta, or positron emission, or electron capture often leaves some of the energy resulting from these changes in the nucleus. As a result, the nucleus is raised to an excited level. None of these excited nuclei can remain in this high-energy state. Nuclei release this energy returning to ground state or to the lowest possible stable energy level. The energy released is in the form of gamma radiation (high energy photons) and has an energy equal to the change in the energy state of the nucleus. Gamma and x rays behave similarly but differ in their origin; gamma emissions originate in the nucleus while x rays originate in the orbital electron structure or from rapidly changing the velocity of an electron (e.g., as occurs when shielding high energy beta particles or stopping the electron beam in an x ray tube).

#### D.3 ESTIMATION OF ENERGY DEPOSITION IN HUMAN TISSUES

Two forms of potential radiation exposures can result: internal and external. The term exposure denotes physical interaction of the radiation emitted from the radioactive material with cells and tissues of the human body. An exposure can be "acute" or "chronic" depending on how long an individual or organ is exposed to the radiation. Internal exposures occur when radionuclides, which have entered the body (e.g., through the inhalation, ingestion, or dermal pathways), undergo radioactive decay resulting in the deposition of energy to internal organs. External exposures occur when radiation enters the body directly from sources located outside the body, such as radiation emitters from radionuclides on ground surfaces, dissolved in water, or dispersed in the air. In general, external exposures are from material emitting gamma radiation, which readily penetrate the skin and internal organs. Beta and alpha radiation from external sources are far less penetrating and deposit their energy primarily on the skin's outer layer. Consequently, their contribution to the absorbed dose of the total body dose, compared to that deposited by gamma rays, may be negligible.

Characterizing the radiation dose to persons as a result of exposure to radiation is a complex issue. It is difficult to: (1) measure internally the amount of energy actually transferred to an organic material and to correlate any observed effects with this energy deposition; and (2) account for and predict secondary processes, such as collision effects or biologically triggered effects, that are an indirect consequence of the primary interaction event.

## D.3.1 Dose/Exposure Units

- **D.3.1.1 Roentgen.** The roentgen (R) is a unit of x or gamma-ray exposure and is a measured by the amount of ionization caused in air by gamma or x radiation. One roentgen produces  $2.58 \times 10^{-4}$  coulomb per kilogram of air. In the case of gamma radiation, over the commonly encountered range of photon energy, the energy deposition in tissue for a dose of 1 R is about 0.0096 joules (J) /kg of tissue.
- **D.3.1.2 Absorbed Dose and Absorbed Dose Rate.** The absorbed dose is defined as the energy imparted by the incident radiation to a unit mass of the tissue or organ. The unit of absorbed dose is the rad; 1 rad = 100 erg/gram = 0.01 J/kg in any medium. An exposure of 1 R results in a dose to soft tissue of approximately 0.01 J/kg. The SI unit is the gray which is equivalent to 100 rad or 1 J/kg. Internal and external exposures from radiation sources are not usually instantaneous but are distributed over extended periods of time. The resulting rate of change of the absorbed dose to a small volume of mass is referred to as the absorbed dose rate in units of rad/unit time.

**D.3.1.3 Working Levels and Working Level Months.** Working level (WL) is a measure of the atmospheric concentration of radon and its short-lived progeny. One WL is defined as any combination of short-lived radon daughters (through polonium-214), per liter of air, that will result in the emission of  $1.3 \times 10^5$  MeV of alpha energy. An activity concentration of 100 pCi radon-222/L of air, in equilibrium with its daughters, corresponds approximately to a potential alpha-energy concentration of 1 WL. The WL unit can also be used for thoron daughters. In this case,  $1.3 \times 10^5$  MeV of alpha energy (1 WL) is released by the thoron daughters in equilibrium with 7.5 pCi thoron/L. The potential alpha energy exposure of miners is commonly expressed in the unit Working Level Month (WLM). One WLM corresponds to exposure to a concentration of 1 WL for the reference period of 170 hours, or more generally

WLM = concentration (WL) x exposure time (months) (one "month" = 170 working hours).

# **D.3.2 Dosimetry Models**

Dosimetry models are used to estimate the dose from internally deposited to radioactive substances. The models for internal dosimetry consider the amount of radionuclides entering the body, the factors affecting their movement or transport through the body, distribution and retention of radionuclides in the body, and the energy deposited in organs and tissues from the radiation that is emitted during spontaneous decay processes. The dose pattern for radioactive materials in the body may be strongly influenced by the route of entry of the material. For industrial workers, inhalation of radioactive particles with pulmonary deposition and puncture wounds with subcutaneous deposition have been the most frequent. The general population has been exposed via ingestion and inhalation of low levels of naturally occurring radionuclides as well as radionuclides from nuclear weapons testing.

The models for external dosimetry consider only the photon doses to organs of individuals who are immersed in air or are exposed to a contaminated object.

**D.3.2.1 Ingestion.** Ingestion of radioactive materials is most likely to occur from contaminated foodstuffs or water or eventual ingestion of inhaled compounds initially deposited in the lung. Ingestion of radioactive material may result in toxic effects as a result of either absorption of the radionuclide or irradiation of the gastrointestinal tract during passage through the tract, or a combination of both. The fraction of a radioactive material absorbed from the gastrointestinal tract is variable, depending on the specific element, the physical and chemical form of the material ingested, and the diet, as well as some other metabolic and physiological factors. The absorption of some elements is influenced by age, usually with higher absorption in the very young.

**D.3.2.2 Inhalation.** The inhalation route of exposure has long been recognized as being a major portal of entry for both nonradioactive and radioactive materials. The deposition of particles within the lung is largely dependent upon the size of the particles being inhaled. After the particle is deposited, the retention will depend upon the physical and chemical properties of the dust and the physiological status of the lung. The retention of the particle in the lung depends on the location of deposition, in addition to the physical and chemical properties of the particles. The converse of pulmonary retention is pulmonary clearance. There are three distinct mechanisms of clearance which operate simultaneously. Ciliary clearance acts only in the upper respiratory tract. The second and third mechanisms act mainly in the deep respiratory tract. These are phagocytosis and absorption. Phagocytosis is the engulfing of foreign bodies by alveolar macrophages and their subsequent removal either up the ciliary "escalator" or by entrance into the lymphatic system. Some inhaled soluble particles are absorbed into the blood and translocated to other organs and tissues.

#### **D.3.3 Internal Emitters**

An internal emitter is a radionuclide that is inside the body. The absorbed dose from internally deposited radioisotopes depends on the energy absorbed per unit tissue by the irradiated tissue. For a radioisotope distributed uniformly throughout an infinitely large medium, the concentration of absorbed energy must be equal to the concentration of energy emitted by the isotope. An infinitely large medium may be approximated by a tissue mass whose dimensions exceed the range of the particle. All alpha and most beta radiation will be absorbed in the organ (or tissue) of reference. Gamma-emitting isotope emissions are penetrating radiation, and a substantial fraction of gamma energy may be absorbed in tissue. The dose to an organ or tissue is a function of the effective retention half-time, the energy released in the tissue, the amount of radioactivity initially introduced, and the mass of the organ or tissue.

#### D.4 BIOLOGICAL EFFECTS OF RADIATION

When biological material is exposed to ionizing radiation, a chain of cellular events occurs as the ionizing particle passes through the biological material. A number of theories have been proposed to describe the interaction of radiation with biologically important molecules in cells and to explain the resulting damage to biological systems from those interactions. Many factors may modify the response of a living organism to a given dose of radiation. Factors related to the exposure include the dose rate, the energy of the radiation, and the temporal pattern of the exposure. Biological considerations include factors such as species, age, sex, and the portion of the body exposed. Several excellent reviews of the biological effects of radiation have been published, and the reader is referred to these for a more in-depth discussion (Brodsky 1996; Hobbs and McClellan 1986; ICRP 1984; Mettler and Moseley 1985; Rubin and Casarett 1968).

## D.4.1 Radiation Effects at the Cellular Level

According to Mettler and Moseley (1985), at acute doses up to 10 rad (100 mGy), single strand breaks in DNA may be produced. These single strand breaks may be repaired rapidly. With doses in the range of 50–500 rad (0.5–5 Gy), irreparable double-stranded DNA breaks are likely, resulting in cellular reproductive death after one or more divisions of the irradiated parent cell. At large doses of radiation, usually greater than 500 rad (5 Gy), direct cell death before division (interphase death) may occur from the direct interaction of free-radicals with essentially cellular macromolecules. Morphological changes at the cellular level, the severity of which are dose-dependent, may also be observed.

The sensitivity of various cell types varies. According to the Bergonie-Tribondeau law, the sensitivity of cell lines is directly proportional to their mitotic rate and inversely proportional to the degree of differentiation (Mettler and Moseley 1985). Rubin and Casarett (1968) devised a classification system that categorized cells according to type, function, and mitotic activity. The categories range from the most sensitive type, "vegetative intermitotic cells," found in the stem cells of the bone marrow and the gastrointestinal tract, to the least sensitive cell type, "fixed postmitotic cells," found in striated muscles or long-lived neural tissues.

Cellular changes may result in cell death, which if extensive, may produce irreversible damage to an organ or tissue or may result in the death of the individual. If the cell recovers, altered metabolism and function may still occur, which may be repaired or may result in the manifestation of clinical symptoms. These changes may also be expressed at a later time as tumors or cellular mutations, which may result in abnormal tissue

# D.4.2 Radiation Effects at the Organ Level

In most organs and tissues the injury and the underlying mechanism for that injury are complex and may involve a combination of events. The extent and severity of this tissue injury are dependent upon the radiosensitivity of the various cell types in that organ system. Rubin and Casarett (1968) describe and schematically display the events following radiation in several organ system types. These include: a rapid renewal system, such as the gastrointestinal mucosa; a slow renewal system, such as the pulmonary epithelium; and a nonrenewal system, such as neural or muscle tissue. In the rapid renewal system, organ injury results from the direct destruction of highly radiosensitive cells, such as the stem cells in the bone marrow. Injury may also result from constriction of the microcirculation and from edema and inflammation of the basement membrane, designated as the histohematic barrier (HHB), which may progress to fibrosis. In slow renewal and nonrenewal systems, the radiation may have little effect on the parenchymal cells, but ultimate parenchymal atrophy and death over several months result from HHB fibrosis and occlusion of the microcirculation.

#### D.4.3 Low Level Radiation Effects

Cancer is the major latent harmful effect produced by ionizing radiation and the one that most people exposed to radiation are concerned about. The ability of alpha, beta, and gamma radiation to produce cancer in virtually every tissue and organ in laboratory animals has been well-demonstrated. The development of cancer is not an immediate effect. Radiation-induced leukemia has the shortest latent period at 2 years, while other radiation induced cancers have latent periods >20 years. The mechanism by which cancer is induced in living cells is complex and is a topic of intense study. Exposure to ionizing radiation can produce cancer at any site within the body; however, some sites appear to be more common than others, such as the breast, lung, stomach, and thyroid.

DNA is a major target molecule during exposure to ionizing radiation. Other macromolecules, such as lipids and proteins, are also at risk of damage when exposed to ionizing radiation. The genotoxicity of ionizing radiation is an area of intense study, as damage to the DNA is ultimately responsible for many of the adverse toxicological effects ascribed to ionizing radiation, including cancer. Damage to genetic material is basic to developmental or teratogenic effects, as well. However, for effects other than cancer, there is little evidence of human effects at low levels of exposure.

#### D.5 UNITS IN RADIATION PROTECTION AND REGULATION

# D.5.1 Dose Equivalent and Dose Equivalent Rate

Dose equivalent or rem is a special radiation protection quantity that is used, for administrative and radiation safety purposes only, to express the absorbed dose in a manner which considers the difference in biological effectiveness of various kinds of ionizing radiation. The ICRU has defined the dose equivalent, H, as the product of the absorbed dose, D, and the quality factor, Q, at the point of interest in biological tissue. This relationship is expressed as  $H = D \times Q$ . The dose equivalent concept is applicable only to doses that are not great enough to produce biomedical effects.

The quality factor is a dimensionless quantity that depends in part on the stopping power for charged particles, and it accounts for the differences in biological effectiveness found among the types of radiation. Originally relative biological effectiveness (RBE) was used rather than Q to define the quantity, rem, which was of use in risk assessment. The generally accepted values for quality factors for

various radiation types are provided in Table D-3. The dose equivalent rate is the time rate of change of the dose equivalent to organs and tissues and is expressed as rem/unit time or sievert/unit time.

Table D-3. Quality Factors (Q) and Absorbed Dose Equivalencies

Type of radiation	Quality factor (Q)	Absorbed dose equal to a unit dose equivalent*
X, gamma, or beta radiation	1	1
Alpha particles, multiple-charged particles, fission fragments and heavy particles of unknown charge	20	0.05
Neutrons of unknown energy	10	0.1
High-energy protons	10	0.1

<sup>\*</sup>Absorbed dose in rad equal to 1 rem or the absorbed dose in gray equal to 1 sievert.

Source: USNRC. 1999. Standards for the protection against radiation, table 1004(b).1. 10 CFR 20.1004. U.S. Nuclear Regulatory Commission, Washington, D.C.

# **D.5.2 Relative Biological Effectiveness**

RBE is used to denote the experimentally determined ratio of the absorbed dose from one radiation type to the absorbed dose of a reference radiation required to produce an identical biologic effect under the same conditions. Gamma rays from cobalt-60 and 200–250 keV x-rays have been used as reference standards. The term RBE has been widely used in experimental radiobiology, and the term quality factor used in calculations of dose equivalents for radiation safety purposes (ICRP 1977; NCRP 1971; UNSCEAR 1982). RBE applies only to a specific biological end point, in a specific exposure, under specific conditions to a specific species. There are no generally accepted values of RBE.

# D.5.3 Effective Dose Equivalent and Effective Dose Equivalent Rate

The absorbed dose is usually defined as the mean absorbed dose within an organ or tissue. This represents a simplification of the actual problem. Normally when an individual ingests or inhales a radionuclide or is exposed to external radiation that enters the body (gamma), the dose is not uniform throughout the whole body. The simplifying assumption is that the detriment will be the same whether the body is uniformly or non-uniformly irradiated. In an attempt to compare detriment from absorbed dose of a limited portion of the body with the detriment from total body dose, the ICRP (1977) has derived a concept of effective dose equivalent. The effective dose equivalent, H<sub>E</sub>, is

$$H_E =$$
(the sum of)  $W_t H_t$ 

where  $H_t$  is the dose equivalent in the tissue,  $W_t$  is the weighting factor, which represents the estimated proportion of the stochastic risk resulting from tissue, T, to the stochastic risk when the whole body is uniformly irradiated for occupational exposures under certain conditions (ICRP 1977). Weighting factors for selected tissues are listed in Table D-4.

The ICRU (1980), ICRP (1984), and NCRP (1985) now recommend that the rad, roentgen, curie, and rem be replaced by the SI units: gray (Gy), Coulomb per kilogram (C/kg), Becquerel (Bq), and sievert (Sv), respectively. The relationship between the customary units and the international system of units (SI) for radiological quantities is shown in Table D-5.

Table D-4. Weighting Factors for Calculating Effective Dose **Equivalent for Selected Tissues** 

		Weighting factor	
Tissue	ICRP60	NCRP115/ ICRP60	NRC
Bladder	0.040	0.05	_
Bone marrow	0.143	0.12	0.12
Bone surface	0.009	0.01	0.03
Breast	0.050	0.05	0.15
Colon	0.141	0.12	_
Liver	0.022	0.05	_
Lung	0.111	0.12	0.12
Esophagus	0.034	0.05	_
Ovary	0.020	0.05	_
Skin	0.006	0.01	_
Stomach	0.139	0.12	_
Thyroid	0.021	0.05	0.03
Gonads	0.183	0.20	0.25
subtotal	0.919	0.99	0.70
Remainder	0.081	0.05	0.30

ICRP60 = International Commission on Radiological Protection, 1990 Recommendations of the ICRP;

NCRP115 = National Council on Radiation Protection and Measurements. 1993. Risk Estimates for Radiation Protection, Report 115. Bethesda, Maryland; NRC = Nuclear Regulatory Commission.

NRC = Nuclear Regulatory Commission, Title 10, Code of Federal Regulations, Part 20

Table D-5. Comparison of Common and SI Units for Radiation Quantities

Quantity	Customary units	Definition	SI units	Definition
Activity (A)	curie (Ci)	$3.7x10^{10}$ transformations s <sup>-1</sup>	becquerel (Bq)	S <sup>-1</sup>
Absorbed dose (D)	rad (rad)	10 <sup>-2</sup> Jkg <sup>-1</sup>	gray (Gy) Jkg <sup>-1</sup>	
Absorbed dose rate (Š)	rad per second (rad s <sup>-1</sup> )	10 <sup>-2</sup> Jkg <sup>-1</sup> s <sup>-1</sup>	gray per second (Gy s <sup>-1</sup> )	Jkg <sup>-1</sup> s <sup>-1</sup>
Dose equivalent (H)	rem (rem)	10 <sup>-2</sup> Jkg <sup>-1</sup>	sievert (Sv)	Jkg <sup>-1</sup>
Dose equivalent rate ( )	rem per second (rem s <sup>-1</sup> )	10 <sup>-2</sup> Jkg <sup>-1</sup> s <sup>-1</sup>	sievert per second (Sv s <sup>-1</sup> )	Jkg <sup>-1</sup> s <sup>-1</sup>
Linear energy transfer (LET)	kiloelectron volts per micrometer (keV µm <sup>-1</sup> )	1.602x10 <sup>-10</sup> Jm <sup>-1</sup>	kiloelectron volts per micrometer (keV μm <sup>-1</sup> )	1.602x10 <sup>-10</sup> Jm <sup>-1</sup>

Jkg<sup>-1</sup> = Joules per kilogram; Jkg<sup>-1</sup>s<sup>-1</sup> = Joules per kilogram per second; Jm<sup>-1</sup> = Joules per meter; s<sup>-1</sup> = per second

#### REFERENCES FOR APPENDIX D

ATSDR. 1990a. Toxicological profile for thorium. U.S. Department of Health and Human Services. Public Health Service. Agency for Toxic Substances and Disease Registry. Atlanta, GA.

ATSDR. 1990b. Toxicological profile for radium. U.S. Department of Health and Human Services. Public Health Service. Agency for Toxic Substances and Disease Registry. Atlanta, GA.

ATSDR. 1990c. Toxicological profile for radon. U.S. Department of Health and Human Services. Public Health Service. Agency for Toxic Substances and Disease Registry. Atlanta, GA.

ATSDR. 1999. Toxicological profile for uranium. U.S. Department of Health and Human Services. Public Health Service. Agency for Toxic Substances and Disease Registry. Atlanta, GA.

BEIR III. 1980. The effects on populations of exposure to low levels of ionizing radiation. Committee on the Biological Effects of Ionizing Radiations, National Research Council. Washington, DC: National Academy Press.

BEIR IV. 1988. Health risks of radon and other internally deposited alpha emitters. Committee on the Biological Effects of Ionizing Radiations, National Research Council. Washington, DC: National Academy Press.

BEIR V. 1988. Health effects of exposure to low levels of ionizing radiation. Committee on the Biological Effects of Ionizing Radiations, National Research Council. Washington, DC: National Academy Press.

Brodsky A. 1996. Review of radiation risks and uranium toxicity with application to decisions associated with decommissioning clean-up criteria. Hebron, Connecticut: RSA Publications.

Cember H. 1996. Introduction to health physics. New York., NY: McGraw Hill.

Early P, Razzak M, Sodee D. 1979. Nuclear medicine technology. 2nd ed. St. Louis: C.V. Mosby Company.

Eichholz G. 1982. Environmental aspects of nuclear power. Ann Arbor, MI: Ann Arbor Science.

Hendee W. 1973. Radioactive isotopes in biological research. New York, NY: John Wiley and Sons.

Hobbs C, McClellan R. 1986. Radiation and radioactive materials. In: Doull J, et al., eds. Casarett and Doull's Toxicology. 3rd ed. New York, NY: Macmillan Publishing Co., Inc., 497-530.

ICRP. 1977. International Commission on Radiological Protection. Recommendations of the International Commission on Radiological Protection. ICRP Publication 26. Vol 1. No. 3. Oxford: Pergamon Press.

ICRP. 1979. International Commission on Radiological Protection. Limits for intakes of radionuclides by workers. ICRP Publication 20. Vol. 3. No. 1-4. Oxford: Pergamon Press.

ICRP. 1979. Limits for Intakes of Radionuclides by Workers. Publication 30. International Commission on Radiological Protection. Pergamon Press.

# COBALT D-14 APPENDIX D

ICRP. 1984. International Commission on Radiological Protection. A compilation of the major concepts and quantities in use by ICRP. ICRP Publication 42. Oxford: Pergamon Press.

ICRP. 1990. International Commission on Radiological Protection 1990 Recommendations of the ICRP

ICRU. 1980. International Commission on Radiation Units and Measurements. ICRU Report No. 33. Washington, DC.

James A. 1987. A reconsideration of cells at risk and other key factors in radon daughter dosimetry. In: Hopke P, ed. Radon and its decay products: Occurrence, properties and health effects. ACS Symposium Series 331. Washington, DC: American Chemical Society, 400-418.

James A, Roy M. 1987. Dosimetric lung models. In: Gerber G, et al., ed. Age-related factors in radionuclide metabolism and dosimetry. Boston: Martinus Nijhoff Publishers, 95-108.

Kondo S. 1993. Health effects of low-level radiation. Kinki University Press, Osaka, Japan (available from Medical Physics Publishing, Madison, Wisconsin).

Kato H, Schull W. 1982. Studies of the mortality of A-bomb survivors. Report 7 Part 8, Cancer mortality among atomic bomb survivors, 1950-78. Radiat Res 90;395-432.

Mettler F, Moseley R. 1985. Medical effects of ionizing radiation. New York: Grune and Stratton.

NCRP. 1971. Basic radiation protection criteria. National Council on Radiation Protection and Measurements. Report No. 39. Washington, DC.

NCRP. 1985. A handbook of radioactivity measurements procedures. 2nd ed. National Council on Radiation Protection and Measurements. Report No. 58. Bethesda, MD:

NCRP. 1993. Risk estimates for radiation protection. National Council on Radiation Protection and Measurements. Report 115. Bethesda, Maryland

Otake M, Schull W. 1984. Mental retardation in children exposed in utero to the atomic bombs: A reassessment. Technical Report RERF TR 1-83, Radiation Effects Research Foundation, Japan.

Rubin P, Casarett G. 1968. Clinical radiation pathology. Philadelphia: W.B. Sanders Company, 33.

UNSCEAR. 1977. United Nations Scientific Committee on the Effects of Atomic Radiation. Sources and effects of ionizing radiation. New York: United Nations.

UNSCEAR. 1982. United Nations Scientific Committee on the Effects of Atomic Radiation. Ionizing radiation: Sources and biological effects. New York: United Nations.

UNSCEAR. 1986. United Nations Scientific Committee on the Effects of Atomic Radiation. Genetic and somatic effects of ionizing radiation. New York: United Nations.

UNSCEAR. 1988. United Nations Scientific Committee on the Effects of Atomic Radiation. Sources, effects and risks of ionization radiation. New York: United Nations.

UNSCEAR. 1993. United Nations Scientific Committee on the Effects of Atomic Radiation. Sources and effects of ionizing radiation. New York: United Nations.

# COBALT D-15 APPENDIX D

USNRC. 1999. Standards for the protection against radiation, table 1004(b).1. 10 CFR 20.1004. U.S. Nuclear Regulatory Commission, Washington, D.C.